

times higher than in those from patients with no history of treatment. However, the prevalence of monoresistant strains was low (5.3%, 4.3%, 0.3%, and 4.3% for isoniazid, rifampicin, ethambutol, and streptomycin, respectively) compared with the prevalence of multidrug-resistant strains whose rate reached a peak of 30.4%.

Drug-resistant TB in countries with good national control programs, such as in Western Europe, is not commonly a major health problem, although increasing immigration prompts public health authorities to maintain vigilant surveillance systems. The results of our study indicate that throughout Italy, prevalence of resistance to firstline drugs and multidrug resistance among isolates from new cases was consistently low over the 4-year survey period. Prevalence of multidrug resistance among isolates from previously treated patients was high, although a downward trend could be demonstrated during the last 2 years. Since almost 2 out of 10 isolates resistant to rifampicin were multidrug resistant, using rapid molecular methods to identify rifampicin resistance in questionable cases appears cost-effective to facilitate early detection and control of multidrug-resistant TB (10). Resistance to isoniazid is associated with immigration from countries where isoniazid was used extensively in the past. This information is a useful tool for clinicians, as isoniazid resistance may be suspected early in the disease and properly treated. Finally, the finding of substantial multidrug resistance among isolates from previously treated patients, combined with the evidence that immigrants from areas where isoniazid resistance is endemic contribute substantially to the number of new TB cases in Italy every year, strongly suggests that public health action is needed to improve treatment outcomes.

This work was funded independently by the Istituto Superiore di Sanità-Rome (National TB Project) and the World Health Organization. It was also supported by a grant (TBC1) from the Associazione Italiana Pneumologi Ospedalieri (AIPO).

**Giovanni B. Migliori,\***

**Rosella Centis,\***

**Lanfranco Fattorini,\***

**Giorgio Besozzi,\* Cesare Saltini,\*  
Claudio Scarparo,\* Daniela Cirillo,\*  
Andrea Gori,\* Antonio Cassone,\*  
and Claudio Piersimoni\***

\*SMIRA (Italian Multicentre Study on Resistance to Antituberculosis Drugs) Coordinating Committee

#### References

1. Kochi A, Vareldzis B, Styblo K. Multidrug-resistant tuberculosis and its control. *Res Microbiol* 1993;144:104–10.
2. Anti-tuberculosis drug resistance in the world. The WHO/IUATLD Global Project on anti-tuberculosis drug resistance surveillance. World Health Organization, Geneva, 1997. WHO/TB/97.229:1–227.
3. WHO/IUATLD Global Working Group on antituberculosis drug resistant surveillance. Guidelines for surveillance of drug resistance in tuberculosis. World Health Organization, Geneva, 1997. WHO/TB/96.216:1–36.
4. Centis R, Ianni A, Migliori GB, on behalf of the Tuberculosis section of the National AIPO Study Group and the SMIRA Group. Evaluation of tuberculosis treatment results in Italy, report 1998. *Monaldi Arch Chest Dis* 2000;55:293–8.
5. Migliori GB, Ambrosetti M, Fattorini L, Penati V, Vaccarino P, Besozzi G, et al. Surveillance of anti-tuberculosis drug resistance: results of the 1998/1999 proficiency testing in Italy. *Int J Tuberc Lung Dis* 2000;4: 940–6.
6. Canetti G. Quelques imprécisions dans les méthodes couramment employées pour la détermination de l'isoniazido-résistance du bacille tuberculeux, leur ampleur et leur inconvénients. *Bull Int Union Tuberc* 1955;25:157–78.
7. Roberts GD, Goodman NL, Heifets L, Larsh HW, Lindner TH, McClatchy JK, et al. Evaluation of the BACTEC radiometric method for recovery of mycobacteria and drug susceptibility testing of *Mycobacterium tuberculosis* from acid-fast smear-positive specimens. *J Clin Microbiol* 1983;18:689–96.
8. Rusch-Gerdes S, Domehl C, Nardi G, Gismondo MR, Welscher HM, Pfyffer GE. Multicenter evaluation of the mycobacteria growth indicator tube for testing susceptibility of *Mycobacterium tuberculosis* to first-line drugs. *J Clin Microbiol* 1999;37:45–8.
9. Laszlo A, Rahman M, Raviglione MC, Bustreo F, and the WHO/IUATLD Network of Supranational Reference Laboratories. Quality assurance programme for drug susceptibility testing of *Mycobacterium tuberculosis* in the WHO/IUATLD Supranational Laboratory Network: first round of proficiency testing. *Int J Tuberc Lung Dis* 1997;1:231–8.
10. Garcia de Viedma D. Rapid detection of resistance in *Mycobacterium tuberculosis*: a review discussing molecular approach. *Clin Microbiol Infect* 2003;9:349–59.

Address for correspondence: Claudio Piersimoni, Department of Clinical Microbiology, General Hospital "Umberto I," Via Conca, I-60020, Ancona, Italy; fax: 39-071-596-4184; email: piersim@tin.it

## Mollaret-like Cells in Patients with West Nile Virus Infection

**To the Editor:** We have read with interest many of the articles concerning West Nile virus (WNV) published in the July 2003 issue of *Emerging Infectious Diseases*. Last summer Ohio was one of the leading states with WNV infection in humans. Consequently, requests for tests for this pathogen have increased. Unfortunately, the turnaround time for testing these specimens may be delayed because of shipping difficulties, the limited number of laboratories that can perform these assays, and an increase in requests at testing facilities.

Cytologic examination of cerebrospinal fluid (CSF) from patients with WNV has not been studied.

Although cytologic examination of CSF from patients with encephalitis is likely nonspecific, it may provide supportive information of the suspected disease process, and is useful for excluding other conditions, such as neoplasia. Of the 22 patients that were hospitalized at our institution last year with WNV meningoencephalitis, documented by serologic tests and/or reverse transcription-polymerase chain reaction, CSF of 4 of these patients was submitted for cytologic examination. Of these 4, 3 had a sufficient number of cells in the CSF specimen (47, 213, and 495 cell/ $\mu$ L) to afford cytologic examination, whereas one had a paucicellular CSF, with only 2 white blood cells/ $\mu$ L. The cytologic features from the 3 patients,  $>10$  cells/ $\mu$ L consistently demonstrated a mixture of lymphocytes at various stages of activation and occasional large monocytic-like cells with cerebriform nuclei reminiscent of the Mollaret cells described in CSF of patients with recurrent meningitis (Figure).

Mollaret described cells with enlarged nuclei and cerebriform nuclear contours in CSF of patients with recurrent, aseptic meningitis (1).

Although he believed these were of endothelial origin, immunohistochemical studies have subsequently shown that they are monocytes (2). This type of meningitis, now commonly known as Mollaret meningitis, has been associated with herpes simplex virus encephalitis, but the definitive cause of all cases remains unclear (3).

One of the patients infected with WNV meningoencephalitis who had Mollaret-like cells in CSF died. Postmortem neuropathologic examination showed an extensive perivascular lymphocytic infiltrate which contained mononuclear cells consistent with the Mollaret-like cells in CSF. These mononuclear cells were stained with an immunohistochemical stain directed against the CD68 antigen, which supports a monocytic origin (4). Further studies are needed to delineate the consistency of Mollaret-like cells in CSF of patients with WNV meningoencephalitis. Finding Mollaret-like cells admixed with activated lymphocytes may be a useful, readily-available test that provides supportive evidence of viral encephalitis in the appropriate clinical setting, until more definitive tests are available.

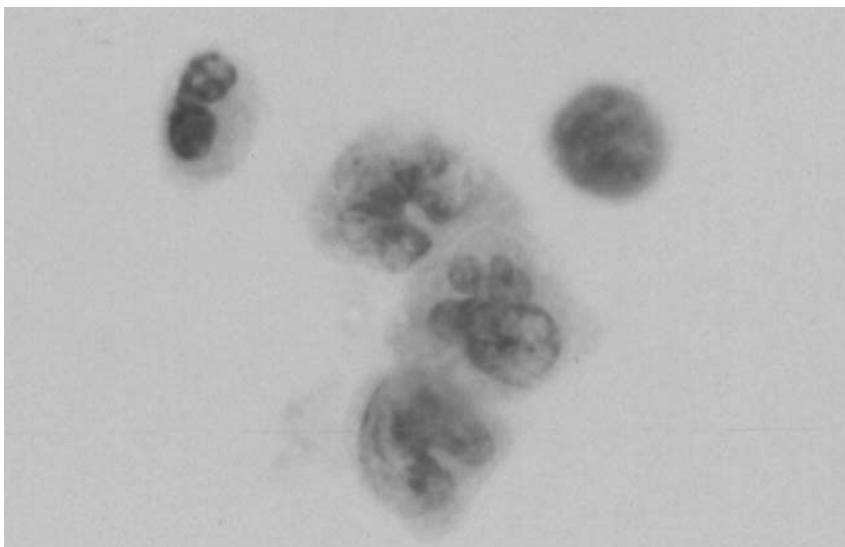


Figure. Three Mollaret-like cells are present (center), with a neutrophil (upper left) and a lymphocyte (upper right) in cerebrospinal fluid from a patient with West Nile virus encephalitis, confirmed by reverse transcription-polymerase chain reaction and serologic testing (Papanicolaou stain; magnification  $\times 500$ ).

Gary W. Procop,\*  
Belinda Yen-Lieberman,\*  
Richard A. Prayson,\*  
and Steve M. Gordon\*

\*The Cleveland Clinic Foundation,  
Cleveland, Ohio

#### References

1. Mollaret MP. La meningite endothelio-leucocitaire multirecurrente benigne: Syndrome nouveau ou maladie nouvelle? *Rev Neurol* 1981;9:81-84.
2. Stoppe G, Stark E, Patzold U. Mollaret's meningitis: CSF immunohistologic examinations. *J Neurol* 1987;234:103-6.
3. Tedder DG, Ashley R, Tyler KL, Levin MJ. Herpes simplex virus infection as a cause of benign recurrent lymphocytic meningitis. *Ann Intern Med* 1994;121:334-8.
4. Kelly TW, Prayson RA, Ruiz AI, Isada CM, Gordon SM. The neuropathology of West Nile virus meningoencephalitis. A report of two cases and review of the literature. *Am J Clin Pathol* 2003;119:749-53.

Address for correspondence: Gary W. Procop, Section Head, Clinical Microbiology, L40, the Cleveland Clinic Foundation, 9500 Euclid Avenue, Cleveland, OH 44915; fax: 216-445-6984; email: procopg@ccf.org

## Newly Isolated *Vibrio cholerae* Non-O1, Non-O139 Phages

**To the Editor:** The epidemic cholera caused by *Vibrio cholerae* O1 appeared in Latin America in 1991 after a 100-year absence. Following its explosive appearance in Peru, travelers on the Amazon River brought cholera to Brazil by April 1991. It spread southward along the Atlantic Coast of Brazil, reaching Rio de Janeiro in February 1993.

Phage typing is a useful tool for studying the source or origin of *V. cholerae* for epidemiologic importance. Because of limitations of the Basu and Mukerjee scheme, a new