

3. Miyoshi T, Uchino K, Matsuo M, Ikeda Y, Yoshida H, Sibata H, et al. Characteristics of norovirus outbreaks during a non-epidemic season. *Jpn J Infect Dis*. 2006;59:140–1.
4. Muscatello DJ, Churches T, Kaldor J, Zheng W, Chiu C, Correll P, et al. An automated, broad-based, near real-time public health surveillance system using presentations to hospital emergency departments in New South Wales, Australia. *BMC Public Health*. 2005;5:141.
5. Widdowson M-A, Cramer EH, Hadley L, Bresee JS, Beard RS, Bulens SN, et al. Outbreaks of acute gastroenteritis on cruise ships and on land: identification of a predominant circulating strain of norovirus—United States, 2002. *J Infect Dis*. 2004;190:27–36.
6. Bull RA, Tu ET, McIver CJ, Rawlinson WD, White PA. Emergence of a new norovirus genotype II.4 variant associated with global outbreaks of gastroenteritis. *J Clin Microbiol*. 2006;44:327–33.
7. White PA, Hansman GS, Li A, Dable J, Isaacs M, Ferson M, et al. Norwalk-like virus 95/96-US strain is a major cause gastroenteritis outbreaks in Australia. *J Med Virol*. 2002;68:113–8.
8. Telfer B, Munnoch S. OzFoodnet—enhancing foodborne disease surveillance across Australia. In: Annual report. Sydney (Australia): New South Wales and Hunter Area Health Service; 2005.
9. Kroneman A, Vennema H, van Duynhoven Y, Duizer E, Koopmans M. High number of norovirus outbreaks associated with GGII.4 variant in The Netherlands and elsewhere: does this herald a worldwide increase? *Euro Surveill*. 2004;8(52).
10. Bull RA, Hansman GS, Clancy LE, Tanaka MM, Rawlinson WD, White PA. Norovirus recombination in ORF1/ORF2 overlap. *Emerg Infect Dis*. 2005;11:1079–85.

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## *Echinostoma malayanum* Infection, the Philippines

**To the Editor:** In 2002, the Department of Health-Provincial Health Team (DOH-PHT) of Surigao del Norte reported 102 cases of fasciolid-like infections in the municipality of Santa Monica, Siargao Island, the Philippines. The reports were based on characteristic large operculate eggs having been found in routine stool examinations conducted during schistosomiasis surveys. *Fasciola hepatica* infection was the initial diagnosis considered. In 2005, a collaborative team from DOH-PHT, the National Institutes of Health, the University of the Philippines Manila, and the Local Government Unit/Rural Health Unit conducted a field investigation to determine the cumulative prevalence of intestinal helminthic infections in adult patients whose conditions had been previously diagnosed as fasciolid-like infection and to determine the causative species of trematode.

The study group consisted of 70 adult patients from the *barangay* (local government unit of 50–100 families) of Libertad, for whom fasciolid infections had been noted on previous surveys. Researchers confirmed infections and collected adult trematodes for species identification. All patients underwent bowel preparation with bisacodyl (Dulcolax) 10-mg tablets (2 tablets taken immediately after a meal on night before deworming), followed by praziquantel (25 mg/kg in 2 doses 4 h apart), and 30-g magnesium sulfate granules, dissolved in 1 cup of milk, given 1 h after the second dose of praziquantel. Stools were processed by using the Kato-Katz method (1) and examined microscopically by medical technologists from the Diagnostic Parasitology Laboratory, College of Public Health, University of the Philippines Manila, for intestinal helminth

ova. In addition, a clinical history was taken and a complete physical examination was conducted for each patient volunteer after stool submission. Eating preferences and habits were specifically noted.

The research was approved by the Department of Health Center for Health Development of the Caraga region. Informed consent was obtained before procedures were done and treatment was given to infected patients.

Cumulative prevalence for soil-transmitted helminth infections among the 70 patients was 51.4%. Prevalence according to species was *Trichuris* spp. 42.9%, *Ascaris* spp. 17.1%, and hookworm spp. 1.4%. *Schistosoma japonicum* infection rate was 10%. Stool samples from 8 (11.4%) patients had large (120–130  $\mu\text{m} \times 80\text{--}90 \mu\text{m}$ ), brownish, operculated eggs; 3 had a total of 13 adult flukes. Microscopy showed small leaflike flukes 8–9 mm long and 2.5–3.5 mm wide. After the organisms were processed and stained with aceto-carmin and fast green stains, diagnostic features of *Echinostoma malayanum* (Leiper 1911) were noted. Adult trematodes were within known species size range (5–10 mm  $\times$  2.5–3.0 mm) and had elongated bodies and bluntly rounded ends. The ventral sucker (acetabulum) was prominent and larger than the anterior oral sucker. Paired testes were deeply branched and positioned high in the posterior half of the body, extending above the midplane with a single anterior globular ovary. The uterus was entirely anterior to the ovary, and vitellaria (glands) were abundant along both lateral portions of the worm, ending just posterior to the esophagus. The oral sucker had a horseshoe-shaped anterior collar with 43 circumoral spines, which differentiates this species from *E. ilocanum* (49–51 collar spines), another trematode species endemic to the Philippines.

In terms of eating habits, patients reported that fish were commonly eaten raw, after being dipped in a salt

and vinegar mixture, locally known as *kinilaw*. Other methods of fish preparation were *tinola* (boiled), *ginataan* (stewed in coconut milk), and *sinugba* (charcoal-grilled). All echinostome-infected patients had a history of having eaten snails, *kuhol* and *kiambu-ay*, prepared raw with coconut milk and lime juice (*kinilaw*), especially when found in greater abundance during the rainy season.

Human echinostome infection results from ingestion of metacercariae that encyst in secondary intermediate hosts, usually freshwater snails, tadpoles, or fish. *E. malayanum* uses various species of gastropod mollusks for primary and secondary intermediate developmental stages (2–5). Certain species of fish may also serve as secondary intermediate hosts (2). Several mollusks that may serve as primary and secondary intermediate hosts have been identified in the Philippines, including *Lymnaea (Bullastra) cumingiana*, *Radix quadrasi*, and *Physastra hungerfordiana* for *E. malayanum*, and *Pila luzonica* for *E. ilocanum* (6,7).

To our knowledge, this is the first report of *E. malayanum* infections in the southern Philippines. Local eating habits are a strong factor in echinostome infections. The general lack of awareness by health staff and the community was a big factor in the poor identification of the disease. Clinical and laboratory staff and healthcare providers need training about echinostome infections and other intestinal foodborne trematode infections. Similar environmental, sanitary, and eating practices in the region suggest that the same parasitoses should be considered to be widespread in the area. Redirecting vital resources of the local health and government units of the Caraga region to the periphery and building local capacity will help empower authorities to provide public health services in rural areas, strengthen public health programs, and further develop public health infrastructure. More-

over, a successful control program against chronic intestinal parasitoses could serve as a paradigm for local health system development of effective control measures for other endemic diseases.

### Acknowledgments

We express sincere gratitude for the support, cooperation, and assistance provided by the Local Government Unit/Rural Health Unit of Santa Monica, Surigao del Norte, Provincial Health Office, DOH-PHT Surigao del Norte, V.L. Makabali Memorial General Hospital, and the generous residents of Santa Monica.

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### References

1. World Health Organization. Bench aids for the diagnosis of intestinal parasites. Geneva: the Organization; 1994. Plate 3.
2. Monzon RB, Kitikoon V, Thammapalerd N, Temcharoen P, Sornmani S, Viyanant V. Ecological observations on *Lymnaea (Bullastra) cumingiana*. Southeast Asian J Trop Med Public Health. 1993;24:563–9.
3. Garrison PE. A new intestinal trematode of man. Philippine Journal of Science. 1908;B3:385–93.
4. Waikagul J. Intestinal fluke infections in Southeast Asia. Southeast Asian J Trop Med Public Health. 1991;22(Suppl): 158–62.
5. Radomyos P, Radomyos B, Tangtrongchitr A. Multi-infection with helminths in adults from northeast Thailand as determined by post-treatment fecal examination of adult worms. Trop Med Parasitol. 1994;45: 133–5.
6. Radomyos B, Wongsaroj T, Wilairatana P, Radomyos P, Praevanich R, Meesomboon V, et al. Opisthorchiasis and intestinal fluke infections in northern Thailand. Southeast Asian J Trop Med Public Health. 1998;29:123–7.
7. Belizario VY, de Leon WU, Bersabe MJ, Baird JK, Bangs MJ. A focus of human infection by *Haplorchis taichui* (Trematoda: Heterophyidae) in the southern Philippines. J Parasitol. 2004;90:1165–9.

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## Zoonotic Pathogens in *Ixodes scapularis*, Michigan

**To the Editor:** *Ixodes scapularis*, the black-legged tick, is the predominant vector of reportable human vectorborne disease in the United States. It transmits agents that cause Lyme borreliosis, human anaplasmosis, and human babesiosis. *I. scapularis*-borne disease is becoming more frequent as this tick expands its range from tick-endemic foci in the northeastern and upper midwestern United States.

Despite Michigan's proximity to large tick-endemic areas (Wisconsin and Minnesota to the west and Indiana to the south), active and passive surveillance data indicated that the only populations of *I. scapularis* established in the state before 2002 were in Menominee County in the Upper Peninsula (1,2). However, wildlife sampling and tick dragging in 2002–2003 suggested that *I. scapularis* had begun to invade southwestern Michigan (3), with nearby populations in northwestern Indiana (4) as the putative source.