

## References

1. Allander T, Tammi MT, Eriksson M, Bjerckner A, Tiveljung-Lindell A, Andersson B. Cloning of a human parvovirus by molecular screening of respiratory tract samples. *Proc Natl Acad Sci U S A*. 2005;102:12891–6.
2. McIntosh K. Human bocavirus: developing evidence for pathogenicity. *J Infect Dis*. 2006;194:1197–9.
3. Ma X, Endo R, Ishiguro N, Ebihara T, Ishiko H, Ariga T, et al. Detection of human bocavirus in Japanese children with lower respiratory tract infections. *J Clin Microbiol*. 2006;44:1132–4.
4. Sloots TP, McErlean P, Speicher DJ, Arden KE, Nissen MD, Mackay IM. Evidence of human coronavirus HKU1 and human bocavirus in Australian children. *J Clin Virol*. 2006;35:99–102.
5. Smuts H, Hardie D. Human bocavirus in hospitalized children, South Africa. *Emerg Infect Dis*. 2006;12:1457–8.
6. Weissbrich B, Neske F, Schubert J, Tollmann F, Blath K, Blessing K, et al. Frequent detection of bocavirus DNA in German children with respiratory tract infections. *BMC Infect Dis*. 2006;6:109.
7. Schenk T, Huck B, Forster J, Berner R, Neumann-Haefelin D, Falcone V. Human bocavirus DNA detected by quantitative real-time PCR in two children hospitalized for lower respiratory tract infection. *Eur J Clin Microbiol Infect Dis*. 2007;26:147–9.
8. Kesebir D, Vazquez M, Weibel C, Shapiro ED, Ferguson D, Landry ML, et al. Human bocavirus infection in young children in the United States: molecular epidemiological profile and clinical characteristics of a newly emerging respiratory virus. *J Infect Dis*. 2006;194:1276–82.
9. Manning A, Russell V, Eastick K, Leadbetter GH, Hallam N, Templeton K, et al. Epidemiological profile and clinical associations of human bocavirus and other human parvoviruses. *J Infect Dis*. 2006;194:1283–90.
10. Arnold JC, Singh KK, Spector SA, Sawyer MH. Human bocavirus: prevalence and clinical spectrum at a children's hospital. *Clin Infect Dis*. 2006;43:283–8.

Address for correspondence: Valeria Falcone, Department of Virology, Freiburg University Medical Center, Hermann-Herder-Strasse 11, 79104 Freiburg, Germany; email: valeria.kapper-falcone@uniklinik-freiburg.de

The opinions expressed by authors contributing to this journal do not necessarily reflect the opinions of the Centers for Disease Control and Prevention or the institutions with which the authors are affiliated.

## Pure Red Blood Cell Aplasia and Isoniazid Use

**To the Editor:** Isoniazid is a first-line drug in the treatment of tuberculosis; its uses include prevention as well as cure. Isoniazid is usually well tolerated, although its common side effects include gastrointestinal discomfort, rash, allergy, hepatitis, and peripheral neuropathy. Hematologic disorders such as eosinophilia, thrombocytopenia, and autoimmune hemolytic anemia are rarely reported (1). Pure red cell aplasia (PRCA) is an uncommon disorder in adults. PRCA occurs secondary to drug exposure in 5% of patients;  $\approx 30$  drugs have been implicated (2), and few reports involving isoniazid have been published. We report a case of isoniazid-induced PRCA.

In 2006, a 79-year-old woman sought care at a public assistance hospital in Paris; she reported having had asthenia, breathlessness, and decreased appetite for 2 weeks. She had had node-negative, localized gastric adenocarcinoma 2 years earlier, which had been treated by partial gastrectomy, and pulmonary tuberculosis in 1948, which had been treated by partial pneumonectomy. As a result of a pleuropulmonary tuberculosis relapse diagnosed 6 weeks earlier, she was receiving antituberculous treatment with rifampin (400 mg/day), isoniazid (150 mg/day), ethambutol (800 mg/day), and pyrazinamide (1,000 mg/day). She received no other concomitant medications.

Her initial physical examination showed nothing abnormal. Laboratory analysis showed hemoglobin 6.3 g/dL, leukocyte and thrombocyte counts within normal limits, 1% reticulocytes, and zero schizocytes. Other test results (liver and renal function, serum folate and vitamin B12 levels, lactic dehydrogenase levels, C-reactive protein, serum protein electrophoresis, direct and indirect Coombs tests, and antinu-

clear antibody tests) were within normal limits, as were viral serologic test results (HIV, hepatitis B virus, hepatitis C virus, parvovirus B19). Upper and lower digestive tract endoscopic examination and carcinoembryonic antigen showed no abnormalities, which lessened the likelihood of a tumor relapse. Bone marrow aspiration from the sternum showed a hypocellular marrow with complete absence of the erythroid series, a normal myeloid series, and megakaryocytes. The marrow findings were consistent with PRCA.

In view of previous reports of isoniazid-induced PRCA (3–5), we suspected this drug to be responsible in this case. Isoniazid was withdrawn, and other antituberculous drugs were continued. The patient's hemoglobin level rose to 10.6 g/dL and reticulocyte count to 3% over 10 days; no blood transfusion was required.

Drug-induced PRCA is a rare blood disorder in adults and has already been reported in isoniazid-treated patients (3–5). For this patient, other causes for anemia (e.g., drug-induced hemolytic anemia, digestive malignancies, viral causes known to date, hematologic malignancies, and autoimmune disorders) were excluded (2).

The favorable outcome after isoniazid withdrawal increased the likelihood that isoniazid was the cause. Although rechallenge with isoniazid could have confirmed isoniazid as the cause, it is not an ethical option because of the hazardous adverse effects.

The exact mechanism for isoniazid-induced PRCA remains unclear, but the demonstration of antibodies reacting with nucleated red blood cells in  $\approx 50\%$  of cases suggests an induction of autoimmunity (4,5). This hypothesis is supported by previously reported cases in which PRCA relapses occurred when treatment with isoniazid was resumed (3,5). The intrinsic imputability of isoniazid also relies on the lack of a dose-effect relationship and the delay between the introduction

of isoniazid and the onset of PRCA, which can occur up to 6 months after start of treatment (3).

Clinicians treating patients with tuberculosis must be aware of this adverse reaction because failure to identify and discontinue isoniazid in patients with such a condition might lead to their illness and death. Given the ongoing worldwide HIV pandemic and the increase in tuberculosis it induces, such adverse effects are more likely to be reported in the next few years.

**Pierre Louergue,\* Olivier Mir,†  
and Robin Dhote‡**

\*Assistance Publique–Hôpitaux de Paris, Hôpital Necker, Paris, France; †Assistance Publique–Hôpitaux de Paris, Hôpital Cochin, Paris, France; and ‡Assistance Publique–Hôpitaux de Paris, Hôpital Avicenne, Bobigny, France

**References**

1. Goldman AL, Braman SS. Isoniazid: a review with emphasis on adverse effects. *Chest*. 1972;62:71–7.
2. Fisch P, Handgretinger R, Schaefer HE. Pure red cell aplasia. *Br J Haematol*. 2000;111:1010–22.
3. Goodman SB, Block MH. A case of red cell aplasia occurring as a result of anti-tuberculous therapy. *Blood*. 1964;24:616–23.
4. Sen R, Singh U, Yadav MS, Raj B, Sen J. Isoniazid induced pure red cell aplasia. *Ind J Tuberc*. 1989;36:41–3.
5. Dixit R, Dixit R, Dixit K. Isoniazid induced pure red cell aplasia. *Indian Journal of Allergy Asthma and Immunology*. 2003;17:93–5.

Address for correspondence: Olivier Mir, Service de Médecine Interne, Unité d'Oncologie Médicale, Assistance Publique–Hôpitaux de Paris, Hôpital Cochin, Université Paris 5, Faculté de Médecine, 27, Rue du Faubourg Saint Jacques, 75679 Paris, CEDEX 14, France; email: olivier.mir@cch.aphp.fr

The opinions expressed by authors contributing to this journal do not necessarily reflect the opinions of the Centers for Disease Control and Prevention or the institutions with which the authors are affiliated.

## Failure of Isoniazid Chemoprophylaxis during Infliximab Therapy

**To the Editor:** A patient with ankylosing spondylitis was treated with infliximab, a tumor necrosis factor (TNF) blocker that has been associated with reactivation of latent tuberculosis (TB). Because of reactivity in testing with purified protein derivative, isoniazid chemoprophylaxis was started 2 weeks before infliximab therapy. Four months later, a cavitary lung infection developed in the patient, caused by isoniazid-resistant *Mycobacterium kansasii*.

To our knowledge, this is the first documented case of failure of isoniazid prophylaxis due to the emergence of isoniazid-resistant mycobacteria in patients receiving infliximab therapy. TNF blockers have contributed to the control of rheumatic diseases (1). Many of the damaging inflammatory mechanisms that they inhibit are important in maintaining TB in the latent phase. Consequently, drugs that target TNF functions have been associated with an increased risk of TB (2). For these reasons, prophylactic chemotherapy should be offered to patients with latent TB (3). We show the failure of isoniazid chemoprophylaxis in a patient receiving infliximab therapy in whom lung infection developed, caused by isoniazid-resistant *M. kansasii*.

A 39-year-old man with ankylosing spondylitis was admitted to Jimenez Diaz Foundation hospital, Madrid, because of fever and lung infiltrates. He had been receiving anti-inflammatory drug therapy without amelioration of his symptoms. Therefore, treatment with infliximab was considered. Fifteen years before, the patient's father had had pulmonary TB caused by *M. tuberculosis* that was susceptible to first-line antituberculous drugs, and the patient was given chemoprophylaxis with isoniazid, 300

mg/day, during a 9-month period. Before beginning infliximab therapy, the patient was again given chemoprophylaxis with isoniazid, 300 mg/day, because a tuberculin test with 5 units of purified protein derivative showed an induration of 18 mm at 72 hours. Results of chest radiographs were normal, and cultures for mycobacteria were negative. Results of HIV testing were also negative.

After 4 months of infliximab therapy, fever, cough, and sputum production developed. New radiographs showed bilateral upper lung field infiltrates with cavitary lesions. Three acid-fast stains of sputum were positive, and treatment with rifampin, isoniazid, pyrazinamide, and ethambutol was started.

A heavy growth of photochromogenic mycobacteria was detected in 3 sputum cultures. The isolate was identified as *M. kansasii* genotype 1 by using common biochemical tests and PCR–restriction fragment length polymorphism analysis of the *hsp65* gene (4). Susceptibility tests showed resistance to isoniazid ( $\geq 5$   $\mu\text{g/mL}$ ), streptomycin, pyrazinamide, p-aminosalicylic acid, and kanamycin but susceptibility to rifampin, ethambutol, and fluoroquinolones.

Treatment was continued with a combination of rifampin, levofloxacin, and ethambutol. Sputum cultures taken after 4, 6, and 9 months of antimicrobial drug therapy were negative. After 20 months of treatment, the patient was doing well with a partial resolution of lung infiltrates, and new cultures were negative.

Isoniazid chemoprophylaxis can effectively lessen the likelihood of active TB in patients treated with TNF antagonists (5). However, at least 1 failure of TB chemoprophylaxis in a severely immunocompromised patient treated with infliximab and methotrexate has been published (6). Our patient is unique because the mycobacterial lung infection seemed to emerge as a result of the lack of activity of iso-