

Benton, Dick Braun, Rand Carpenter, Sunni Carr, John Dunn, Alice Green, Aaron Hecht, Kraig Humbaugh, Vicki Lambert, David Lincicome, Stephanie Mayfield, Susan McCool, Gary McCracken, John New, Wade Northington, Lucky Pittman, David Pelren, John Poe, Brooke Slack, Steve Thomas, Rick Toomey, Joe Turpen, Karen Waldrop, and David Withers.

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**Anne Griggs, M. Kevin Keel,  
Kevin Castle, and David Wong**

Author Affiliations: National Park Service, Mammoth Cave, Kentucky, USA (A. Griggs); Southeastern Cooperative Wildlife Disease Study, Athens, Georgia, USA (M.K. Keel); National Park Service, Ft. Collins, Colorado, USA (K. Castle); and National Park Service, Albuquerque, New Mexico, USA (D. Wong)

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**References**

1. Blehert DS, Hicks AC, Behr M, Meteyer CU, Berlowski-Zier BM, Buckles EL, et al. Bat white-nose syndrome: an emerging fungal pathogen? *Science*. 2009;323:227. <http://dx.doi.org/10.1126/science.1163874>
2. Lorch JM, Meteyer CU, Behr M, Boyles JG, Cryan P, Hicks AC, et al. Experimental infection of bats with *Geomyces destructans* causes white-nose syndrome. *Nature*. 2011;480:376–8. <http://dx.doi.org/10.1038/nature10590>
3. Chaturvedi V, Springer DJ, Behr MJ, Ramani R, Li X, Peck MK, et al. Morphological and molecular characterizations of psychrophilic fungus *Geomyces destructans* from New York bats with white-nose syndrome (WNS). *PLoS ONE*. 2010;5:e10783. <http://dx.doi.org/10.1371/journal.pone.0010783>
4. Foley J, Clifford D, Castle K, Cryan P, Ostfeld RS. Investigating and managing the rapid emergence of white-nose syndrome, a novel, fatal, infectious disease of hibernating bats. *Conserv Biol*. 2011;25:223–31.

5. Castle KT, Cryan PM. White-nose syndrome in bats: a primer for resource managers. *Park Science*. 2010;27:20–5 [cited 2012 Jan 16]. [http://www.fort.usgs.gov/Products/Publications/pub\\_abstract.asp?PubId=22941](http://www.fort.usgs.gov/Products/Publications/pub_abstract.asp?PubId=22941)
6. Toomey R, Thomas S. White-nose syndrome response plan, Mammoth Cave National Park. Mammoth Cave (KY): United States Department of the Interior, National Park Service; 2011.
7. Carr SL. White-nose syndrome confirmed in Kentucky. Frankfort (KY): Kentucky Department of Fish and Wildlife Resources; 2011 [cited 2012 Jan 16]. <http://fw.ky.gov/newsrelease.asp?nid=943>
8. Ellison LE, O'Shea TJ, Bogan MA, Everette AL, Schneider DM. Existing data on colonies of bats in the United States: summary and analysis of the U.S. Geological Survey's bat population database. Fort Collins (CO): United States Geological Survey; 2003. Information and Technology Report no. 21461 [cited 2012 Jan 16]. <http://www.fort.usgs.gov/Products/Publications/21461/21461.pdf>
9. One Health Initiative Task Force. One Health: a new professional imperative. Schaumburg (IL): American Veterinary Medical Association; 2008.
10. Higgins CL. The National Park System, a living laboratory for One Health. Florida Department of Health. One Health Newsletter. Winter 2011;4(1):7–8 [cited 2012 Jan 16]. [http://www.doh.state.fl.us/environment/medicine/One\\_Health/OHNLwinter2011.pdf](http://www.doh.state.fl.us/environment/medicine/One_Health/OHNLwinter2011.pdf)

Address for correspondence: David Wong, National Park Service, Office of Public Health, 801 Vassar Dr NE, Albuquerque, NM 87106, USA; email: [david\\_wong@nps.gov](mailto:david_wong@nps.gov)

**NDM-1-producing  
*Klebsiella  
pneumoniae*,  
Croatia**

**To the Editor:** The novel metallo-β-lactamase named New Delhi metallo-β-lactamase (NDM-1) was identified from *Klebsiella pneumoniae* and *Escherichia coli* isolates in Sweden from a patient previously

hospitalized in India (1). NDM-1 is spreading rapidly worldwide to nonclonally related isolates, many of which are directly or indirectly tracked to the Indian subcontinent (2). A carbapenem-resistant *K. pneumoniae* strain, KLZA, was isolated in May 2009 from the culture of a blood sample from of a 40-year-old man on the day after his admission to a surgical intensive care unit of the Clinical Hospital Center in Zagreb, Croatia. The patient had been transferred after 5 days of hospitalization in Bosnia and Herzegovina following a car accident. The clinical history mentioned antimicrobial drug treatment that did not include carbapenems (gentamicin, metronidazole, and ceftriaxone) and no link to the Indian subcontinent. Antimicrobial drug susceptibility testing was performed by Vitek2 (bioMérieux, Marcy-l'Etoile, France) and broth microdilution and interpreted according to the latest documents from the European Committee on Antimicrobial Susceptibility Testing ([www.eucast.org/clinical\\_breakpoints/](http://www.eucast.org/clinical_breakpoints/), version 1.1).

The strain proved resistant to imipenem and meropenem, to all broad-spectrum cephalosporins, and to aminoglycosides and susceptible to ciprofloxacin and tigecycline (Table). We checked for *bla*<sub>VIM</sub>, *bla*<sub>IMP</sub>, *bla*<sub>SPM</sub>, *bla*<sub>GIM</sub>, *bla*<sub>SIM</sub>, and *bla*<sub>NDM</sub> resistance genes by using PCR. A PCR product was obtained only with the NDM primers, after being purified (QIAquick PCR Purification Kit, QIAGEN, Hilden, Germany), its sequence showed 100% identity with *bla*<sub>NDM-1</sub>.

Strain genotyping was performed by multilocus sequence typing to determine the sequence type (ST) of the isolate and to establish a comparison with previously reported NDM-1-producing isolates. Allelic numbers were obtained on the basis of sequences of 7 housekeeping genes at [www.pasteur.fr/recherche/genopole/PF8/mlst/Kpneumoniae](http://www.pasteur.fr/recherche/genopole/PF8/mlst/Kpneumoniae).

Table. MIC of the KLZA strain of *Klebsiella pneumoniae* and its transconjugant and recipient

| Antimicrobial drug | MIC, mg/L                 |                             |                   |
|--------------------|---------------------------|-----------------------------|-------------------|
|                    | <i>K. pneumoniae</i> KLZA | <i>Escherichia coli</i> J53 | <i>E. coli</i> T1 |
| Imipenem           | 8                         | <0.06                       | 4                 |
| Meropenem          | 8                         | <0.06                       | 4                 |
| Ertapenem          | 16                        | <0.06                       | 8                 |
| Ceftazidime        | >128                      | <0.06                       | 128               |
| Cefotaxime         | >128                      | <0.06                       | 32                |
| Cefepime           | 32                        | <0.06                       | 64                |
| Aztreonam          | >128                      | 0.25                        | >128              |
| Ciprofloxacin      | 0.5                       | <0.06                       | 0.12              |
| Gentamicin         | 8                         | 0.25                        | 0.25              |
| Amikacin           | 16                        | 0.5                         | 0.5               |
| Tigecycline        | 1                         | 0.25                        | 0.25              |
| Colistin           | <0.5                      | <0.5                        | <0.5              |

html. Multilocus sequence typing identified *K. pneumoniae* KLZA as an ST25 strain, which significantly differs from the ST14 type found in the index NDM-1-producing strain and from other isolates originating from India (1) and then in other countries. ST25 *K. pneumoniae* was also found in *K. pneumoniae* isolates in Geneva (3). Other *K. pneumoniae* STs harboring NDM-1 were ST15, ST16, and ST147 (4–7).

Resistance was transferred by conjugation to *E. coli* J53, with selection based on growth on agar in the presence of ceftazidime (10 mg/L) and azide (100 mg/L). The conjugant T1 showed resistance to  $\beta$ -lactams, including all carbapenems, as well as decreased susceptibility to ciprofloxacin.

The KLZA strain and its transconjugant harbored other determinant of resistance, namely *bla*<sub>CTX-M-15</sub>, *bla*<sub>CMY-16</sub>, and *qnrA6*. Plasmid incompatibility groups, determined by a PCR-based replicon typing method, belonged to the incA/C replicon type.

This report of an NDM-1-producing *K. pneumoniae* in Croatia adds to those of other cases in patients from patients hospitalized in the Balkan area. The patient in this report had no apparent link to the Indian subcontinent.

In a survey conducted by the European Centre for Disease Prevention and Control

to gather information about the spread of NDM-1-producing *Enterobacteriaceae* in Europe and reporting cases from 13 countries during 2008–2010, five of the 55 persons with known travel histories had traveled to the Balkan region during the month before diagnosis of their infection: 2 to Kosovo and 1 each to Serbia, Montenegro, and Bosnia and Herzegovina. All had received hospital care in Balkan countries because of an illness or accident that occurred during the journey (7). Two of the latter cases (4,8) and a case from Germany (9) were subsequently published. No patient had any apparent link to the Indian subcontinent.

Although the way NDM-1 isolates might have been imported to western Europe not only from the Indian subcontinent but also from Balkan countries (10) has been highlighted, awareness of western Europe as a possible area of endemicity remains limited. The aforementioned report from Germany, although recognizing that the patient had been repatriated after hospitalization in Serbia, declared “no evidence about contact with people from regions where NDM-1-producing enterobacteria are endemic” (9). This limited awareness shows the threat of neglecting to screen patients who are transferred from countries thought not to be at risk for NDM-1. Furthermore, it means that specimen are not sent to

the local reference laboratories and recognized as positive for NDM-1, thus permitting wide dissemination of NDM-1-producing enterobacteria in the community (4). The accumulating evidence of NDM-1 from the Balkan area could suggest a possible multifocal spread of this enzyme, with the Balkans as a possible second area of endemicity, in addition to the Indian subcontinent, and prompts for widespread epidemiologic surveillance.

**Annarita Mazzariol,  
Zrinka Bošnjak, Piero Ballarini,  
Ana Budimir, Branka Bedenić,  
Smilja Kalenić,  
and Giuseppe Cornaglia**

Author affiliations: University of Verona, Verona, Italy (A. Mazzariol, P. Ballarini, G. Cornaglia); and University of Zagreb, Zagreb, Croatia (Z. Bošnjak, A. Budimir, B. Bedenić, S. Kalenić)

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## References

1. Yong D, Toleman MA, Giske CG, Cho HS, Sundman K, Lee K, et al. Characterization of a new metallo- $\beta$ -lactamase gene, *bla*<sub>NDM-1</sub>, and a novel erythromycin esterase gene carried on a unique genetic structure in *Klebsiella pneumoniae* sequence type 14 from India. *Antimicrob Agents Chemother*. 2009;53:5046–54. <http://dx.doi.org/10.1128/AAC.00774-09>
2. Rolain JM, Parola P, Cornaglia G. New Delhi metallo-beta-lactamase (NDM-1): towards a new pandemic? *Clin Microbiol Infect*. 2010;16:1699–701. <http://dx.doi.org/10.1111/j.1469-0691.2010.03385.x>
3. Poirel L, Schrenzel J, Cherkouki A, Bernabeu S, Renzi G, Nordmann P. Molecular analysis of NDM-1-producing enterobacterial isolates from Geneva, Switzerland. *J Antimicrob Chemother*. 2011;66:1730–3. <http://dx.doi.org/10.1093/jac/dkr174>
4. Bogaerts P, Bouchahrouf W, de Castro RR, Deplano A, Berhin C, Piérard D, et al. Emergence of NDM-1-producing *Enterobacteriaceae* in Belgium. *Antimicrob Agents Chemother*. 2011;55:3036–8. <http://dx.doi.org/10.1128/AAC.00049-11>
5. Mulvey MR, Grant JM, Plewes K, Roscoe D, Boyd DA. New Delhi metallo- $\beta$ -lactamase in *Klebsiella pneumoniae* and *Escherichia coli*, Canada. *Emerg Infect Dis*. 2011;17:103–6. <http://dx.doi.org/10.3201/eid1701.101358>

6. Sidjabat H, Nimmo GR, Walsh TR, Binotto E, Htin A, Hayashi Y. Carbapenem resistance in *Klebsiella pneumoniae* due to the New Delhi metallo- $\beta$ -lactamase. *Clin Infect Dis*. 2011;52:481–4. <http://dx.doi.org/10.1093/cid/ciq178>
7. Struelens MJ, Monnet DL, Magiorakos AP, O'Connor FS, Giesecke J. European NDM-1 Survey Participants. New Delhi metallo-beta-lactamase 1-producing *Enterobacteriaceae*: emergence and response in Europe. *Euro Surveill*. 2010;15:pii:19716.
8. Hammerum AM, Toleman MA, Hanse F, Kristensen B, Lester CH, Walsh TR, et al. Global spread of New Delhi metallo- $\beta$ -lactamase 1. *Lancet Infect Dis*. 2010;10:829–30. [http://dx.doi.org/10.1016/S1473-3099\(10\)70276-0](http://dx.doi.org/10.1016/S1473-3099(10)70276-0)
9. Göttig S, Pfeifer Y, Wichelhas TA, Zacharowski K, Bingold T, Averhoff B, et al. Global spread of New Delhi metallo- $\beta$ -lactamase 1. *Lancet Infect Dis*. 2010;10:828–9. [http://dx.doi.org/10.1016/S1473-3099\(10\)70275-9](http://dx.doi.org/10.1016/S1473-3099(10)70275-9)
10. Livermore DM, Walsh TR, Toleman M, Woodford N. Balkan NDM-1: escape or transplant? *Lancet Infect Dis*. 2011;11:164. [http://dx.doi.org/10.1016/S1473-3099\(11\)70048-2](http://dx.doi.org/10.1016/S1473-3099(11)70048-2)

Address for correspondence: Annarita Mazzariol, Department of Pathology and Diagnostics, University of Verona, Strada Le Grazie, 8 37134 Verona, Italy; email: annarita.mazzariol@univr.it

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## Adherence to Oseltamivir Guidelines during Influenza Pandemic, the Netherlands

**To the Editor:** In the Netherlands, the outbreak of pandemic influenza A (H1N1) 2009 led to a 100-fold increase from 2008 in prescriptions for the antiviral neuraminidase inhibitor oseltamivir (1). The guidelines for prescribing oseltamivir during the 2009 pandemic were adapted throughout the year. After August 7,

prescribers were advised to restrict prescriptions to patients with influenza symptoms plus 1 additional risk factor (2) (Table).

Community pharmacists dispensed oseltamivir as a 5-day course of sachets produced exclusively for the Dutch government program and documented all prescriptions. Our objective was to assess whether oseltamivir dispensed through community pharmacies was prescribed according to the national guideline for the pandemic virus and to investigate how patients used oseltamivir. The Institutional Review Board of the Division of Pharmacoepidemiology and Clinical Pharmacology of Utrecht University approved the study.

Pharmacists in 19 pharmacies belonging to the Utrecht Pharmacy Practice Network for Education and Research (UPPER) selected all patients who had filled a prescription for oseltamivir during May 1, 2009–February 8, 2010. These patients were contacted by phone and, after giving consent, completed a structured questionnaire. The questionnaire contained questions about potential risk factors, the reason for receiving the oseltamivir prescription (influenza symptoms or other reasons), and whether the oseltamivir course was started and completed.

Of the 630 patients eligible for contact, 361 (57.3%) completed the questionnaire. To assess whether the current guidelines were adhered to, because of the changes in policy throughout the year, we analyzed only the 300 respondents who had filled the oseltamivir prescription at the height of the pandemic, i.e., after August 7, 2009.

A total of 156 (52.0%) participants were female patients; most participants were 18–59 years of age. Of the 212 patients >18 years of age, education level was available for 195; of these, 55 (28.2%) had a low education level, 94 (48.2%) a middle education level, and 46 (23.6%) a high education level.

Of the 300 respondents, 111 (37.0%) received a prescription while they did not meet guideline criteria (Table). They had risk factors but did not experience influenza symptoms (67 [22.3%] of all respondents); had influenza symptoms but not risk factors (34 [11.3%]); or had neither influenza symptoms nor any risk factors (10 [3.3%]).

Compared with respondents who had a low education level, respondents >18 years of age who had a middle or high education level were 2× more likely to receive an oseltamivir prescription that was not in accordance with guideline criteria (odds ratio 2.20; 95% CI 1.12–4.32). Sex and age were not associated with the likelihood of receiving off-guideline oseltamivir.

Of the 189 respondents who received oseltamivir in accordance with guideline criteria, 184 (97.4%) started treatment and 167 (90.8%) completed the oseltamivir course. Of the 111 respondents who received a prescription for oseltamivir that was not in accordance with guideline criteria, 62 (55.9%) started treatment, and 56 (90.3%) completed the course.

We showed that during the pandemic the guideline criteria were not met by nearly one third of patients who received an oseltamivir prescription. Patients with a higher education level more often received a prescription, suggesting that they are more informed or empowered than patients with a lower education level to request a prescription. Another explanation for the inadequate adherence to guideline criteria is that prescribers themselves were not immediately aware of the current criteria, possibly because of changes throughout the year.

In addition, in nearly half of instances in which guideline criteria were not met but in which oseltamivir was prescribed, the patients did not start the oseltamivir course. These prescriptions could have been used for stockpiling, which