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References

1. Segura M. *Streptococcus suis*: an emerging human threat. *J Infect Dis.* 2009;199:4–6. <http://dx.doi.org/10.1086/594371>
2. Wertheim HF, Nghia H, Taylor W, Schultz C. *Streptococcus suis*: an emerging human pathogen. *Clin Infect Dis.* 2009;48:617–25. <http://dx.doi.org/10.1086/596763>
3. Yu H, Jing H, Chen Z, Zheng H, Zhu X, Wang H, et al. Human *Streptococcus suis* outbreak, Sichuan, China. *Emerg Infect Dis.* 2006;12:914–20. <http://dx.doi.org/10.3201/eid1206.051194>
4. Fittipaldi N, Collis T, Prothero B, Gottschalk M. *Streptococcus suis* meningitis, Hawaii. *Emerg Infect Dis.* 2009;15:2067–9. <http://dx.doi.org/10.3201/eid1512.090825>
5. Gottschalk M, Higgins R, Boudreau M. Use of polyvalent coagglutination reagents for serotyping of *Streptococcus suis*. *J Clin Microbiol.* 1993;31:2192–4.
6. King SJ, Leigh JA, Heath PJ, Luque I, Tarradas C, Dowson CG, et al. Development of a multilocus sequence typing scheme for the pig pathogen *Streptococcus suis*: identification of virulent clones and potential capsular serotype exchange. *J Clin Microbiol.* 2002;40:3671–80. <http://dx.doi.org/10.1128/JCM.40.10.3671-3680.2002>
7. Fittipaldi N, Xu J, Lacouture S, Tharavichitkul P, Osaki M, Sekizaki T, et al. Lineage and virulence of *Streptococcus suis* serotype 2 isolates from North America. *Emerg Infect Dis.* 2011;17:2239–44. <http://dx.doi.org/10.3201/eid1712.110609>
8. Silva LM, Baums C, Rehm T, Wisselink H, Goethe R, Valentin-Weigand P. Virulence-associated gene profiling of *Streptococcus suis* isolates by PCR. *Vet Microbiol.* 2006;115:117–27. <http://dx.doi.org/10.1016/j.vetmic.2005.12.013>
9. Versalovic J, Koeuth T, Lupski JR. Distribution of repetitive DNA sequences in eubacteria and application to fingerprinting of bacterial genomes. *Nucleic Acids Res.* 1991;19:6823–31. <http://dx.doi.org/10.1093/nar/19.24.6823>
10. Gottschalk M, Segura M, Xu J. *Streptococcus suis* infections in humans: the Chinese experience and the situation in North America. *Anim Health Res Rev.* 2007;8:29–45. <http://dx.doi.org/10.1017/S1466252307001247>

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Streptococcus suis and Porcine Reproductive and Respiratory Syndrome, Vietnam

To the Editor: *Streptococcus suis*, an opportunistic pathogen of swine, is an emerging zoonotic pathogen among humans (1). In Vietnam, *S. suis* is the leading cause of human acute bacterial meningitis (2). Infection in humans is associated with direct exposure to infected pigs or infected raw or undercooked pork products (3). Of the 35 *S. suis* serotypes, only a limited number are pathogenic for pigs, and clinical cases in humans have most frequently been attributed to serotype 2 (SS2) (1). In Vietnam during September 2006–November 2007, the carrier rate of *S. suis* among slaughterhouse pigs was 41% (222/542); SS2 was the most frequently identified serotype in 14% (45/317) of *S. suis* isolations (4).

Porcine respiratory and reproductive syndrome (PRRS) is a major disease affecting the swine industry globally; the severity of PRRS in pigs can be increased by co-infection with *S. suis* (5). In 2010, PRRS outbreaks in swine were reported in 49 of 63 Vietnamese provinces (online Technical Appendix Figure, wwwnc.cdc.gov/EID/article/19/2/12-0470-Techapp1.pdf) (6). To understand the potential implications of swine PRRS out-

breaks for human *S. suis* disease, we investigated co-infections of *S. suis* and PRRS virus (PRRSV) in sick pigs in 3 provinces of Vietnam during the PRRS outbreaks in 2010 (online Technical Appendix Figure).

We sampled 108 farms reporting pigs that had a clinical syndrome consistent with PRRSV infections in the provinces of Thai Binh (May), Tien Giang (July), and Soc Trang (July). Samples were blood from sick febrile pigs and postmortem tissue from freshly culled pigs. To confirm swine PRRS outbreaks, we performed reverse transcription real-time PCR on 1 randomly selected plasma sample from each farm (7). A total of 103 (95%) plasma samples from 103 farms tested positive for PRRSV (Chinese genotype). We additionally selected 3 PRRSV-positive farms per province for comprehensive PRRSV screening of all 42 sampled pigs; 100% of samples from the 9 farms were PRRSV positive. After swine outbreaks ended, blood samples from 52 healthy pigs from 10 farms that had no recent history of PRRS were collected from Tien Giang Province (March 2011). None of the 52 plasma samples from the 10 control farms tested positive for PRRSV.

We investigated the presence of SS2 in blood and tissue samples from pigs on PRRS- and non-PRRS-affected farms by bacterial culture (online Technical Appendix Table). A total of 534 specimens from sick pigs yielded 9 (1.7%) SS2 isolates. One (2%) of 52 specimens from the healthy control pigs yielded a non-SS2 *S. suis* isolate. *S. suis* has been proposed to contribute to the spread of antimicrobial resistance genes to other human pathogenic streptococci (8). The antimicrobial susceptibility results of 9 SS2 isolates by disk diffusion (9) revealed a high prevalence (6/9, 66%) of resistance to tetracycline, tobramycin, enrofloxacin, and either marbofloxacin or chloramphenicol.

PCR amplification of the *16SrD-NA* gene (10) and the *cps2J* gene (2)

was performed on all blood samples to detect *S. suis* and SS2, respectively. Ninety-two (18%) of 521 sick pigs from PRRSV outbreak farms were systemically infected with *S. suis*. In contrast, no healthy pigs from control farms were positive for *S. suis* by PCR (online Technical Appendix Table). The SS2-*cps2J*-specific PCR was positive for 58 (11%) of 521 samples, and the *S. suis*-*16SrDNA* PCR was positive for 55 (11%). Twenty-one of the *16SrDNA*-positive samples also were positive for *cps2J*-PCR, which indicated that 34 (7%) sick pigs were infected with non-SS2 strains. Therefore, SS2 accounted for most (58 [63%] of 92) *S. suis*-positive detections. The bacterial load of SS2 in blood ranged from 1×10^3 CFU/mL⁻¹ to 8.3×10^6 CFU/mL⁻¹ (median 9.2×10^3 CFU/mL⁻¹). Overall, SS2 was found in 58 (11%) sick pigs and on 33 (32%) PRRS outbreak farms. The higher prevalence (92 [18%]) of systemic infections of *S. suis* and SS2 with high bacterial load in pigs from PRRS outbreak farms compared with prevalence on nonoutbreak farms (1 [2%] of 52) suggests increased systemic *S. suis* infections during swine

PRRS outbreaks ($p = 0.001$, Fisher exact test).

We investigated the possible association between swine PRRS outbreaks and human *S. suis* infection. Case reports of confirmed human infections during 2007–2010 at the 2 tertiary referral hospitals in Hanoi and Ho Chi Minh City were reviewed. The number of human *S. suis* infection cases increased in August 2010 in southern Vietnam and doubled in northern Vietnam during May–August and October–November 2010 (Figure). Swine PRRS outbreaks were reported during June–September and March–December 2010 in southern and northern provinces, respectively (6) (online Technical Appendix Figure). Most patients with *S. suis* infection during these periods resided in provinces reporting swine PRRS outbreaks. Our data suggest a possible temporal association between swine PRRS outbreaks and human *S. suis* infections.

We demonstrated increased prevalence of systemic *S. suis* and SS2 infection in pigs co-infected with PRRSV during the 2010 swine outbreaks in Vietnam. The results indicate an

increased risk for potential zoonotic transmission of *S. suis* to humans during outbreaks of PRRS in swine.

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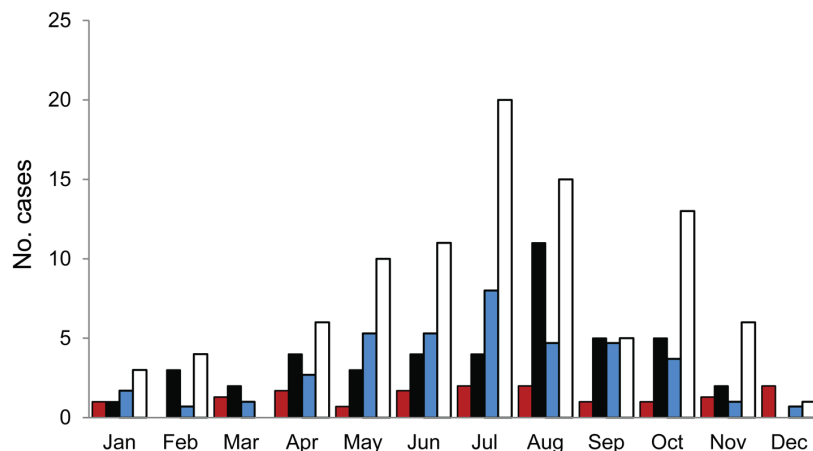


Figure. Monthly distribution of human *Streptococcus suis* infections in 2 referral hospitals, Vietnam, 2007–2010. Humans infected with *S. suis* during 2007–2009 are presented as mean total cases per month. Dark gray and black bars represent the number of *S. suis* case-patients at the Hospital for Tropical Diseases in Ho Chi Minh City during 2007–2009 and 2010, respectively. Light gray and white bars represent human *S. suis* cases at the National Hospital for Tropical Diseases in Hanoi during 2007–2009 and 2010, respectively.

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References

- Gottschalk M, Xu J, Calzas C, Segura M. *Streptococcus suis*: a new emerging or an old neglected zoonotic pathogen? *Future Microbiol.* 2010;5:371–91. <http://dx.doi.org/10.2217/fmb.10.2>
- Mai NT, Hoa NT, Nga TV, Linh le D, Chau TT, Sinh DX, et al. *Streptococcus suis* meningitis in adults in Vietnam. *Clin Infect Dis.* 2008;46:659–67. <http://dx.doi.org/10.1086/527385>
- Nghia HD, Tu le TP, Wolbers M, Thai CQ, Hoang NV, Nga TV, et al. Risk factors of *Streptococcus suis* infection in Vietnam. A case–control study *PLoS One.* 2011;6:e17604. [PubMed http://dx.doi.org/10.1371/journal.pone.0017604](http://dx.doi.org/10.1371/journal.pone.0017604)
- Ngo TH, Tran TB, Tran TT, Nguyen VD, Campbell J, Pham HA, et al. Slaughterhouse pigs are a major reservoir of *Streptococcus suis* serotype 2 capable of causing human infection in southern Vietnam. *PLoS One.* 2011;6:e17943. <http://dx.doi.org/10.1371/journal.pone.0017943>
- Xu M, Wang S, Li L, Lei L, Liu Y, Shi W, et al. Secondary infection with *Streptococcus suis* serotype 7 increases the virulence of highly pathogenic porcine reproductive and respiratory syndrome virus in pigs. *Virol J.* 2010;7:184. <http://dx.doi.org/10.1186/1743-422X-7-184>
- Tien NN. Situation of blue ear pig diseases in Vietnam and the outbreak control activities [in Vietnamese]. *Veterinary Sciences and Techniques.* 2011;18:12–20.
- Xiao XL, Wu H, Yu YG, Cheng BZ, Yang XQ, Chen G, et al. Rapid detection of a highly virulent Chinese-type isolate of porcine reproductive and respiratory syndrome virus by real-time reverse transcriptase PCR. *J Virol Methods.* 2008;149:49–55. <http://dx.doi.org/10.1016/j.jviromet.2008.01.009>
- Palmieri C, Varaldo PE, Facinelli B. *Streptococcus suis*, an emerging drug-resistant animal and human pathogen. *Front Microbiol.* 2011;2:235.
- Clinical and Laboratory Standards Institute. Performance standards for antimicrobial disk and dilution susceptibility tests for bacteria isolated from animals; approved standard. 3rd ed. (document M31–A3). Wayne (PA): The Institute; 2008.
- Marois C, Bougeard S, Gottschalk M, Kobisch M. Multiplex PCR assay for detection of *Streptococcus suis* species and serotypes 2 and 1/2 in tonsils of live and dead pigs. *J Clin Microbiol.* 2004;42:3169–75. <http://dx.doi.org/10.1128/JCM.42.7.3169-3175.2004>

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Hepatitis E Virus Seroprevalence among Men Who Have Sex with Men, United Kingdom

To the Editor: Immunosuppression might be associated with chronic carriage of hepatitis E virus (HEV) (1,2). HIV-infected persons could be at increased risk for HEV acquisition (3). If HIV infection is a risk factor for HEV, the risk will probably be mediated by associated behavioral factors. Men who have sex with men (MSM) are known to be at risk for transmission of enteric infection (4). Because of increasing prevalence of chronic liver disease induced by various causes among HIV-infected persons, it is necessary to determine whether these patients are at risk for HEV acquisition and possible hepatic decompensation (5).

We aimed to establish the contribution of HIV infection and MSM to seroprevalence of HEV among banked serum specimens. We used an unlinked, anonymous HIV seroprevalence survey of sexual health clinic attendees in England, Wales, and Northern Ireland, compared results from testing of residual serum samples collected for routine syphilis testing from sentinel clinics, and analyzed basic epidemiologic data (6). We exam-

ined serum samples collected during a 3-year period (2006–2008) and stored at -80°C . All samples were from male patients, 20–44 years of age. IgG against HEV was measured by using ELISA (Wantai; Fortress Diagnostics, Antrim, UK). To further increase the specificity for a seroprevalence analysis, and in accordance with previous work (7), we defined only samples with an optical density/cutoff value ≥ 1.5 as reactive and those in the range 1.0–1.5 as weakly reactive.

We analyzed 422 serum samples collected during 2008, comprising 146 samples from MSM with positive HIV test results, 135 from MSM with negative HIV test results, and 141 from heterosexual men with negative HIV test results. Thirty (7.1%) serum samples showed IgG reactivity against HEV and 3 (0.7%) additional samples showed weak reactivity. We examined the effect of HIV infection on prevalence of IgG against HEV by comparing samples from HIV-infected MSM with those from HIV-negative MSM. Seroprevalence rates did not differ significantly (HIV-positive MSM 7.5%; HIV-negative MSM 10.4%; $p = 0.4$).

We then examined the effect of being MSM as a risk factor for HEV infection. Prevalence of IgG against HEV among HIV-negative heterosexual men was 3.5%, significantly lower than that among MSM (odds ratio 3.1, $p = 0.025$, for comparison with non-HIV-infected MSM). We examined the relationship of status of IgG against HEV among MSM to the presence of an acute non-HIV sexually transmitted infection (STI) at the time of serum sampling. No association was found (acute STI, 14 [9.1%] of 154 vs. no acute STI, 11 [8.7%] of 127; $p = 0.9$). Similarly, no statistical association was found between HEV antibody status and the location of the clinic that provided the serum sample (London, 21 [10.0%] of 211; United Kingdom excluding London, 4 [5.7%] of 70; $p = 0.3$). As has been observed for the general UK population (7), we

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Technical Appendix

Technical Appendix Table. Systemic *Streptococcus suis* infections in sick pigs from farms with confirmed PRRS in 3 provinces, Vietnam, 2010

Province	No. PRRS outbreak farms*	No. pigs on farms	No. sick pigs reported	No. blood samples†	No. tissue sample sets‡	No. bacteria culture positive	No. <i>S. suis</i> culture positive	No. (%) <i>cps2J</i> RT-PCR (SS2) positive	No. (%) <i>16SrDNA</i> PCR (<i>S. suis</i>) positive	No. (%) non-SS2 <i>S. suis</i> positive§	Total no. (%) <i>S. suis</i> positive¶
Thai Binh	12	530	351	34	0	0	0	32 (94.1)	3 (8.8)	0	32 (94.1)
Soc Trang	41	1,055	927	232	3	9	4	11 (4.7)	15 (6.5)	9 (3.9)	20 (8.6)
Tien Giang	50	2,006	1,715	255	10	8	5	15 (5.9)	37 (14.5)	25 (9.8)	40 (15.7)
Total	103	3,591	2,993	521	13	17	9	58 (11.1)	55 (10.5)	34 (6.5)	92 (17.7)

*PRRS outbreak farms were confirmed by 1) sick pigs with a clinical syndrome consistent with PRRSV infection in farms located in provinces reported for PRRS outbreaks and 2) positive reverse transcription real-time PCR reaction to detect for the presence of *nsp2* gene from PRRS virus in 1 randomly selected blood sample/farm.

†Blood was collected from febrile pigs only. PRRS, porcine reproductive and respiratory syndrome.

‡Tissue sets comprised lung, pulmonary lymph node, liver, spleen and tonsils.

§Samples positive for 16SrDNA but negative for *cps2J*.

¶Combined *cps2J* or/and 16SrDNA positive samples.

