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## Characterization of *Mycobacterium orygis*

**To the Editor:** We thank Gey van Pittius and colleagues for their addition to the markers that identify *Mycobacterium orygis* as a distinct subspecies in the *M. tuberculosis* complex (1). Its isolation from a wild buffalo broadens the host range of *M. orygis*. Gey van Pittius and colleagues raise 3 issues: the utility of the *gyrB*<sup>oryx</sup> single-nucleotide polymorphism (SNP) being equally specific as the reported SNP in *Rv2042*<sup>38</sup>, the presence of genomic regions RD701 and RD702 in *M. orygis*, and the addition of the sequence type (ST) 701 spoligotype to *M. orygis*-specific spoligotypes.

We agree that use of the *gyrB*<sup>oryx</sup> mutation is more practical for routine daily use because this gene helps identify several subspecies of the *M. tuberculosis* complex. However, use of the partial *Rv2042* sequencing is similarly practical because it can be combined with sequencing of the adjacent *pncA* gene, which enables identification of several *M. tuberculosis* complex species and some subspecies (i.e., *M. orygis*, *M. bovis*, *M. canettii*) (2), to identify the CAS genotype of *M. tuberculosis* (J. van Ingen, unpub. data) and, to some degree, assess susceptibility to pyrazinamide (3).

With the added data, we can conclude that *M. orygis* is an *M. tuberculosis* complex subspecies defined by the presence of genomic regions RD1, RD2, RD4, RD5a, RD6, RD13–RD16, RD701, and RD702, by the C-to-G SNP in *mmpL6*<sup>551</sup>, and by the deletion of regions RD3, RD5b, RD7–RD12, RDoryx\_1, RDoryx\_4, and RDoryx\_wag22. Subspecies-specific SNPs are present in *gyrB* and *Rv2042*. Spoligotypes ST587, ST701, and closely related types are characteristic of *M. orygis*, and this subspecies yields 17–20 copies of insertion sequence *6110* and a distinct 24-locus variable number tandem repeats pattern (4,5). Given the rapid progress in genome sequencing, additional markers specific for the different subspecies will further enrich this panel of differences.

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## ***Mycobacterium tuberculosis* Beijing Type Mutation Frequency**

**To the Editor:** A striking finding in the study by de Steenwinkel et al. (1) is the high frequency of mutation to rifampin resistance by 2 *Mycobacterium tuberculosis* Beijing strains, which might play a role in the association between the Beijing strains and multidrug-resistant tuberculosis. Earlier reported frequency of mutation to rifampin resistance by *M. tuberculosis* has been  $10^{-8}$  CFU (2,3), including the Beijing genotype (3,4). Of note, the Beijing 2002–1585 strain, for which frequency of mutation to rifampin resistance is  $10^{-3}$  CFU (1 mutant/1,000 CFU), showed a moderate frequency of  $10^{-8}$  CFU in another study (4). We think that a mutation frequency increase of  $100,000\times$  is remarkably high. In contrast, rifampin-resistant mutants of the Beijing 1585 strain did not emerge in low-density cultures ( $5 \times 10^5$  CFU/mL) used for time-kill kinetics experiments, al-

though frequency of mutation to rifampin resistance was determined to be  $10^{-3}$  CFU.

Mutation frequency is determined by fluctuation assays. To exclude preexisting mutants, which would bias the mutation frequency by so-called jackpots, a series of low-inoculum cultures is typically used (5). However, for unknown reasons, de Steenwinkel et al. used only 1 high-density culture of  $10^{10}$  CFU of each strain to determine mutation frequency. This strategy is not recommended because mutations can occur early or late, resulting in substantial mutation frequency fluctuation between test episodes. A strain with known mutation rates should preferably be included to rule out possible technical errors.

We propose the following explanations for the remarkable results: 1) the rifampin concentration for selecting mutants might have been too low, enabling growth of some colonies of drug-susceptible bacteria; 2) rifampin mutants arose early or preexisted in the cultivation of Beijing strains 1585 and 1607, producing jackpots; or 3) the 2 Beijing isolates might contain rifampin-resistant subpopulations (heteroresistance). The capacity of the Beijing strain to develop and, especially, transmit multidrug-resistant tuberculosis remains to be further analyzed.

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**In Response:** We explain the differing frequencies of mutation to rifampin resistance mentioned by Werngren (1). First, the strains of *Mycobacterium tuberculosis* that we tested differed from those previously tested (2). Second, we used different rifampin concentrations in subculture plates. For Beijing strain 2002–1585, Bergval et al. (3) found a mutation frequency of  $4\text{--}24 \times 10^{-8}$  at a subculture concentration of 8 mg/L, whereas we found a mutation frequency of  $3\text{--}4 \times 10^{-3}$  at a subculture concentration of 1 mg/L and a lower mutation frequency at 2 mg/L. Thus, the concentration of drugs in subculture plates is crucial to mutation frequency assays. Absent a subculture concentration standard, we applied rifampin at 1 mg/L (4) because bacteria growing at this concentration are considered resistant to rifampin. Our mutation frequency and time-kill kinetics assay results are not contradictory