

Percentage similarity	24-loci MIRU-VNTR																				Spoligotype	Patient				
	0580	2996	0802	1955	0960	2163b	3192	1644	0424	0577	2165	4052	0154	2531	4348	2401	2059	2687	3007	4156			2347	2461	3771	3690
100%	3s	4	3	3	2	3	3	2	2	4	2	8	2	5	2	2	2	1	3	2	4	1	3	3	H3	A
	3s	4	3	3	2	3	3	2	2	4	2	8	2	5	2	2	2	1	3	2	4	1	3	3	H3	B
	3s	4	3	3	2	3	3	2	2	4	2	8	2	5	2	2	2	1	3	2	4	1	3	3	H3	C

Figure. Spoligotype and 24-loci MIRU-VNTR typing results for *Mycobacterium tuberculosis* complex isolates recovered from 3 patients with multidrug-resistant tuberculosis (TB). Patient A (index case-patient), Burma-born man with TB, incarcerated in Singapore correctional facility; patient B, Singapore-born man with HIV infection and TB, who transported prisoners in Singapore; patient C, Singapore-born man with HIV infection and TB, who shared cell with patient A. MIRU-VNTR, mycobacterial interspersed repetitive units-variable number tandem repeat.

medical teams been aware of his recent MDR TB exposure.

A recent update documented the highest rates of global MDR TB in 2009 and 2010 (9). Our experience reported here underscores the need to be constantly mindful of this infectious disease threat in our increasingly borderless world, even in countries where incidence of MDR TB is low.

**Cynthia B.E. Chee,
Li-Yang Hsu, Li-Hwei Sng,
Yee-Sin Leo, Jeffery Cutter,
and Yee-Tang Wang**

Author affiliations: Tan Tock Seng Hospital, Singapore (C.B.E. Chee, Y.-S. Leo, Y.-T. Wang); National University of Singapore, Singapore (L.-Y. Hsu); Singapore General Hospital, Singapore (L.-H. Sng); and Ministry of Health, Singapore (J. Cutter)

DOI: <http://dx.doi.org/10.3201/eid1907.120372>

References

1. Ministry of Health. Communicable disease surveillance report in Singapore 2010. Singapore: The Ministry; 2011.
2. Chee CBE, KhinMar KW, Cutter J, Wang YT. The imminent threat of multidrug-resistant tuberculosis in Singapore. *Singapore Med J* 2012;53:238–40.
3. Kyi Win KM, Chee CBE, Shen L, Wang YT, Cutter J. Tuberculosis among foreign-born persons, Singapore, 2000–2009. *Emerg Infect Dis*. 2011;17:517–9.
4. Kamerbeek J, Schouls L, Kolk A, van Agterveld M, van Soolingen D, Kuijper S, et al. Simultaneous detection and strain differentiation of *Mycobacterium tuberculosis* for diagnosis and epidemiology. *J Clin Microbiol*. 1997;35:907–14.

5. Supply P, Allix C, Lesjean S, Cardoso-Oelemann M, Rusch-Gerdes S, Willery E, et al. Proposal for standardization of optimized mycobacterial interspersed repetitive unit-variable-number tandem repeat typing of *Mycobacterium tuberculosis*. *J Clin Microbiol*. 2006;44:4498–510. <http://dx.doi.org/10.1128/JCM.01392-06>
6. Valway SE, Greifinger RB, Papania M, Kilburn JO, Woodley C, DiFerdinando GT, et al. Multidrug-resistant tuberculosis in the New York State prison system, 1990–1991. *J Infect Dis*. 1994;170:151–6. <http://dx.doi.org/10.1093/infdis/170.1.151>
7. Edlin BR, Tokars JI, Grieco MH, Crawford JT, Williams J, Sordillo EM, et al. An outbreak of multidrug-resistant tuberculosis among hospitalised patients with the acquired immunodeficiency syndrome. *N Engl J Med*. 1992;326:1514–21. <http://dx.doi.org/10.1056/NEJM199206043262302>
8. Wells CD, Cegielski JP, Nelson LJ, Laserson KF, Holtz TH, Finlay A, et al. HIV infection and multidrug-resistant tuberculosis: the perfect storm. *J Infect Dis*. 2007;196(Suppl 1):S86–107. <http://dx.doi.org/10.1086/518665>
9. Zignol M, van Gemert W, Falzon D, Sismanidis C, Glaziou P, Floyd K, et al. Surveillance of anti-tuberculosis drug resistance in the world: an updated analysis, 2007–2010. *Bull World Health Organ*. 2012;90:111–119D. <http://dx.doi.org/10.2471/BLT.11.092585>

Address for correspondence: Cynthia B. E. Chee, TB Control Unit, Department of Respiratory and Critical Care Medicine, Tan Tock Seng Hospital, 144 Moulmein Rd, Singapore 308089; email: cynthia_chee@ttsh.com.sg

All material published in *Emerging Infectious Diseases* is in the public domain and may be used and reprinted without special permission; proper citation, however, is required.

Human Infection with Marten Tapeworm

To the Editor: Cysticercosis-like human infections with the tapeworm *Taenia crassiceps*, which infects foxes as terminal hosts, have been reported (1,2). We report a case of a cysticercosis-like eye infection caused by the tapeworm *T. martis* (marten tapeworm) in a woman.

The patient was a 43-year-old German woman who sought care during July 2010, after 4 days of perceiving flashing lights in her visual field and a paracentral scotoma in her left eye. Visual acuity in both eyes was 20/20. Examination of the left fundus revealed a mobile subretinal tumor at the temporal upper retinal branch vessel with adjacent intraretinal and subhyaloid bleeding (Figure, panels A–C; online Video, wwwnc.cdc.gov/EID/article/19/7/12-1114-F1.htm). The subretinal tumor resembled a cestode larva.

The patient reported no other symptoms at that time. Laboratory evaluation found no eosinophilia or elevation of total IgE. Serologic testing results were negative for antibodies against the following parasites: *Taenia solium*, *Echinococcus multilocularis*, *E. granulosus*, *Dirofilaria immitis*, *Strongyloides* spp., and *Toxocara canis*. Fecal testing results were negative for worm eggs. Images from ultrasonography of the liver and magnetic resonance imaging of the head were unremarkable. The patient's travel history included—in addition to southern European countries—trips to Nepal and Thailand 15 years previously.

At the time of examination, the patient lived in a small village near Freiburg (im Breisgau) in southwestern Germany. She grew vegetables in the family garden, which was next to a forest. Her 3 children and husband did not report any health problems. For the past 6 years, the family had owned

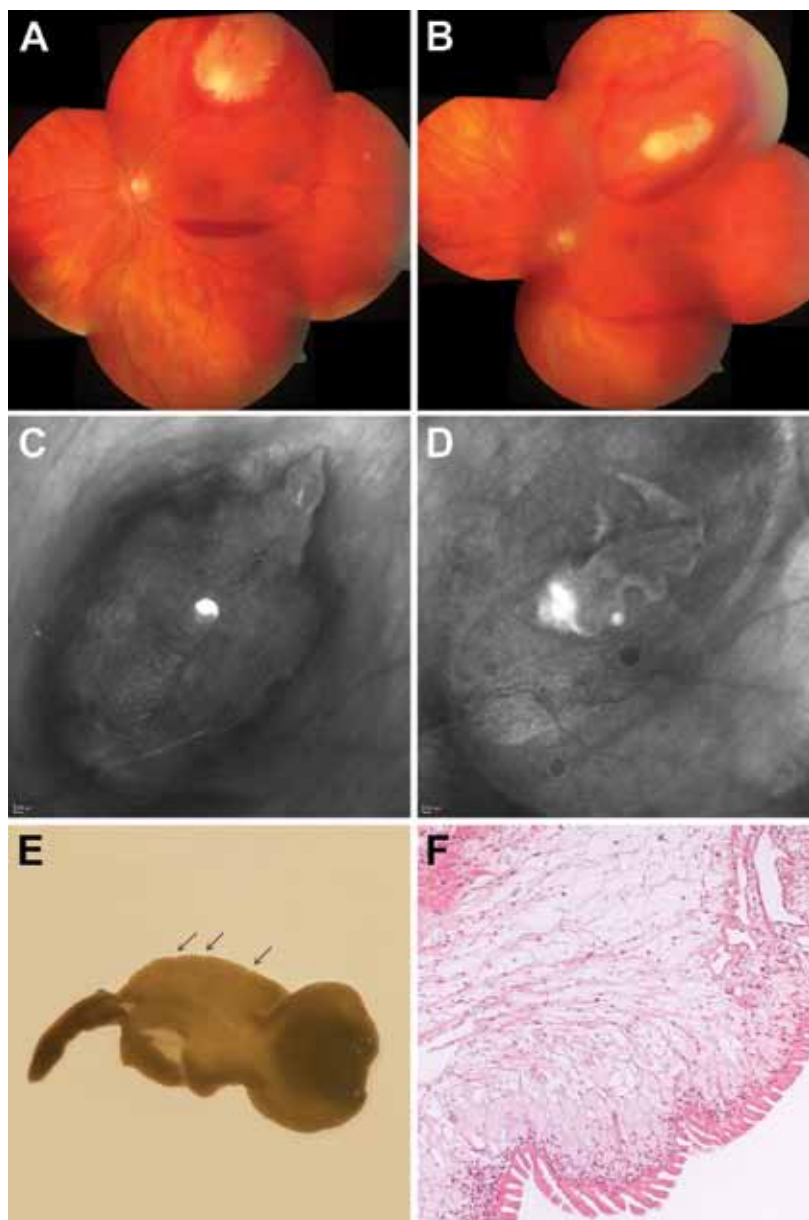


Figure. Cysticercosis-like eye infection caused by the tapeworm *Taenia martis* in a woman. A) Fundus at the patient's initial visit, before medical therapy. The cyst lies subretinally at the temporal upper branch vessels; adjacent intraretinal and subretinal bleeding and central subhyaloid bleeding can be seen. B) After 8 days of medical therapy, the cyst size had decreased markedly. The physis of the larva (A and B) is reminiscent of the armatetrathyridium (or fimbricercus), a larval form typical for the tapeworm subspecies *T. martis martis*. C) Cyst at patient's initial visit. D) Cyst at time of surgery. E) Surgically removed monocephalic cysticercus-like larva with inverted parenchymatous portion, withdrawn scolex, and attenuated posterior end. The tegumental surface is transversely striated and exhibits inward folds (arrows). F) Histologic section of the *Taenia martis* tapeworm cyst showing morphologic characteristics also commonly seen in cysticercosis cysts caused by *T. solium* tapeworms. The syncytial bladder wall consists of a rugate external, a nucleated intermediate, and an internal reticular layer with lacunate branches of the excretory duct system. Filamentous extensions of contractile muscles project into the parenchyma, which is interspersed with a few calcareous corpuscles. In addition, the *T. martis* cyst shows a preponderance of uniformly organized, elongate and slender tegumental processes, which are usually not seen in histologic sections of cyst walls caused by *T. solium* tapeworms. Hematoxylin and eosin stain; objective magnification $\times 10$.

a dog, which received antiparasitic medications on a regular basis; recent checks for intestinal parasitic infection found no ova.

The suspected cause of the woman's illness was cysticercosis caused by the larva of *T. solium*; systemic antiparasitic therapy was started (albendazole 400 mg 2 \times /d, dexamethasone 20 mg/d). The size of the larva diminished (Figure, panel D; Video), but the patient remained symptomatic. Therefore, after 8 days of therapy, the cyst was removed by retinotomy. A few days later, peripheral retinal detachment occurred and was treated by a second vitrectomy and intravitreal gas injection. Because of the repeated gas tamponade, a gas cataract developed, which necessitated cataract surgery. At the end of March 2011, the patient's visual acuity had returned to 20/20 in both eyes.

The removed cyst showed the characteristic macroscopic and histologic features of a cysticercus bladder wall (Figure, panels E, F). To determine the exact species by using molecular methods, we isolated DNA from the cyst, conducted different PCRs selective for mitochondrial genes, determined the corresponding sequences, and used a BLAST search (3) to compare these sequences with publically available sequences. Sequences of the following mitochondrial genes were determined by using the given primers and later submitted to GenBank: small ribosomal subunit (primers 12S *Taenia* FF 5'-CACAGTGCCAGCAT-CYGCGGT-3' and 12S *Taenia* RR 5'-GAGGGTGACGGGCGGTGT-GTAC-3', PCR product of 426 bp, GenBank accession no. JX415820); NADH dehydrogenase subunit 1 (primers: NAD1-FF 5'-ATTGGKT-TATTCAGAGTTTTTCTGATT-TA-3' and NAD1-RR 5'-CTCMC-CATAATCAAATGGACTACG-3', 394 bp, GenBank accession no. JX415819); and the cytochrome-*c* oxidase subunit 1 (determined by using previously published primers

[4,5]; 376 bp, GenBank accession no. JX415821). All sequences showed highest identity with *T. martis* (99%–100%) but substantially lower identity with *T. crassiceps* (91%–97%) and *T. solium* (87%–89%) tapeworms.

Thus, molecular methods unequivocally identified the larva as that of a *T. martis* tapeworm. *T. martis* tapeworms (cestodes) live and produce eggs in the intestines of definitive hosts, weasels (family *Mustelidae*), which also includes pine martens, stone martens, polecats, badgers, wolverines, and stoats (6). The intermediate hosts are prey animals of the definitive hosts, such as arvicoline (voles and muskrats) or murid rodents. When intermediate hosts ingest eggs, cysticerci develop in the pleural and peritoneal cavities. *T. martis* tapeworms probably occur worldwide wherever suitable definitive and intermediate hosts are present (6,7). A study in southwest Germany reported that 36% of stone martens were infected with *T. martis* tapeworms (6).

Although nearly all patients who had cysticercosis-like infections caused by *T. crassiceps* tapeworms were immunosuppressed (1,2), we found no signs of immunosuppression in the patient reported here. The only identified risk factor for this patient was consumption of homegrown vegetables, which could have been contaminated by marten feces.

The clinical and histologic appearance of the organism in this patient suggested cysticercosis caused by a *T. solium* tapeworm. However, the specific diagnosis of *T. martis* tapeworm infection was possible only by use of molecular methods. Thus, human infections with *T. martis* and other animal tapeworms might occur at times but might be misdiagnosed as *T. solium* cysticercosis. For therapy, the rules and considerations are probably the same as those for *T. solium* cysticercosis, as described (8,9). Concerning antiparasitic therapy, one must be aware of possible complications

caused by intraocular immunologic reactions. As demonstrated by the case reported here, surgical removal of a subretinal larva is connected with the risk for retinal detachment and cataract formation. The identification of the responsible tapeworm is useful for epidemiologic reasons, for determining the source of infection. We therefore suggest using molecular methods to determine the exact species of parasites removed from human tissue.

Acknowledgment

We gratefully acknowledge Heidrun von Thien for her technical assistance. We thank the patient for her collaboration and her approval to publish this case report, and we thank the board of directors of the Bernhard Nocht Institute for Tropical Medicine, Rolf Horstmann, Bernhard Fleischer, and Egbert Tannich for their support.

**Philipp Eberwein,
Alexandra Haeupler,
Fabian Kuepper, Dirk Wagner,
Winfried V. Kern, Birgit Muntau,
Paul Racz,
Hansjuergen Agostini,
and Sven Poppert**

Author affiliations: Albert-Ludwigs-Universität, Freiburg, Germany (P. Eberwein, F. Kuepper, D. Wagner, W.V. Kern, H. Agostini); and Bernhard Nocht Institute for Tropical Medicine, Hamburg, Germany (A. Haeupler, B. Muntau, P. Racz, S. Poppert)

DOI: <http://dx.doi.org/10.3201/eid1907.121114>

References

- Goesseringer N, Lindenblatt N, Michic-Probst D, Grimm F, Giovanoli P. *Taenia crassiceps* upper limb fasciitis in a patient with untreated acquired immunodeficiency syndrome and chronic hepatitis C infection—the role of surgical debridement. *J Plast Reconstr Aesthet Surg*. 2011;64:e174–6. <http://dx.doi.org/10.1016/j.bjps.2011.02.011>
- Heldwein K, Biedermann HG, Hamperl WD, Bretzel G, Loscher T, Laregina D, et al. Subcutaneous *Taenia crassiceps* infection in a patient with non-Hodgkin's lymphoma. *Am J Trop Med Hyg*. 2006;75:108–11.
- Altschul SF, Madden TL, Schaffer AA, Zhang J, Zhang Z, Miller W, et al. Gapped BLAST and PSI-BLAST: a new generation of protein database search programs. *Nucleic Acids Res*. 1997;25:3389–402.
- Bowles J, Blair D, McManus DP. Genetic variants within the genus *Echinococcus* identified by mitochondrial DNA sequencing. *Mol Biochem Parasitol*. 1992;54:165–73. [http://dx.doi.org/10.1016/0166-6851\(92\)90109-W](http://dx.doi.org/10.1016/0166-6851(92)90109-W)
- Bowles J, McManus DP. NADH dehydrogenase 1 gene sequences compared for species and strains of the genus *Echinococcus*. *Int J Parasitol*. 1993;23:969–72. [http://dx.doi.org/10.1016/0020-7519\(93\)90065-7](http://dx.doi.org/10.1016/0020-7519(93)90065-7)
- Loos-Frank B, Zeyhle E. The intestinal helminths of the red fox and some other carnivores in southwest Germany. *Z Parasitenkd*. 1982;67:99–113. <http://dx.doi.org/10.1007/BF00929518>
- Mathy A, Hanosset R, Adant S, Losson B. The carriage of larval *Echinococcus multilocularis* and other cestodes by the muskrat (*Ondatra zibethicus*) along the Ourthe River and its tributaries (Belgium). *J Wildl Dis*. 2009;45:279–87.
- Palomares F, Palencia G, Ambrosio JR, Ortiz A, Jung-Cook H. Evaluation of the efficacy of albendazole sulphoxide and praziquantel in combination on *Taenia crassiceps* cysts: in vitro studies. *J Antimicrob Chemother*. 2006;57:482–8. <http://dx.doi.org/10.1093/jac/dki484>
- Nash TE, Garcia HH. Diagnosis and treatment of neurocysticercosis. *Neurology reviews*. 2011;7:584–94.

Address for correspondence: Sven Poppert, Bernhard-Nocht-Institut für Tropenmedizin Bernhard-Nocht-Strasse 74, 20359 Hamburg, Germany; email: sven@poppert.eu

Letters

Letters commenting on recent articles as well as letters reporting cases, outbreaks, or original research are welcome. Letters commenting on articles should contain no more than 300 words and 5 references; they are more likely to be published if submitted within 4 weeks of the original article's publication. Letters reporting cases, outbreaks, or original research should contain no more than 800 words and 10 references. They may have 1 Figure or Table and should not be divided into sections. All letters should contain material not previously published and include a word count.