

References

- Harris JB, LaRocque RC, Qadri F, Ryan ET, Calderwood SB. Cholera. *Lancet*. 2012;379:2466–76. [http://dx.doi.org/10.1016/S0140-6736\(12\)60436-X](http://dx.doi.org/10.1016/S0140-6736(12)60436-X)
- Republique d’Haiti Ministère de la Santé Publique et de la Population. Ministère de la Santé Publique et de la Population (MSPP) rapport de cas [cited 2014 Mar 18]. http://mspp.gouv.ht/site/downloads/Rapport%20Web_18.03_Avec_Courbes_Departementales.pdf.
- Clinical and Laboratory Standards Institute. Performance standards for antimicrobial susceptibility testing; twenty-first informational supplement. CLSI Document M100–S21. Wayne (PA): The Institute; 2013.
- Sjölund-Karlsson M, Reimer A, Folster JP, Walker M, Dahourou GA, Batra DG, et al. Drug-resistance mechanisms in *Vibrio cholerae* O1 outbreak strain, Haiti, 2010. *Emerg Infect Dis*. 2011;17:2151–4.
- Steenland MW, Joseph GA, Lucien MA, Freeman N, Hast M, Nygren BL, et al. Laboratory-confirmed cholera and rotavirus among patients with acute diarrhea in four hospitals in Haiti, 2012–2013. *Am J Trop Med Hyg*. 2013;89:641–6. <http://dx.doi.org/10.4269/ajtmh.13-0307>
- Katz LS, Petkau A, Beaulaurier J, Tyler S, Antonova ES, Turnsek MA, et al. Evolutionary dynamics of *Vibrio cholerae* O1 following a single-source introduction to Haiti. *MBio*. 2013;4:e00398-13. <http://dx.doi.org/10.1128/mBio.00398-13>
- Zankari E, Hasman H, Cosentino S, Vestergaard M, Rasmussen S, Lund O, et al. Identification of acquired antimicrobial resistance genes. *J Antimicrob Chemother*. 2012;67:2640–4. <http://dx.doi.org/10.1093/jac/dks261>
- Carattoli A, Bertini A, Villa L, Falbo V, Hopkins KL, Threlfall EJ. Identification of plasmids by PCR-based replicon typing. *J Microbiol Methods*. 2005;63:219–28. <http://dx.doi.org/10.1016/j.mimet.2005.03.018>
- Young HK, Amyes SG. Plasmid trimethoprim resistance in *Vibrio cholerae*: migration of the type I dihydrofolate reductase gene out of the *Enterobacteriaceae*. *J Antimicrob Chemother*. 1986;17:697–703. <http://dx.doi.org/10.1093/jac/17.6.697>
- Call DR, Singer RS, Meng D, Broschat SL, Orfe LH, Anderson JM, et al. *bla*_{CMV-2}-positive IncA/C plasmids from *Escherichia coli* and *Salmonella enterica* are a distinct component of a larger lineage of plasmids. *Antimicrob Agents Chemother*. 2010;54:590–6. <http://dx.doi.org/10.1128/AAC.00055-09>

Address for correspondence: Jason P. Folster, Centers for Disease Control and Prevention, 1600 Clifton Rd NE, Atlanta, GA 30329-4027, USA; email: gux8@cdc.gov

Human Co-Infection with Avian and Seasonal Influenza Viruses, China

To the Editor: In April 2013, a case of co-infection with avian-origin influenza A(H7N9) virus and seasonal influenza A(H3N2) virus was reported in Jiangsu Province, China (1). This case raised concern over the possible occurrence of new reassortants with enhanced transmissibility among humans. Because of the nature of the dynamic reassortment of A(H7N9) virus with A(H9N2) virus in the environment and in poultry (2,3), close surveillance for possible new reassortment in human patients with A(H7N9) infection is needed. We report co-infection in 2 patients in Hangzhou, the capital Zhejiang Province, China, in January 2014. The co-infections involved influenza A(H7N9) virus and a seasonal A(H1N1)pdm09 virus (1 patient) or a seasonal influenza B virus (1 patient).

Of 60 patients with laboratory-confirmed influenza A(H7N9) infections in Hangzhou in April 2013 and in January–February 2014, testing of pharyngeal swab samples indicated that 2 patients were also positive for seasonal influenza virus. The pharyngeal samples were tested by real-time reverse transcription PCR according to protocols provided by the Chinese National Influenza Center. Informed consent for this study was provided by each patient’s spouse.

On January 6, 2014, patient 1 (male, 58 years of age), a resident of Xiaoshan District, had a high fever (39.6°C) and a cough; at a hospital, he received a diagnosis of severe acute interstitial pneumonia. The patient had a history of chronic myelogenous leukemia; his history of exposure to live poultry was not clear. On January 13, infection with influenza A(H7N9) virus was laboratory confirmed; viral RNA from a pharyngeal swab sample collected before oseltamivir treatment was positive for the following: influenza A virus (cycle threshold [C_t] = 26), H7 (C_t = 27), N9 (C_t = 26), influenza A(H1N1)pdm09 virus H1 (C_t = 30), and N1 (C_t = 30). The 2 viruses were named A/Hangzhou/10–1/2014(H7N9) and A/Hangzhou/10–2/2014(H1N1)pdm09. The patient received oseltamivir while in the hospital but died on January 18.

On January 5, patient 2 (male, 54 years of age), also from Xiaoshan District, had fever and a cough; at a hospital, he received a diagnosis of severe acute pneumonia. He had a history of aplastic anemia and had been exposed to live poultry 1 week before symptom onset. On January 18, infection with influenza A(H7N9) virus was laboratory confirmed. Viral RNA from a pharyngeal swab sample collected before oseltamivir treatment was positive for the following: influenza A virus (C_t = 22), H7 (C_t = 23), N9 (C_t = 22), and influenza B virus (C_t = 22). The viruses were named A/Hangzhou/17–1/2014(H7N9) and B/Hangzhou/17–2/2014. This patient received oseltamivir but died on January 22.

The hemagglutinin (HA) and neuraminidase (NA) sequences of viruses from these 2 patients were determined by Sanger sequencing. The specific primers used are listed in online Technical Appendix Table 1 (<http://wwwnc.cdc.gov/EID/article/20/11/14-0897-Techapp1.pdf>). The accession numbers of these sequences and the reference sequences for phylogenetic analyses are

listed in online Technical Appendix Table 2. Phylogenetic analyses (4) revealed that these 2 influenza A(H7N9) viruses were clustered into the clade of A/Shanghai/2/2013(H7N9)-like viruses (Figure). Some amino acid features within the HA and NA of these 2 viruses were the same as those in the A/Shanghai/2/2013(H7N9) strain: L226 and G228 in HA, believed to control host receptor specificity; the cleavage site in HA,

relevant for virulence; a deletion in NA stalk (position 69–73), associated with the adaption to gallinaceous hosts; and R294 in NA, related to virus sensitivity to oseltamivir (5). The HA and NA sequences of A/Hangzhou/10–2/2014(H1N1) pdm09 and B/Hangzhou/17–2/2014 were very close to those of A(H1N1) pdm09 virus and B/Yamagata-lineage viruses that had recently circulated in China (6,7).

Co-infection with A(H7N9) virus and seasonal influenza viruses is probably associated with the overlap of A(H7N9) virus and seasonal virus circulation in both time and space and with increased prevalence of influenza virus infections within the population. From November 2012 through March 2014, outbreaks of A(H7N9) infection (in April 2013 and in January–February 2014) were concurrent with increases in seasonal influenza

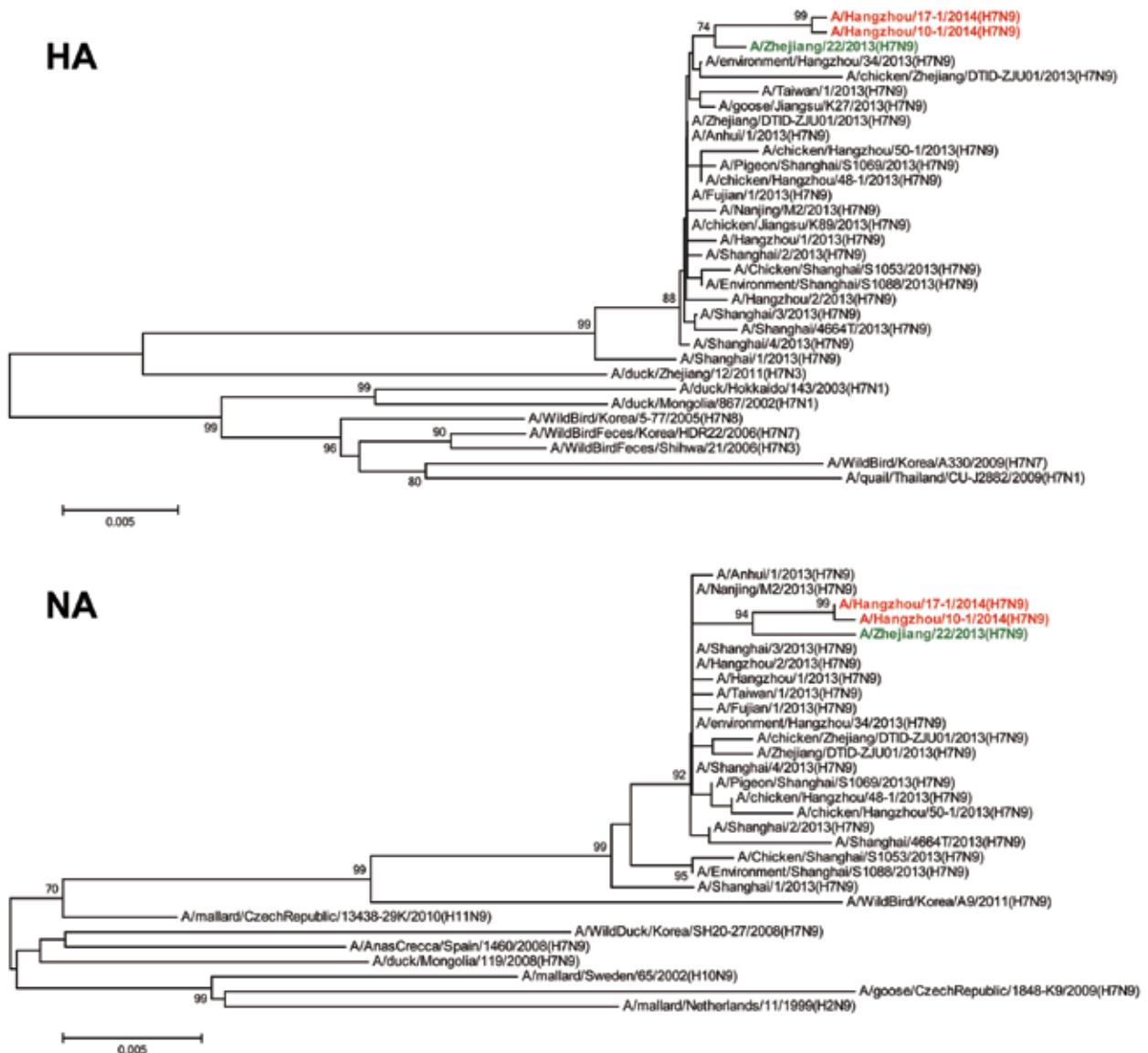


Figure. Phylogenetic analyses of hemagglutinin (A) and neuraminidase (B) of influenza A(H7N9) viruses. The trees were constructed by using the neighbor-joining method with bootstrap analysis ($n = 1,000$) in the MEGA5.0 program (4). Red indicates the 2 viruses isolated from co-infected patients in Hangzhou, China, and green indicates the first strain isolated during the second wave of the influenza A(H7N9) outbreak in China, which started in October 2013. Scale bars indicate nucleotide substitutions per site.

virus infections in Hangzhou (online Technical Appendix Figure). Prompt control of A(H7N9) infection outbreaks and vaccination against seasonal influenza viruses could reduce the potential for co-infections with A(H7N9) virus and seasonal viruses.

Taken together with the previous finding of human co-infection with A(H7N9) virus and A(H3N2) virus (1), our results show that human co-infection with A(H7N9) virus and each of the 3 seasonal influenza viruses currently circulating worldwide can occur. Avian influenza viruses, including A(H7N9), preferentially replicate in the lower respiratory tract of humans (8,9). In contrast, seasonal influenza viruses preferentially infect the upper respiratory tract of humans (10). Coexistence of A(H7N9) virus with either A(H1N1)pdm09 virus or influenza B virus in the pharyngeal swab samples from 2 patients suggests that the upper respiratory tract could provide a location for the A(H7N9) virus to reassort with other influenza viruses. The possibility that seasonal influenza viruses might provide some gene segments that increase the human-to-human transmissibility of possible new reassortants is cause for concern. For detection of such new influenza virus reassortants, extensive surveillance to identify influenza virus co-infections is necessary.

This work was supported by the Hangzhou Key Medicine Discipline Fund for Public Health Laboratory, sponsored by the Hangzhou government, and the S&T Innovation Group of Key Technology for Public Health Surveillance and Emergency Preparedness and Response, sponsored by the Zhejiang government (2011R50021).

**Jun Li, Yu Kou, Xinfen Yu,
Yongxiang Sun, Yinyan Zhou,
Xiaoying Pu, Tao Jin,
Jingcao Pan, and George F. Gao**

Author affiliations: Hangzhou Center for Disease Control and Prevention, Hangzhou, China (J. Li, Y. Kou, X. Yu, Y. Zhou,

X. Pu, J. Pan); Xiaoshan District Center for Disease Control and Prevention, Hangzhou (Y. Sun); BGI-Shenzhen, Shenzhen, China (T. Jin); and Chinese Academy of Sciences Key Laboratory of Pathogenic Microbiology and Immunology, Beijing, China (G.F. Gao)
DOI: <http://dx.doi.org/10.3201/eid2011.140897>

References

- Zhu Y, Qi X, Cui L, Zhou M, Wang H. Human co-infection with novel avian influenza A H7N9 and influenza A H3N2 viruses in Jiangsu province, China. *Lancet*. 2013;381:2134 [http://dx.doi.org/10.1016/S0140-6736\(13\)61135-6](http://dx.doi.org/10.1016/S0140-6736(13)61135-6).
- Yu X, Jin T, Cui Y, Pu X, Li J, Xu J, et al. Influenza H7N9 and H9N2 viruses: coexistence in poultry linked to human H7N9 infection and genome characteristics. *J Virol*. 2014;88:3423–31 <http://dx.doi.org/10.1128/JVI.02059-13>.
- Cui L, Liu D, Shi W, Pan J, Qi X, Li X, et al. Dynamic reassortments and genetic heterogeneity of the human-infecting influenza A (H7N9) virus. *Nat Commun*. 2014;5:3142.
- Tamura K, Peterson D, Peterson N, Stecher G, Nei M, Kumar S. MEGA5: molecular evolutionary genetics analysis using maximum likelihood, evolutionary distance, and maximum parsimony methods. *Mol Biol Evol*. 2011;28:2731–9. <http://dx.doi.org/10.1093/molbev/msr121>
- Gao R, Cao B, Hu Y, Feng Z, Wang D, Hu W, et al. Human infection with a novel avian-origin influenza A (H7N9) virus. *N Engl J Med*. 2013;368:1888–97 <http://dx.doi.org/10.1056/NEJMoa1304459>.
- Li J, Shao TJ, Yu XF, Pan JC, Pu XY, Wang HQ, et al. Molecular evolution of HA gene of the influenza A H1N1 pdm09 strain during the consecutive seasons 2009–2011 in Hangzhou, China: several immune-escape variants without positively selected sites. *J Clin Virol*. 2012;55:363–6 <http://dx.doi.org/10.1016/j.jcv.2012.08.006>.
- Tan Y, Guan W, Lam TT, Pan S, Wu S, Zhan Y, et al. Differing epidemiological dynamics of influenza B virus lineages in Guangzhou, southern China, 2009–2010. *J Virol*. 2013;87:12447–56 <http://dx.doi.org/10.1128/JVI.01039-13>.
- Chen Y, Liang W, Yang S, Wu N, Gao H, Sheng J, et al. Human infections with the emerging avian influenza A H7N9 virus from wet market poultry: clinical analysis and characterisation of viral genome. *Lancet*. 2013;381:1916–25 [http://dx.doi.org/10.1016/S0140-6736\(13\)60903-4](http://dx.doi.org/10.1016/S0140-6736(13)60903-4).
- Chan MC, Chan RW, Chan LL, Mok CK, Hui KP, Fong JH, et al. Tropism and innate host responses of a novel avian influenza A H7N9 virus: an analysis of

ex-vivo and in-vitro cultures of the human respiratory tract. *Lancet Respir Med*. 2013;1:534–42 [http://dx.doi.org/10.1016/S2213-2600\(13\)70138-3](http://dx.doi.org/10.1016/S2213-2600(13)70138-3).

- Couceiro JN, Paulson JC, Baum LG. Influenza virus strains selectively recognize sialyloligosaccharides on human respiratory epithelium; the role of the host cell in selection of hemagglutinin receptor specificity. *Virus Res*. 1993;29:155–65 [http://dx.doi.org/10.1016/0168-1702\(93\)90056-S](http://dx.doi.org/10.1016/0168-1702(93)90056-S).

Address for correspondence: Jingcao Pan, Microbiology Laboratory, Hangzhou Center for Disease Control and Prevention, Room 221, Building 5, Mingshi Rd, Jianqiao Zhen, Jianggan District, Hangzhou, Zhejiang Province, China; email: jingcaopan@gmail.com

Misidentification of *Diphyllobothrium* Species Related to Global Fish Trade, Europe

To the Editor: *Diphyllobothriosis*, infection by tapeworms of the genus *Diphyllobothrium* (Cestoda: Diphylllobothriidae) (1), is a well-known disease of humans. In Europe, infections caused by 3 species of *Diphyllobothrium* have recently been reported in humans: *D. latum* is considered to be the principal species infecting persons in Europe (1); 4 cases of *D. dendriticum* infection and 6 cases of *D. nihonkaiense* infection have also been reported (2,3). Except for those caused by *D. latum*, which is autochthonous in northeastern Europe and subalpine lakes, most of the cases in Europe have been imported or caused by consumption of fish imported from areas to which the parasites are endemic (1,3,4).

Diphyllobothriosis is not endemic to Spain, but 7 cases of *D. latum*