Human Papillomavirus Prevalence in Oropharyngeal Cancer before Vaccine Introduction, United States

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We conducted a study to determine prevalence of HPV types in oropharyngeal cancers in the United States and establish a prevaccine baseline for monitoring the impact of vaccination. HPV DNA was extracted from tumor tissue samples from patients in whom cancer was diagnosed during 1995–2005. The samples were obtained from cancer registries and Residual Tissue Repository Program sites in the United States. HPV was detected and typed by using PCR reverse line blot assays. Among 557 invasive oropharyngeal squamous cell carcinomas, 72% were positive for HPV and 62% for vaccine types HPV16 or 18. Prevalence of HPV-16/18 was lower in women (53%) than in men (66%), and lower in non-Hispanic Black patients (31%) than in other racial/ethnic groups (68%-80%). Results indicate that vaccines could prevent most oropharyngeal cancers in the United States, but their effect may vary by demographic variables.

Oropharyngeal cancers include malignancies that occur where the oral cavity and pharynx merge, including in the palatine and lingual tonsils, the posterior 1/3 (base) of the tongue, the soft palate, and the posterior pharyngeal wall. Current worldwide incidence has been estimated at

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The 2 HPV vaccines approved by the US Food and Drug Administration protect against infection with HPV-16 and HPV-18, which are the high-risk types most frequently associated with cervical cancer. A candidate 9-valent vaccine that includes types in the existing quadrivalent vaccine (HPV types 6, 11, 16, and 18) and 5 additional high-risk types (31, 33, 45, 52, and 58) is in clinical trial. Supported by evidence that existing vaccines effectively reduce oral HPV infections, these formulations may also reduce incidence of oropharyngeal cancers (7). When monitoring the population-level effect of HPV vaccination on oropharyngeal cancer occurrence in the United States, data on the incidence and type-specific prevalence of this disease are essential. Previously, the prevalence of cases attributable to viral infection and the consequent effects of vaccine programs were approximated from small published studies (8-13), which estimated HPV to be detected in 37%-60% of OPSCC in North America. Considering the range of prevalence, the heterogeneity of study populations, and differences in sample preparation and HPV detection methods used in these studies, it is not clear that this range of estimates reflects the true scope of HPV-associated OPSCC in the United States. Therefore, the objectives of this study were to determine prevalence of HPV types detected in oropharyngeal cancers in the United States and to establish a prevaccine baseline for monitoring the impact of vaccination.

DOI: http://dx.doi.org/10.3201/eid2005.131311

¹Members of HPV Typing of Cancers Workgroup who contributed to this study are listed at the end of this article.

Materials and Methods

Cancer Tissue Specimens

As part of the Centers for Disease Control Cancer Registry Sentinel Surveillance System study (M. Saraiya, unpub data), a systematic review of cases of oropharyngeal cancer diagnosed during 1995-2005 was performed. The cases were selected from 7 participating registries, including 4 central cancer registries in Florida, Kentucky, Louisiana, and Michigan and 3 Surveillance, Epidemiology, and End Results program (SEER [http://seer.cancer. gov/]) cancer registry-based residual tissue repositories in Los Angeles County, CA; Hawaii; and Iowa. The following anatomic regions (by ICD-O-3 codes) were included: C01.9 and C02.4 (base of the tongue and lingual tonsil); C09.0, C09.1, C09.8, C09.9, and c14.2 (tonsil); C14.0, C14.2, C14.8, C02.8, C10.2, C10.8, and C10.9 (other oropharynx) (13). Of 4,073 cases matching these criteria, we requested samples from 1,271 case-patients representative of the whole case-patient population regarding sex, age, and race/ethnicity. One archived, formalin-fixed paraffinembedded tissue sample, representative of the primary tumor, was selected by the submitting pathology laboratory. If tissue from the primary tumor was unavailable, a sample from a metastatic lesion in a lymph node was accepted because HPV prevalence is usually maintained in OPSCC-positive lymph nodes (14). With the exception of 32 cases from Hawaii and 11 from Los Angeles County, which had been sampled by Chaturvedi et al. (12), cases were selected exclusively for this study. Each participating state and CDC received approval from their institutional review boards for the study; CDC approved the overall study.

DNA Extraction and HPV Typing

All laboratory methods were described previously (15,16). Six consecutive 5-µm sections were cut from each selected tissue block; special precautions were used to avoid cross-contamination. The first and last sections were stained with hematoxylin and eosin and reviewed by a study pathologist (ERU) to confirm the presence of viable tumor tissue. DNA was extracted from two 5-µm sections by using high temperature-assisted tissue lysis (17) and further purification was carried out by automated extraction by using Chemagic MSM1 (PerkinElmer, Waltham, MA, USA). HPV types were determined from 2 commercial assays by using an algorithm which was evaluated earlier for this application (18,19). First, all DNA extracts were tested by using the Linear Array HPV Genotyping Assay (Linear Array; Roche Diagnostics, Indianapolis, IN, USA) and a HPV-52-specific PCR to resolve ambiguous positive results from the XR probe of the Linear Array HPV test (20). Samples that had negative or inadequate linear array results (negative for HPV and cellular β -globin controls) were

retested with the INNO-LiPA HPV Genotyping Assay (Innogenetics, Gent, Belgium). HPV status was recorded for 40 types: 6, 11, 16, 18, 26, 31, 33, 35, 39, 40, 42, 43, 44, 45, 51, 52, 53, 54, 55, 56, 58, 59, 61, 62, 64, 66, 67, 68, 69, 70, 71, 72, 73, 74, 81, 82, 83, 84, IS39, and 89 as tested; and HPV of an unknown type (HPV-X) for additional unspecified types as indicated by LiPA results.

Analysis

Prevalence was assessed as percentage positive from the total number of cases with valid results. HPV types 16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, 66, and 68 were considered to have a high risk for oncogenic potential (*21*) and all other types, including HPV-X, to have a low risk, showing low or no known oncogenic potential.

Hierarchical categories for HPV status were assigned as follows: HPV-16 includes all cases positive for this type regardless of other results. HPV-18 includes all cases positive for HPV-18, but not for HPV-16; other 9-valent, highrisk HPV types included in the next generation of the HPV vaccine: HPV-31, -33, -45, -52, -58, but not HPV-16 or -18; other high-risk HPV cases positive for any high risk not included in the previous categories: HPV-35, -39, -51, -66, -68; and low-risk HPV: all other cases positive for any remaining low-risk HPV types.

Statistical analysis was restricted to case-patients that had confirmed invasive OPSCC. Case-patient age at diagnosis was stratified into 4 groups: <50, 50– 59, 60–69, and \geq 70 years. Cancer stages were crudely classified as local, regional, or distant (metastatic) by SEER classifications. Differences in prevalence of positive results for high-risk HPV or HPV-16/18, categorized by patient's age, sex, race/ ethnicity, and the anatomic location of cancer, were evaluated by using the χ^2 or Fisher exact test whenever possible. Multivariate analysis was performed by using logistic regression with a step-down procedure, adjusting for age, sex, and race/ethnicity as appropriate. All statistical calculations were made by using SAS 9.3 (SAS Institute Inc., Cary, NC, USA).

Results

Of the 1,271 oropharyngeal tumors requested from the participating cancer registries, samples from 588 casepatients were received and successfully tested. Those not received were either unavailable or the remaining tissue was not representative of disease. The demographic characteristics (sex, age) and cancer stage (Table 1) (22) of the cohort from which the tested sample set was collected were similar to those of the untested cohort. Persons from the Asian Pacific Islands were few in number and slightly overrepresented in the final test population.

HPV results for 476 (81.0%) samples were from the linear array and 112 (19%) from LiPA. Most tissue was

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Table 1. Characteristics of oropharyngeal cancer case-patients
who provided samples compared with those of all eligible case-
patients, United States, 1995–2005*

patients, onited otates, re			
	% Case-patient	% Case-patient	
	samples not tested,	samples tested,	
Characteristic	n = 683	n = 588	
Age, y			
<50	18.4	18.5	
50–59	32.1	33.7	
60–69	27.5	27.6	
≥70	22.0	20.2	
Race/ethnicity			
Asian/Pacific Islander	1.3	4.4	
Non-Hispanic Black	11.9	12.6	
Hispanic	8.6	6.8	
Non-Hispanic White	77.6	75.2	
Other	0.6	1.0	
Sex			
F	23.4	25.4	
Μ	76.6	74.5	
Cancer stage			
Local	13.6	19.2	
Regional	52.0	56.5	
Distant	17.7	13.6	
Unknown	16.7	10.7	
*Samples were provided from 7 US registries.			

from the primary site (n = 473), but for 15 samples, only metastatic tissue from lymph nodes was available. Most case-patients (77.6%) were from urban areas or counties with a population >250,000. Most (94.4%) diagnoses were made during 2000 or later. Median age at the time of diagnosis was 58 (range 28–97) years. The male-to-female ratio was 3:1 and most of the cases (75.6%) were in non-Hispanic White persons. SCC, the most common histologic type of oropharyngeal cancer, accounted for 557 (94.7%) of all cases, and the main analysis was restricted to these cases (Table 2).

HPV was detected in 403 of the 557 OPSCC cases (72.4%) with valid typing results and 396 (71.1%) were positive for \geq 1 high-risk type (Table 3). In 68.4% of cases, a single HPV type was found; 3.9% contained 2 types. In 7 cases, only low-risk HPV types were detected: HPV-11, 26, 69, 82 (2 cases), 83, and HPV-X). HPV-16 was present in 337 (60.5%) cases, HPV-18 in 14 (2.5%) cases, and 331 (59.4%) cases were exclusively positive for these 2 types.

Other high-risk types, including HPV-31, -33, -35, -39, -45, and -52, were found at low frequency (Table 3). The relative prevalence in case-patients that had multiple HPV types essentially followed single-type distributions. HPV-16/33 was the most frequent combination (6 cases); HPV-16/18 and HPV-16/31 were the next most frequent, found in samples from 3 case-patients each. Frequencies of all co-detected HPV types are shown in Table 4. More than 2 types were not found in any of the oropharyngeal cancers.

Proportions of high-risk HPV prevalence and HPV-16/18 were statistically different among the registries and by race/ethnicity, stage, and anatomic subsite (Table 2). By sex, prevalence was only different for those infected with HPV-16/18. Age at diagnosis was not statistically different between the stratified groups, but median age at diagnosis among high-risk HPV positive case-patients was 58 (28–92) years and 61 (36–97) years in high-risk negative case-patients (p = 0.023).

According to hierarchical assessment, HPV-16 was found in 337 (60.5%) OPSCC cases, HPV-18 in 11 (2.0%), other 9-valent high-risk types in 32 (5.7%), other high-risk types in 16 (2.9%), and low-risk types in the remaining 7 (1.3%) cases (Figure 1). Of the 15 case-patients for whom lymph node metastases were tested, 14 were positive for high-risk HPV and 13 were positive for HPV-16.

In multivariate analysis for high-risk HPV that included age and sex in the model, only race/ethnicity was a significant independent factor (p = 0.003). Odds for high-risk

Table 2. High-risk and HP	V types	16 and 18 in orog	haryngeal
squamous cell carcinoma	s by der	nographic and tu	mor
characteristics, select Uni			
	Total	High-risk HPV,	,
Characteristic	no.	no. (%) pos.†	no. (%) pos.‡
Registry			
Los Angeles Co., CA	20	17 (85.0)	14 (70.0)
Florida	140	101 (72.1)	89 (63.6)
Hawaii	39	33 (84.6)	32 (82.1)
Iowa	13	4 (30.7)	4 (30.7)
Kentucky	116	74 (63.8)	69 (59.5)
Louisiana	95	75 (78.9)	61 (64.2)
Michigan	134	92 (68.6)	79 (59.0)
p value	NA	<0.001	0.032
Age, y			
<50	106	83 (78.3)	74 (69.8)
50–59	191	142 (74.3)	127 (66.5)
60–69	156	102 (65.4)	89 (57.1)
≥70	104	69 (65.4)	58 (55.8)
p value	NA	0.064	0.053
Sex			
F	141	91 (64.5)	74 (52.5)
Μ	416	305 (74.3)	274 (65.9)
p value	NA	0.053	0.006
Race/ethnicity			
Asian/Pacific Islander	20	16 (80.0)	16 (80.0)
Non-Hispanic Black	71	36 (50.7)	22 (31.0)
Hispanic	39	29 (74.4)	27 (69.2)
Non-Hispanic White	421	310 (73.6)	278 (67.5)
Other	6	5 (83.3)	5 (83.3)
p value	NA	0.002	< 0.001
Tumor stage			
Local	102	60 (58.8)	51 (50.0)
Regional	318	248 (78.Ó)	225 (70.8)
Distant (metastatic)	76	51 (6 7.1)	42 (55.3)
Unknown	61	37 (60.7)	30 (49.2)
p value	NA	0.001	<0.001
Tumor subsite			
Base of tongue	213	149 (70.0)	129 (60.6)
Tonsil	250	201 (80.4)	181 (72.4)
Other	94	46 (48.9)	38 (40.4)
p value	NA	<0.001	<0.001
*p values for differences betw	een cate	oories were calcula	ted by γ ² /Fisher

*p values for differences between categories were calculated by χ^2 /Fisher exact test. HPV, human papilloma virus; pos., positive; Co., county; NA, not applicable.

#HPV-16/18 = positive for HPV-16 and/or HPV-18.

[†]Positive for any of the high-risk types: HPV-16, -18, -31, -33, -35, -39, -45, -51, -52, -56, -58, -59, -66, -68.

Table 3. Human papillomavirus prevalence in oropharyngeal
squamous cell carcinomas, select United States registries, 1995–

2005*		
Variable	No. (%) cases, N = 557	
Characteristic		
HPV (any type)	403 (72.4)	
High risk†	396 (71.1)	
Low risk‡	7 (1.3)	
Negative	154 (27.6)	
Single type	381 (68.4)	
Multiple types§	22 (3.9)	
Туре		
HPV-16	337 (60.5)	
HPV-33	31 (5.6)	
HPV-18	14 (2.5)	
HPV-35	11 (2.0)	
HPV-39	5 (0.9)	
HPV-31	4 (0.7)	
HPV-52	4 (0.7)	
HPV-45	3 (0.5)	
Other HPV types	16 (2.9)	
*HPV, human papillomavirus; high risk, virus type is associated with high		

*HPV, human papillomavirus; high risk, virus type is associated with high oncogenic potential; low risk, virus type is associated with low oncogenic potential.

⁺Positive for any of HPV types 16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, 66, and 68.

‡Positive for types other than those identified as high risk.

§HPV-16/-33 (6 cases), HPV-16/-18 (3 cases), and HPV-16/-31(3 cases).

HPV infections were significantly higher for all other race groups than for non-Hispanic Black persons (p<0.001). When only HPV-16/18 detection was considered, significant differences were found in sex (p = 0.009) and race/ ethnicity in (p<0.001), but not age (p = 0.063), between those infected and those who were not (Table 5).

The 31 cases that had histologic results other than SSC included 7 adenocarcinomas (2 were HPV-16 positive, 5 HPV negative) and 2 small cell or neuroendocrine carcinomas (both HPV negative). Twenty-two cases were carcinomas not further specified, of which 7 tested positive for HPV (4 for HPV-16, and 1 each for HPV-18, HPV-33, HPV-35).

Discussion

Our finding of >70% HPV prevalence in a large sample of the oropharyngeal cancer patients from around the United States suggests that a substantially higher faction may be HPV-related than has been reported in many previous investigations (14). In a systematic review of HPV prevalence studies, including several small investigations of populations in North America, Kreimer et al. estimated that 47% of OPSCC cases were HPV related (11). Chaturvedi et al. more recently estimated weighted HPV prevalence at 72% (12), which is comparable to our findings. A continuous increase in HPV-related OPSCC that was observed during the past 20 years and escalated after 2000 (23) may explain some of the discrepancies found in the literature. More sensitive laboratory methods may also have contributed to the increased HPV prevalence relative to earlier investigations. All studies in North

America considered by Kreimer et al. (11) relied on the MY09/11 or GP5+/6+ consensus primer sets that require intact HPV L1 fragments of 450 and 150 bp, respectively. By contrast, in the study by Chaturvedi et al. (12) and ours, testing incorporated the INNO-LiPA assay, which has SFP primers that target 65-bp amplicons. Assays targeting smaller L1 amplicons can achieve increased sensitivity for HPV detection in formalin-fixed, paraffinembedded tissues known to have smaller DNA templates in their extracts than fresh or frozen samples do (18). Although not identical in specificity to that of the Linear Array, INNO-LiPA had shown comparable performance for detecting single type HPV infections predominantly found in cancer tissues (19).

Our current study results further confirm a trend of increasing incidence of tonsillar cancers with shifting demographic patterns (24, 25). The results further confirmed that HPV-16, detected in 84% of all positive tissues, was by far the most frequent type found in oropharyngeal cancers. Although type 16 also has nominally the highest prevalence in the "normal" oral cavity or oropharynx, other types are usually found at similar frequency (26,27). The ability of HPV-16 to establish persistent infection and its potential to transform might be responsible for its prominence in cancers. Currently available HPV vaccines targeting HPV-16 and -18 may be highly effective against OPSCC (9). A candidate 9-valent vaccine (currently in clinical trials) could have the potential to prevent virtually all HPV-associated oropharyngeal cancers: our data showed that 2.9% of the case-patients were positive for a high-risk type not covered in this formulation. (Figure 1).

The most noticeable differences were observed between racial groups, with notably fewer HPV-positive SCCs in non-Hispanic Black persons (50.7%) compared with non-Hispanic White persons (73.6%), Hispanic persons (74.4%), or Asian Pacific Islanders (80.0%). Other studies that noted similar differences by race/ethnicity found this to be a recent but ongoing development (28). Settle et al. (29), who investigated oral cancer survival, also reported reduced HPV prevalence in Black persons

Table 4 Oronharyngeal sguamous	cell carcinomas with >1 HPV	
Table 4. Oropharyngeal squamous cell carcinomas with >1 HPV		
type, select US registries, 1995-200		
Type combination	No. cases	
HPV-16 and -33	6	
HPV-16 and -18	3	
HPV-16 and -31	3	
HPV-16 and -35	2	
HPV-16 and -45	1	
HPV-16 and -52	1	
HPV-16 and -54	1	
HPV-16 and -59	1	
HPV-16 and -83	1	
HPV-18 and -35	1	
HPV-33 and -39	1	
HPV-39 and -56	1	

*No cases with >2 types were found; HPV, human papillomavirus.

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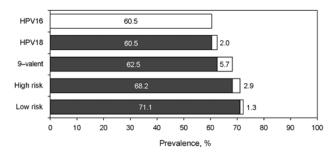


Figure. Hierarchical designation of human papillomavirus (HPV) types to oropharyngeal squamous cell carcinomas. White sections of bars indicate attribution of the specific HPV type or group. Black sections of bars indicate cumulative prevalence of types in higher hierarchy. HPV-16 includes all cases positive for this type regardless of other results. HPV-18 includes all cases positive for HPV-18, but negative for HPV-16. Cases of 9-valent HPV with high-risk HPV types included in the candidate 9-valent HPV vaccine: HPV-31, -33, -45, -52, -58, but not HPV-16 or -18. High-risk: cases positive for any high-risk type not included in the previous categories: HPV-35, -39, -51, -66, -68. Low-risk: cases only positive for HPV types with low or no oncogenic potential.

compared with other race/ethnicity groups and found that this difference was particular to oropharyngeal cancer and not to other cancers of the oral cavity.

In addition to differences by race/ethnicity, HPV prevalence also varied by sex, particularly for HPV-16/18. Prevalence was 66% among men, notably higher than the 53% found among women, which was a finding consistent with results of other investigations (2). Although further data stratification might show even greater dissimilarities, for instance between Black women and White men, the sample sizes for these analyses were modest and confidence intervals were large (data not shown). The precise causes for these discrepancies are unknown and most likely complex, but may be anticipated to influence vaccine efficacy for OPSCC. Prevalence differences observed between the registry states may be, in part, caused by demographic variations. Difference in age at diagnosis between patients with HPV-positive and HPV-negative cases was borderline significant (p = 0.023). Although other studies have also shown that HPV-positive cancers occur in a younger population (30), the role of 3 years difference in median age is unclear. It is possible that differences in behavior associated with causal pathways, such as smoking and drinking, provide a partial explanation. Persistent HPV infection at these anatomic sites may occur early, leading to more rapid and damaging alterations in cell cycle regulation and proliferation than those that occur with other carcinogenic exposures.

Of particular note, high-risk HPV types were detected in 80% of tonsillar SCCs. The microanatomy of the lymphoepithelial tissue of Waldeyer's ring, most notably the lingual and palatine tonsils, may explain this finding. Deep invaginations in this area by the tonsillar crypts may expose immature basal cells to HPV (31). The median age of case-patients was slightly lower than that of those with cancer in other sites (55 years), but proportions of infection, when sex and race/ethnicity were considered, were not different than proportions for the other oropharyngeal sites (data not shown). One limitation of this study is that not all participating sites were able to perform systematic random sampling of case-patient specimens from their eligible pool. The sizable specimen collections from the 4 cancer registries (Michigan, Kentucky, Louisiana, and Florida) were sampled by a simple random or systematic sampling approach, on the basis of the number of eligible cases. Sampling from the SEER tissue repositories (Los Angeles, California; Hawaii; and Iowa) was dependent on what tissue specimens were available. However, the resulting sample population that was ultimately tested represented diverse geographic regions and a wide range of demographic variables regarding sex, age, and race/ethnicity.

It should be noted that the presence of HPV DNA does not confirm its causal role in carcinogenesis. Detection in tumor tissues potentially overestimates the true involvement of the virus because coincidental, transient infections and complementary transforming effects to other factors cannot be distinguished. The natural history of cases in this study could not be assessed in this retrospective cross-sectional study and behavioral data were not available from the participating cancer registries. In particular, information regarding tobacco or alcohol use and HIV status would potentially improve estimation of the proportion of OPSCC caused by HPV alone. Because it is not clear at this point if HPV alone is sufficient to cause oropharyngeal cancer, factors other than use of tobacco products should be considered. Additional HPV markers, such as viral transcription (particularly E6 and E7 mRNA) or characteristic gene expression profiling, may provide further insights in future assessments and show distinction between actively transforming HPV infections and random, transient occurrences (32,33). Similar investigations may also be warranted to explicate the 7 cases

Table 5. Multivariate analysis for HPV and 18 detection in 557 oropharyngeal squamous cell carcinoma samples, select United States registries, 1995–2005*

Variable	OR (95% CI)†	p value
Sex		0.009
F	Ref	
Μ	1.70 (1.14–2.55)	
Race/ethnicity		<0.001
Non-Hispanic Black	Ref	_
Asian/Pacific Islander	8.43 (2.51–28.29)	_
Hispanic	4.73 (2.02–11.1)	_
Non-Hispanic White	4.34 (1.17–97.47)	-

*HPV, human papillomavirus; OR, odds ratio; Ref, reference group. †Odds ratio adjusted for race and ethnicity; age was not significant. in which only low-risk types were found. It is likely that these HPV types were present coincidentally and played no role in malignant transformation, but genomic changes that altered their pathogenic properties to bring them closer to those of high-risk types could provide an intriguing alternative explanation.

Conclusions

This study supports a role for oncogenic HPV in high proportions of oropharyngeal cancers. Future assessments are needed to monitor general prevalence and possible type-specific shifts. Data from the present and future studies will provide a baseline for early assessment of vaccine effects. Because the natural history and pre-cancer stages of oropharyngeal cancers are not established as they are for cervical cancer, direct trials with oropharyngeal neoplasia as the endpoint are not feasible. To obtain meaningful, comparable data for this objective, researchers need a universal definition of the anatomic oropharynx and associated malignancies and agreement on laboratory methods.

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Acknowledgment

We thank all members of the HPV Typing of Cancers Workgroup for their contributions toward this study.

The collection of original specimens from nonrepositories (Kentucky, Florida, Michigan, Louisiana), coordination of genotyping data from both the Surveillance, Epidemiology, and End Results (SEER) program and the National Program of Cancer Registries (NPCR) was largely supported by CDC intramural funds and Vaccine For Children Funds. This project has been supported in part with federal funds by CDC under grant nos. 5U58DP000810-5 (Kentucky), 5U58DP000844-5 (Florida), 5U58DP000812-5 (Michigan), and 5U58DP000769-5 (Louisiana); and with federal funds for Residual Tissue Repositories from the National Cancer Institute SEER Population-based Registry Program, National Institutes of Health, Department of Health and Human Services, under contract nos. N01-PC-35139 (Los Angeles), N01-PC-35143 (Iowa), and N01-PC-35137 (Hawaii).

The collection of data from California used in this publication was largely supported by the California Department of Health Services as part of the statewide cancer reporting program mandated by California Health and Safety Code Section 103885; by the National Cancer Institute, National Institutes of Health, Department of Health and Human Services, under contract no. N01-PC-2010-00035; and grant no. 1U58DP000807-3 from CDC.

B.Y.H. has received consultation and speaker fees from Merck and Co., Inc.

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References

- de Martel C, Ferlay J, Franceschi S, Vignat J, Bray F, Forman D, et al. Global burden of cancers attributable to infections in 2008: a review and synthetic analysis. Lancet Oncol. 2012;13:607–15. http://dx.doi.org/10.1016/S1470-2045(12)70137-7
- Centers for Disease Control and Prevention (CDC). Human papillomavirus-associated cancers—United States, 2004–2008. MMWR Morb Mortal Wkly Rep. 2012;61:258–61.
- Gillison ML. Human papillomavirus–associated head and neck cancer is a distinct epidemiologic clinical, and molecular entity. Semin Oncol. 2004;31:744–54. http://dx.doi.org/10.1053/j.seminoncol.2004.09.011
- D'Souza G, Kreimer AR, Viscidi R, Pawlita M, Fakhry C, Koch WM, et al. Case-control study of human papillomavirus and oropharyngeal cancer. N Engl J Med. 2007;356:1944–56. http://dx.doi.org/10.1056/NEJMoa065497
- Martinez I, Wang J, Hopson KF, Ferris RL, Khan SA. Identification of differentially expressed genes in HPV-positive and HPV-negative oropharyngeal squamous cell carcinomas. Eur J Cancer. 2007;43:415–32. http://dx.doi.org/10.1016/j.ejca.2006.09.001
- Miller DL, Puricelli MD, Stack MS. Virology and molecular pathogenesis of HPV (human papillomovirus)-associated oropharyngeal squamous cell carcinoma. Biochem J. 2012;443:339–53. http://dx.doi.org/10.1042/BJ20112017
- Herrero R, Quint W, Hildesheim A, Gonzalez P, Struijk L, Katki HA, et al. Reduced prevalence of oral human papillomavirus (HPV) 4 years after HPV vaccination in a randomized clinical trial in Costa Rica. PLoS ONE. 2013;8:e68329. http://dx.doi.org/10.1371/journal. pone.0068329

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- Watson M, Saraiya M, Ahmed F, Cardinez CJ, Reichman ME, Weir HK, et al. Using population-based cancer registry data to assess the burden of human papillomavirus-associated cancers in the United States: overview of methods. Cancer. 2008;113 (Suppl):2841–54. http://dx.doi.org/10.1002/cncr.23758
- Gillison ML, Chaturvedi AK, Lowy DR. HPV prophylactic vaccines and the potential prevention of noncervical cancers in both men and women. Cancer. 2008;113:3036–46. http://dx.doi.org/10.1002/ cncr.23764
- Fakhry C, Westra WH, Li S, Cmelak A, Ridge JA, Pinto H, et al. Improved survival of patients with human papillomavirus-positive head and neck squamous cell carcinoma in a prospective clinical trial. J Natl Cancer Inst. 2008;100:261–9. http://dx.doi.org/10.1093/ jnci/djn011
- Kreimer AR, Clifford GM, Boyle PB, Fransceschi S. Human papillomavirus types in head and neck squamous cell carcinomas worldwide: a systematic review. Cancer Epidemiol Biomarkers Prev. 2005;14:467–75. http://dx.doi.org/10.1158/1055-9965. EPI-04-0551
- Chaturvedi AK, Engels EA, Pfeiffer RM, Hernandez BY, Xiao W, Kim E, et al. Human papillomavirus and rising oropharyngeal cancer incidence in the United States. J Clin Oncol. 2011;29:4294– 301. http://dx.doi.org/10.1200/JCO.2011.36.4596
- Ryerson AB, Peters ES, Caughlin SS, Chen VW, Gillison ML, Reichman ME, et al. Burden of potentially human papillomavirusassociated cancer of the oropharynx and oral cavity in the US, 1998– 2003. Cancer. 2008;113:2901–9. http://dx.doi.org/10.1002/cncr.23745
- Mirghani H, Moreau F, Lefèvre M, Tam C, Périé S, Soussan P, et al. Human papillomavirus type 16 oropharyngeal cancer in lymph nodes as a marker of metastases. Arch Otolaryngol Head Neck Surg. 2011;137:910–4. http://dx.doi.org/10.1001/archoto.2011.141
- Gargano JW, Wilkinson EJ, Unger ER, Steinau M, Watson M, Huang Y, et al. Prevalence of human papillomavirus types in invasive vulvar cancers and vulvar intraepithelial neoplasia 3 in the United States before vaccine introduction. J Low Genit Tract Dis. 2012;16:471–9. http://dx.doi.org/10.1097/ LGT.0b013e3182472947
- Steinau M, Unger ER, Hernandez BY, Goodman MT, Copeland G, Hopenhayn C, et al. Human papillomavirus prevalence in invasive anal cancers in the United States prior to vaccine introduction. J Low Genit Tract Dis. 2013; 17:397-403. http://dx.doi.org/10.1097/ LGT.0b013e31827ed372
- Steinau M, Patel SS, Unger ER. Efficient DNA extraction for HPV genotyping in formalin-fixed, paraffin-embedded tissues. J Mol Diagn. 2011;13:377–81. http://dx.doi.org/10.1016/j. jmoldx.2011.03.007
- Hariri S, Steinau M, Rinas A, Gargano J, Ludema C, Unger ER, et al. HPV genotyping in high grade cervical lesions and invasive cervical carcinoma as detected by two commercial DNA assays, North Carolina, 2001–2006. PLoS ONE. 2012;7:e34044. http://dx.doi.org/10.1371/journal.pone.0034044
- Steinau M, Onyekwuluje JM, Scarbrough MZ, Unger ER, Dillner J, Zhou T. Performance of commercial reverse line blot assays for human papillomavirus genotyping. J Clin Microbiol. 2012;50:1539– 44. http://dx.doi.org/10.1128/JCM.06576-11
- Onyekwuluje JM, Steinau M, Swan DC, Unger ER. A real time PCR assay for HPV52 detection and viral load quantification. Clin Lab. 2012;58:61–6.
- Walboomers JM, Jacobs MV, Manos MM, Bosch FX, Kummer JA, Shah K, et al. Human papillomavirus is a necessary cause of

invasive cervical cancer worldwide. J Pathol. 1999;189:12–9. http://dx.doi.org/10.1002/(SICI)1096-9896(199909)189:1<12: AID-PATH431>3.0.CO;2-F

- 22. Watson M, Lyu C, Unger ER, Copeland G, Peters E, Huang Y, et al. Centers for Disease Control and Prevention human papillomavirus typing of cancers study with 7 registries: evaluating representativeness. North American Association of Central Cancer Registries (NAACCR) Annual Conference, 2011. Abstract, no. 58. http://www.naaccr.org/LinkClick.aspx?fileticket=udtv3Esreek%3d &tabid=257&mid=732
- Mehanna H, Beech T, Nicholson T, El-Hariry I, McConkey C, Paleri V, et al. Prevalence of human papillomavirus in oropharyngeal and nonoropharyngeal head and neck cancer-systematic review and meta-analysis of trends by time and region. Head Neck. 2013;35:747–55. http://dx.doi.org/10.1002/hed.22015
- Frisch M, Hjalgrim H, Jaeger AB, Biggar RJ. Changing patterns of tonisllar squamous cell carcinoma in the United States. Cancer Causes Control. 2000;11:489–95. http://dx.doi.org/10.1023/A: 1008918223334
- Warnakulasuriya S. Global epidemiology of oral and oropharyngeal cancer. Oral Oncol. 2009;45:309–16. http://dx.doi.org/10.1016/ j.oraloncology.2008.06.002
- Paolini F, Rizzo C, Sperduti I, Pichi B, Mafera B, Rahimi SS, et al. Both mucosal and cutaneous papillomaviruses are in the oral cavity but only alpha genus seems to be associated with cancer. J Clin Virol 2013;56:72–6 http://dx.doi.org/10.1016/j.jcv.2012.09.016
- Gillison ML, Broutian T, Pickard RK, et al. Prevalence of oral HPV infection in the United States, 2009–2010. JAMA. 2012;307:693– 703. http://dx.doi.org/10.1001/jama.2012.101
- Brown LM, Check DP, Devesa SS. Oropharyngeal cancer incidence trends: diminishing disparities. Cancer Causes Control. 2011;22:753–63. http://dx.doi.org/10.1007/s10552-011-9748-1
- Settle K, Posner MR, Schumaker LM, Tan M, Suntharalingam M, Goloubeva O, et al. Racial survival disparity in head and neck cancer results from low prevalence of human papillomavirus infection in black oropharyngeal cancer patients. Cancer Prev Res (Phila). 2009;2:776–81. http://dx.doi.org/10.1158/1940-6207.CAPR-09-0149
- Chaturvedi AK, Engels EA, Anderson WF, Gillison ML. Incidence trends for human papillomavirus-related and -unrelated oral squamous cell carcinomas in the United States. J Clin Oncol. 2008;26:612–9. http://dx.doi.org/10.1200/JCO.2007.14.1713
- Pai SI, Westra WH. Molecular pathology of head and neck cancer: implications for diagnosis, prognosis, and treatment. Annu Rev Pathol. 2009;4:49–70. http://dx.doi.org/10.1146/annurev. pathol.4.110807.092158
- 32. Jung AC, Briolat J, Millon R, de Reyniès A, Rickman D, Thomas E, et al. Biological and clinical relevance of transcriptionally active human papillomavirus (HPV) infection in oropharynx squamous cell carcinoma. Int J Cancer. 2010;126:1882–94. http://dx.doi. org/10.1002/ijc.24911
- Sethi S, Ali-Fehmi R, Franceschi S, Struijk L, van Doorn L-J, Quint W, et al. Characteristics and survival of head and neck cancer by HPV status: a cancer registry-based study. Int J Cancer. 2012;131:1179–86. http://dx.doi.org/10.1002/ijc.26500

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