

lung, although no specific lesions compatible with this infectious agent were observed.

A pool containing all morbillivirus-positive PCR amplicons for animals 1 and 2 (GenBank accession nos. KT006289 and KT006290), a PCR amplicon for the brain sample from animal 2 (GenBank accession no. KT006291), and a PCR amplicon for the larynx from animal 3 were sequenced. A BLAST search (<http://www.ncbi.nlm.nih.gov/blast/Blast.cgi>) showed that amplified samples were nearly identical to reference PWMV sequences (GenBank accession nos. AF200817 [3] and FJ842381 [8]). The sequence obtained from animal 3 was too short and degenerated to be accurately classified as CeMV, although it showed high homology with PWMV and porpoise morbillivirus.

It has been proposed that pilot whales might be enzootically infected with CeMV (10). These whales might be responsible for maintaining and transmitting CeMV over long distances or to other odontocetes. No die-offs have been observed in these species. However, an outbreak of a lethal morbillivirus infection in long-finned pilot whales caused by a dolphin morbillivirus strain occurred in the Mediterranean Sea during the end of October 2006–April 2007 (7).

Results of this study support the previous hypothesis that pilot whales have a species-adapted morbillivirus but indicate that lethal infections are not as rare as previously believed (3). The tropism of the virus in these cases, the high number of multinucleated syncytial cells, and the severity of the lesions resemble the acute systemic symptoms observed in dolphins infected with morbillivirus (2). Thus, pilot whales in the northeastern Atlantic Ocean could be at risk for infection, especially in one of the main pilot whale-watching regions between La Gomera and Southern Tenerife Islands in the Canary Islands, which has >700,000 visitors each year.

Acknowledgments

We thank other members of the Canary Islands Cetacean Stranding Network (Society for the Study of Cetaceans in the Canary Islands and Canary Islands Conservation) for participating in this study.

This study was supported by National Project CGL2012-39681 (Subprograma BOS); Regional Project SolSub C200801000288 and ProID 20100091; Technical Assistant Contract by Canary Islands Government delegation (TEC0002955); and precompetitive project ULPGC2013-21.

References

- Barrett T, Visser IK, Mamaev L, Goatley L, van Bresse MF, Osterhaust AD. Dolphin and porpoise morbilliviruses are genetically distinct from phocine distemper virus. *Virology*. 1993;193:1010–2. <http://dx.doi.org/10.1006/viro.1993.1217>
- Domingo M, Visa J, Pumarola M, Marco AJ, Ferrer L, Rabanal R, et al. Pathologic and immunocytochemical studies of morbillivirus infection in striped dolphins (*Stenella coeruleoalba*). *Vet Pathol*. 1992;29:1–10. <http://dx.doi.org/10.1177/030098589202900101>
- Taubenberger JK, Tsai MM, Atkin TJ, Fanning TG, Krafft AE, Moeller RB, et al. Molecular genetic evidence of a novel morbillivirus in a long-finned pilot whale (*Globicephalus melas*). *Emerg Infect Dis*. 2000;6:42–5. <http://dx.doi.org/10.3201/eid0601.000107>
- West KL, Sanchez S, Rotstein D, Robertson KM, Dennison S, Levine G, et al. A Longman's beaked whale (*Indopacetus pacificus*) strands in Maui, Hawaii, with first case of morbillivirus in the central Pacific. *Marine Mammal Science*. 2013;29:767–76.
- Groch KR, Colosio AC, Marcondes MC, Zucca D, Diaz-Delgado J, Niemeyer C, et al. Novel cetacean morbillivirus in Guiana dolphin, Brazil. *Emerg Infect Dis*. 2014;20:511–3. <http://dx.doi.org/10.3201/eid2003.131557>
- Stephens N, Duignan PJ, Wang J, Bingham J, Finn H, Bejder L 1st, et al. Cetacean morbillivirus in coastal Indo-Pacific bottlenose dolphins, Western Australia. *Emerg Infect Dis*. 2014;20:666–70. <http://dx.doi.org/10.3201/eid2004.131714>
- Fernández A, Esperon F, Herraiz P, de Los Monteros AE, Clavel C, Bernabe A, et al. Morbillivirus and pilot whale deaths, Mediterranean Sea. *Emerg Infect Dis*. 2008;14:792–4. <http://dx.doi.org/10.3201/eid1405.070948>
- Bellière EN, Esperon F, Fernandez A, Arbelo M, Munoz MJ, Sanchez-Vizcaino JM. Phylogenetic analysis of a new Cetacean morbillivirus from a short-finned pilot whale stranded in the Canary Islands. *Res Vet Sci*. 2011;90:324–8. <http://dx.doi.org/10.1016/j.rvsc.2010.05.038>
- VanDevanter DR, Warrener P, Bennett L, Schultz ER, Coulter S, Garber RL, et al. Detection and analysis of diverse herpesviral species by consensus primer PCR. *J Clin Microbiol*. 1996;34:1666–71.
- Duignan PJ, House C, Geraci JR, Early G, Copland HG, Walsh MT, et al. Morbillivirus infection in two species of pilot whales (*Globicephala* sp.) from the western Atlantic. *Marine Mammal Science*. 1995;11:150–62. <http://dx.doi.org/10.1111/j.1748-7692.1995.tb00514.x>

Address for correspondence: Eva Sierra, Institute for Animal Health, Veterinary School, University of Las Palmas de Gran Canaria, Arucas (Las Palmas), Canary Islands, Spain; email: esierra@becarios.ulpgc.es

Serogroup-specific Seasonality of Verotoxigenic *Escherichia coli*, Ireland

Patricia Garvey, Anne Carroll, Eleanor McNamara, André Charlett, Kostas Danis, Paul J. McKeown

Author affiliations: European Centre for Disease Prevention and Control (ECDC) European Programme for Intervention Epidemiology Training, Stockholm, Sweden (P. Garvey, K. Danis); Health Service Executive–Health Protection Surveillance Centre, Dublin, Ireland (P. Garvey, P.J. McKeown); ECDC Public Health Microbiology Training Programme, Stockholm (A. Carroll); Health Service Executive Public Health Laboratory–Dublin Mid-Leinster, Dublin (A. Carroll, E. McNamara); Public Health England, London, UK (A. Charlett); Institut de Veille Sanitaire, Paris, France (K. Danis)

DOI: <http://dx.doi.org/10.3201/eid2204.151160>

To the Editor: Globally, an increasing number of serogroups of verotoxigenic *Escherichia coli* (VTEC) have been reportedly associated with human illness. The best known is serogroup O157; the World Health Organization also recognizes VTEC O103, O111, O145, and O26 as having the potential to cause severe disease (1). The increasing number of non-O157 VTEC infections is cause for concern. In general, human infections with VTEC are reportedly more common in late summer; the European Centre for Disease Control and Prevention reported that the number of cases across the European Union peaks each year during July–September (2). Similarly, the United States reported that the number of VTEC O157 cases peaks in late summer (3).

Ireland is now one of the countries with the highest incidence of VTEC infection (2). In Ireland, statutory notification of VTEC infection became mandatory in 2004. In common with surveillance internationally, the focus was initially on VTEC O157; since then, testing and surveillance for non-O157 VTEC have improved substantially as a result of increased awareness and availability of diagnostic methods for non-O157 detection. Non-O157 VTEC were first reported in Ireland in 1999 (4), and surveillance data indicated that only 14% of VTEC notifications in 2004 compared with 75% in 2014 were caused by non-O157 VTEC.

In the notification dataset for Ireland, the 2 primary VTEC serogroups (O26 and O157) over many years have seemed to differ in their seasonality; VTEC O26 notifications generally peaked \approx 2 months earlier than VTEC O157 notifications (Figure, panel A). This earlier incidence peak for VTEC O26 has become progressively more consistent as the number of reported VTEC O26 notifications has risen. A study by Rivero et al. also suggested that non-O157 human infections may not exhibit the same seasonal variation observed for VTEC O157 (5).

In this study, we compared the seasonality of the 2 strains by using national notification data for 2004–2014 ($n = 2,569$ notifications for O157 and O26). We estimated the timing of the seasonal peaks (phase of seasonality) for each of the serogroups, and the difference between the 2 phases, by using times series quasi-Poisson regression, fitting terms for temporal trend, and a sine wave with a period of 12 months for seasonality and for interaction by serogroup. We compared the phase shifts of the 2 serogroups by using the Wald test. To rule out the possibility that the observed distributions were influenced by the occurrence of a limited number of outbreaks, we reanalyzed the data for sporadic cases alone and, because risk factors for VTEC infection have been shown to vary by age (6), separately for patients <5 years of age and for older child and adult patients.

The number of predicted cases peaked in July for VTEC O26 and in September for VTEC O157; the 2-month difference in phase (seasonality) by serogroup was significant ($p < 0.0001$) (Figure, panel B). The difference in seasonality remained significant ($p < 0.0001$) for sporadic cases alone; the predicted 2-month difference in seasonality was the same. The serogroup-dependent seasonality also remained when the data were analyzed separately for patients <5 years of age (predicted difference in phase 2 months, $p < 0.0001$) and ≥ 5 years of age (predicted difference in phase 1 month, $p < 0.0001$).

A significant increasing annual trend was also observed, in particular for VTEC O26. However, this increase is probably, at least in part, artifactual because of increased availability and more widespread use of clinical diagnostic tests for non-O157 VTEC in later years.

One possible explanation for the difference in seasonality is that the primary animal reservoirs for the 2 serogroups could differ. Cattle and sheep have been identified

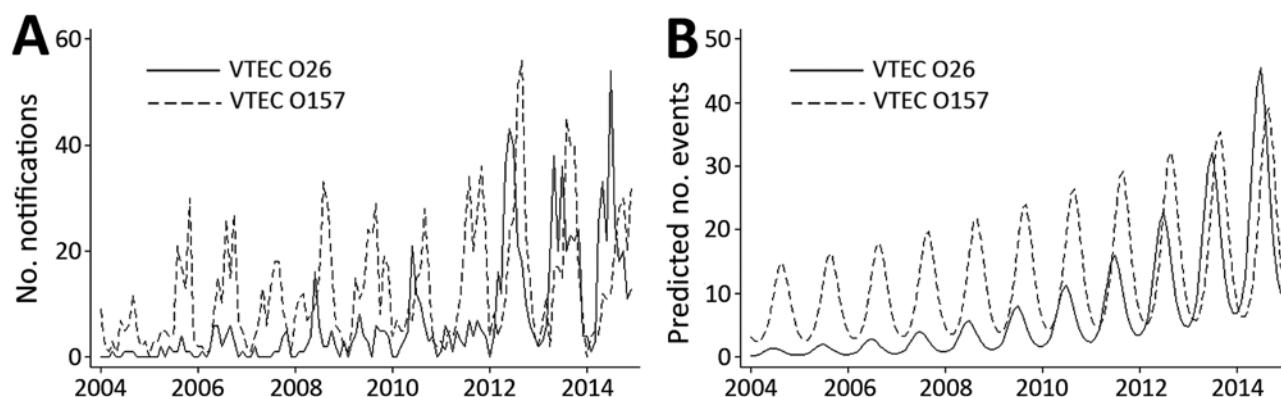


Figure. Verotoxigenic *Escherichia coli* (VTEC) O157 and VTEC O26, Ireland, 2004–2014. A) Seasonal distribution of notifications. B) Predicted seasonal distribution. Data source: Computerised Infectious Disease Reporting System (<https://www.hpsc.ie/NotifiableDiseases>) in Ireland, as of June 24, 2015. Predicted number of cases by month were derived from a cyclical quasi-Poisson model after trend and seasonality and interaction by serogroup were accounted for.

as carriers of O157 and O26 strains in Ireland (7,8). In Germany, cattle density has been shown to be significantly associated with human VTEC O157 incidence but only marginally associated with O26 incidence (9); the same study showed no association between cattle density and VTEC O91 infection, indicating that not all serogroups necessarily share the same reservoirs. Alternatively, animals of the same species may be preferentially colonized with different serogroups at different times of the year or at different developmental ages. Other possible explanations could be variation in survival characteristics between the 2 strains, which results in a different seasonal distribution in the environment, or specific human behavior (e.g., seasonal food) resulting in more frequent exposure to sources of VTEC O157 and VTEC O26 at different times of the year.

The consistent differences in seasonality identified here between the 2 most common VTEC serogroups suggest the existence of noteworthy underlying differences in disease etiology between the strains. Further exploration is recommended.

Acknowledgments

We acknowledge the cooperation of clinicians, laboratory directors, microbiologists, medical scientists, specialists in public health medicine, senior medical officers, surveillance scientists, infection control nurses, and principal environmental health officers in providing the dataset on which this report is based.

References

1. WHO Scientific Working Group. Zoonotic non-O157 Shiga toxin-producing *Escherichia coli* (STEC), 1995. Report of a WHO Scientific Working Group meeting; 1998 Jun 23–26; Berlin, Germany [cited 2016 Feb 5]. http://apps.who.int/iris/bitstream/10665/68880/1/WHO_CSR_APH_98.8.pdf
2. European Centre for Disease Prevention and Control. Annual epidemiological report 2013. Reporting on 2011 surveillance data and 2012 epidemic intelligence data. Stockholm: The Centre; 2013 [cited 2015 Jul 2]. <http://www.ecdc.europa.eu/en/publications/Publications/annual-epidemiological-report-2013.pdf>
3. Sodha SV, Heiman K, Gould LH, Bishop R, Iwamoto M, Swerdlow DL et al. National patterns of *Escherichia coli* O157 infections, USA, 1996–2011. *Epidemiol Infect.* 2015;143:267–73. <http://dx.doi.org/10.1017/S0950268814000880>
4. McMaster C, Roch EA, Willshaw GA, Doherty A, Kinnear W, Cheasty T. Verocytotoxin-producing *Escherichia coli* serotype O26:H11 outbreak in an Irish crèche. *Eur J Clin Microbiol Infect Dis.* 2001;20:430–2.
5. Rivero MA, Passucci JA, Rodríguez EM, Parma AE. Seasonal variation of HUS occurrence and VTEC infection in children with acute diarrhoea from Argentina. *Eur J Clin Microbiol Infect Dis.* 2012;31:1131–5. <http://dx.doi.org/10.1007/s10096-011-1418-4>
6. Werber D, Behnke SC, Fruth A, Merle R, Menzler S, Glaser S, et al. Shiga toxin-producing *Escherichia coli* infection in Germany: different risk factors for different age groups. *Am J Epidemiol.* 2007;165:425–34. <http://dx.doi.org/10.1093/aje/kwk023>
7. Thomas KM, McCann MS, Collery MM, Moschonas G, Whyte P, McDowell DA, et al. Transfer of verocytotoxigenic *Escherichia coli* O157, O26, O111, O103 and O145 from fleece to carcass during sheep slaughter in an Irish export abattoir. *Food Microbiol.* 2013;34:38–45. <http://dx.doi.org/10.1016/j.fm.2012.11.014>
8. Thomas KM, McCann MS, Collery MM, Logan A, Whyte P, McDowell DA, et al. Tracking verocytotoxigenic *Escherichia coli* O157, O26, O111, O103 and O145 in Irish cattle. *Int J Food Microbiol.* 2012;153:288–96. Epub 2011 Nov 29. <http://dx.doi.org/10.1016/j.ijfoodmicro.2011.11.012>
9. Frank C, Kapfhammer S, Werber D, Stark K, Held L. Cattle density and Shiga toxin-producing *Escherichia coli* infection in Germany: increased risk for most but not all serogroups. *Vector Borne Zoonotic Dis.* 2008;8:635–43. <http://dx.doi.org/10.1089/vbz.2007.0237>

Address for correspondence: Patricia Garvey, HSE-Health Protection Surveillance Centre, 25–27 Middle Gardiner Street, Dublin 1, Ireland; email: patricia.garvey@hse.ie

New Delhi Metallo- β -Lactamase-1-Producing *Klebsiella pneumoniae*, Florida, USA¹

Jun-Jie Li, L. Silvia Munoz-Price, Caressa N. Spychala, Dennise DePascale, Yohei Doi

Author affiliations: The Sixth People's Hospital, Shanghai Jiaotong University School of Medicine, Shanghai, China (J.-J. Li); University of Pittsburgh School of Medicine, Pittsburgh, Pennsylvania, USA (J.-J. Li, C.N. Spychala, Y. Doi); Medical College of Wisconsin, Milwaukee, Wisconsin, USA (L.S. Munoz-Price); Jackson Memorial Hospital, Miami, Florida, USA (D. DePascale)

DOI: <http://dx.doi.org/10.3201/eid2204.151176>

To the Editor: New Delhi metallo- β -lactamase (NDM)-producing *Enterobacteriaceae* have swiftly spread worldwide since an initial report in 2008 from a patient who had been transferred from India back home to Sweden (1). Epidemiologically, the global diffusion of NDM-1 producers has been associated with the Indian subcontinent and the Balkan region, which are considered the primary and secondary reservoirs of these pathogens, respectively (1). However, recent reports suggest that countries in the Middle East may constitute another potential reservoir for NDM-1 producers (1). More than 100 NDM-producing isolates have been reported in the United States, most of

¹Preliminary results from this study were presented at the 54th Interscience Conference on Antimicrobial Agents and Chemotherapy, September 5–9, 2014, Washington, DC, USA.