

2. Hennessey M, Fischer M, Staples JE. Zika virus spreads to new areas—region of the Americas, May 2015–January 2016. *MMWR Morb Mortal Wkly Rep.* 2016;65:55–8. <http://dx.doi.org/10.15585/mmwr.mm6503e1>
3. Lanciotti RS, Calisher CH, Gubler DJ, Chang GJ, Vorndam AV. Rapid detection and typing of dengue viruses from clinical samples by using reverse transcriptase-polymerase chain reaction. *J Clin Microbiol.* 1992;30:545–51.
4. Lanciotti RS, Kosoy OL, Laven JJ, Panella AJ, Velez JO, Lambert AJ, et al. Chikungunya virus in US travelers returning from India, 2006. *Emerg Infect Dis.* 2007;13:764–7. <http://dx.doi.org/10.3201/eid1305.070015>
5. Ayers M, Adachi D, Johnson G, Andonova M, Drebot M, Tellier R. A single tube RT-PCR assay for the detection of mosquito-borne flaviviruses. *J Virol Methods.* 2006;135:235–9. <http://dx.doi.org/10.1016/j.jviromet.2006.03.009>
6. Lanciotti RS, Kosoy OL, Laven JJ, Velez JO, Lambert AJ, Johnson AJ, et al. Genetic and serologic properties of Zika virus associated with an epidemic, Yap State, Micronesia, 2007. *Emerg Infect Dis.* 2008;14:1232–9. <http://dx.doi.org/10.3201/eid1408.080287>
7. Lanciotti RS, Lambert AJ, Holodniy M, Saavedra S, Signor LD. Phylogeny of Zika virus in Western Hemisphere, 2015. *Emerg Infect Dis.* 2016;22:933–5. <http://dx.doi.org/10.3201/eid2205.160065>
8. Thomas DL, Sharp TM, Torres J, Armstrong PA, Munoz-Jordan J, Ryff KR, et al. Local transmission of Zika virus—Puerto Rico, November 23, 2015–January 28, 2016. *MMWR Morb Mortal Wkly Rep.* 2016;65:154–8. <http://dx.doi.org/10.15585/mmwr.mm6506e2>
9. Leibovitz Z, Daniel-Spiegel E, Malinger G, Haratz K, Tamarkin M, Gindes L, et al. Microcephaly at birth—the accuracy of three references for fetal head circumference. How can we improve prediction? *Ultrasound Obstet Gynecol.* 2015;2:23. <http://dx.doi.org/10.1002/uog.15801>
10. World Health Organization. Guillain-Barré syndrome—Panama [cited 2016 Mar 29]. <http://www.who.int/csr/don/29-march-2016-gbs-panama/en/>

Address for correspondence: Brechla Moreno, Instituto Conmemorativo Gorgas de Estudios de la Salud, Ave Justo Arosemena calle 35 y 36, Panama City 0816-02593, Panama; email: bmoreno@gorgas.gob.pa

Influenza D Virus Infection in Herd of Cattle, Japan

Shin Murakami, Maiko Endoh, Tomoya Kobayashi, Akiko Takenaka-Uema, James K. Chambers, Kazuyuki Uchida, Masugi Nishihara, Benjamin Hause, Taisuke Horimoto

Author affiliations: University of Tokyo, Tokyo, Japan (S. Murakami, M. Endoh, T. Kobayashi, A. Takenaka-Uema, J.K. Chambers, K. Uchida, M. Nishihara, T. Horimoto); Kansas State University, Manhattan, Kansas, USA (B. Hause)

DOI: <http://dx.doi.org/10.3201/eid2208.160362>

To the Editor: Although the provisionally named influenza D virus was first isolated as an influenza C-like

virus from pigs with respiratory illness in Oklahoma in 2011 (1,2), epidemiologic analyses suggested that cattle are major reservoirs of this virus (3) and the virus is potentially involved in the bovine respiratory disease complex. The high rates of illness and death related to this disease in feedlot cattle are caused by multiple factors, including several viral and bacterial co-infections. Influenza D viruses were detected in cattle and pigs with respiratory diseases (and in some healthy cattle) in China (4), France (5), Italy (6), among other countries, indicating their wide global geographic distribution. Although the influenza D virus, like the human influenza C virus, is known to use 9-O-acetylated sialic acids as the cell receptor (2,7), its zoonotic potential is undefined because of limited research (1,8). We report influenza D virus infection in a herd of cattle in Japan.

To determine the presence of influenza D virus, on January 8, 2016, we used hemagglutination inhibition (HI) to test a convenience sample of 28 serum samples from healthy animals in a herd of female cattle in the Ibaraki Prefecture in central Japan. Two viruses with heterologous antigenicities, D/swine/Oklahoma/1334/2011 (D/OK) and D/bovine/Nebraska/9–5/2012 (D/NE) (9), were used for the assay with receptor-destroying enzyme (Denka: RDE II)-treated samples. Eight samples were positive for antibodies against both viruses, with HI titers of 1:80–1:640 for D/OK and with 2-fold or 4-fold lower HI titers (1:40–1:160) for D/NE in each sample, indicating previous infections in these cows, which ranged in age from ≈2 to 9 years. We also detected HI antibodies in serum samples from other cattle herds in several regions of Japan, although positivity rates varied (T. Horimoto, unpub. data). These data demonstrate the circulation of influenza D virus in Japan, as reported in other countries (3–6), emphasizing that the virus could be distributed worldwide.

Because 4 of the tested cows showed mild respiratory illness in January, we collected serum samples from the same 28 cows on February 3. At that time, only 1 cow still showed clinical signs; we collected a nasal swab sample from this cow. HI testing revealed that, of the 20 cows that had negative results in the first round of testing, 19 were positive for both D/OK and D/NE on the second test, which strongly confirms that influenza D virus infection had occurred and readily spread in this herd during January. However, most cows seemed to be subclinically infected with the virus. We cannot exclude the possibility of influenza D virus being a co-factor in causing respiratory illness because we did not evaluate the role of other viruses and bacteria in disease progression. HI titers (range 1:40–1:320) for D/NE were the same as or only 2 times lower than those for D/OK in all seroconversion samples, unlike the results for the seropositive samples in the first testing (online Technical Appendix, <http://wwwnc.cdc.gov/>

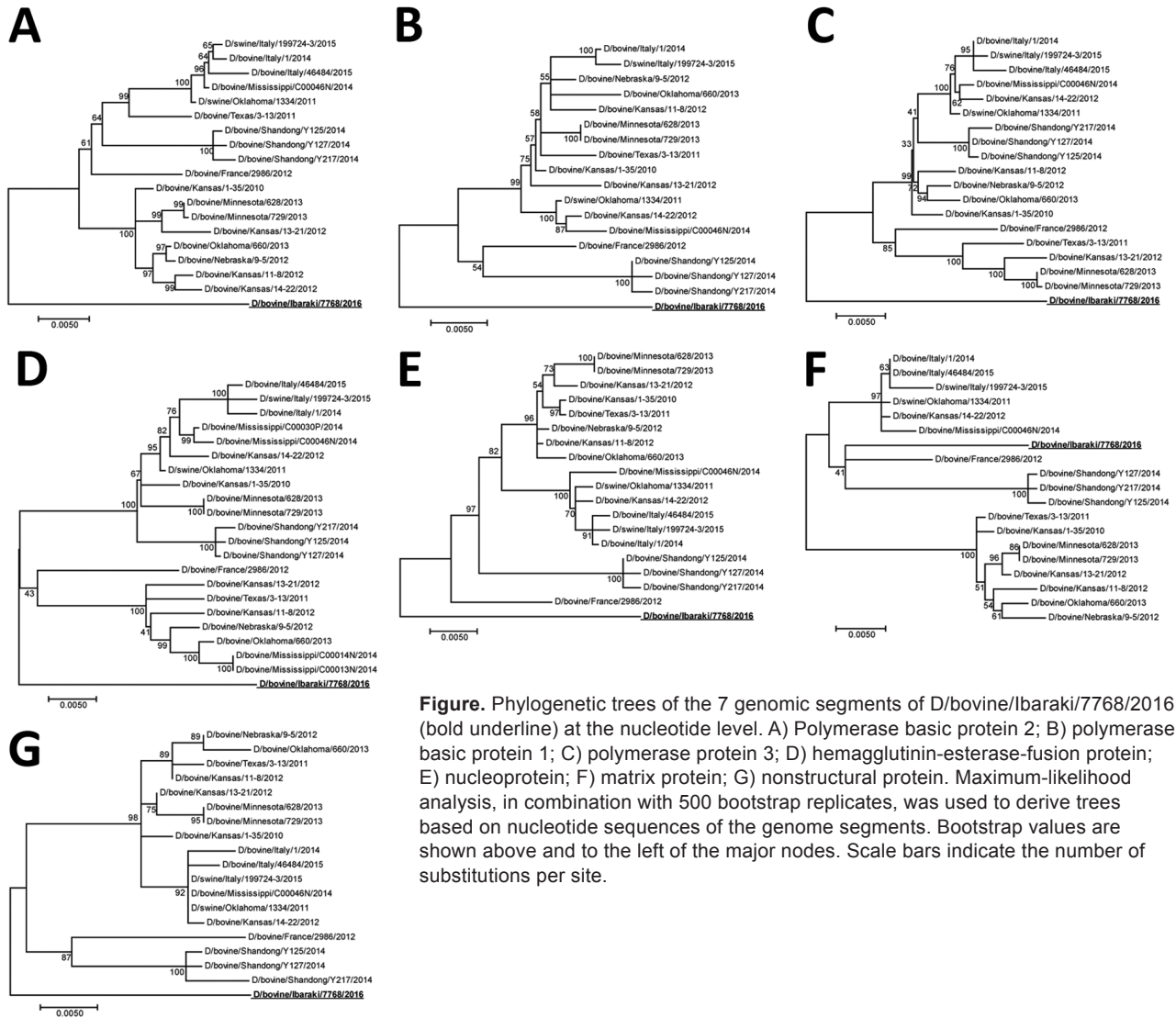


Figure. Phylogenetic trees of the 7 genomic segments of D/bovine/Ibaraki/7768/2016 (bold underline) at the nucleotide level. A) Polymerase basic protein 2; B) polymerase basic protein 1; C) polymerase protein 3; D) hemagglutinin-esterase-fusion protein; E) nucleoprotein; F) matrix protein; G) nonstructural protein. Maximum-likelihood analysis, in combination with 500 bootstrap replicates, was used to derive trees based on nucleotide sequences of the genome segments. Bootstrap values are shown above and to the left of the major nodes. Scale bars indicate the number of substitutions per site.

EID/article/22/8/16-0362-Techapp1.pdf). This result indicates that the virus that spread in this herd in January might be different from the one that infected some cows before the second testing, which suggests the presence of multiple strains with different antigenicities in this area of Japan. No increase in HI titers was observed in the second testing of samples from the 8 cows that were antibody-positive in the first testing.

We used the nasal swab sample from the 1 cow with clinical signs for virus detection, although this cow possessed the HI antibody. Reverse transcription PCR that used specific primers (available from the authors by request) successfully amplified the full genome sequence (GenBank accession nos. LC128433–9 for D/bovine/Ibaraki/7768/2016), which was aligned to the influenza D virus sequence. However, we could not isolate infectious virus by using sensitive cells (2), which might be attributable to the delayed swab sample collection.

Phylogenetic trees generated by using nucleotide sequences of individual segments from the Japan strain (95%–97% nucleotide identities with other strains) by maximum-likelihood analysis with ClustalW (<http://www.clustal.org>) and MEGA version 6.0 (10) indicated that this strain forms independent positions from strains isolated in other countries, although only the matrix segment was included in the same cluster as isolates from France and China (Figure). Although several unique amino acids of each protein exist in the strain from Japan, different from other isolates, their biologic characters are unknown. Among such residues, an amino acid at position 212 of hemagglutinin-esterase-fusion protein determined hemagglutinin antigenicity of the virus; lysine or arginine at this position resulted in heterologous antigenicities (9). The strain from Japan possessed serine at this position, identical to a strain from France (5), likely forming the third group of hemagglutinin antigenicity. Additionally, 1

putative N-glycosylation sequon was missing at positions 249–251 in hemagglutinin-esterase-fusion protein.

In summary, a cattle herd in Japan had influenza D virus infection. Although this study is a case report with a small number of samples, the observation shows a potential role for influenza D virus in the bovine respiratory disease complex.

References

1. Hause BM, Ducatez M, Collin EA, Ran Z, Liu R, Sheng Z, et al. Isolation of a novel swine influenza virus from Oklahoma in 2011 which is distantly related to human influenza C viruses. *PLoS Pathog.* 2013;9:e1003176. <http://dx.doi.org/10.1371/journal.ppat.1003176>
2. Hause BM, Collin EA, Liu R, Huang B, Sheng Z, Lu W, et al. Characterization of a novel influenza virus in cattle and Swine: proposal for a new genus in the *Orthomyxoviridae* family. *MBio.* 2014;5:e00031–14. <http://dx.doi.org/10.1128/mBio.00031-14>
3. Ferguson L, Eckard L, Epperson WB, Long LP, Smith D, Huston C, et al. Influenza D virus infection in Mississippi beef cattle. *Virology.* 2015;486:28–34. <http://dx.doi.org/10.1016/j.virol.2015.08.030>
4. Jiang WM, Wang SC, Peng C, Yu JM, Zhuang QY, Hou GY, et al. Identification of a potential novel type of influenza virus in bovine in China. *Virus Genes.* 2014;49:493–6. <http://dx.doi.org/10.1007/s11262-014-1107-3>
5. Ducatez MF, Pelletier C, Meyer G. Influenza D virus in cattle, France, 2011–2014. *Emerg Infect Dis.* 2015;21:368–71. <http://dx.doi.org/10.3201/eid2102.141449>
6. Chiapponi C, Faccini S, De Mattia A, Baioni L, Barbieri I, Rosignoli C, et al. Detection of influenza D virus among swine and cattle, Italy. *Emerg Infect Dis.* 2016;22:352–4. <http://dx.doi.org/10.3201/eid2202.151439>
7. Song H, Qi J, Khedri Z, Diaz S, Yu H, Chen X, et al. An open receptor-binding cavity of hemagglutinin-esterase-fusion glycoprotein from newly-identified influenza D virus: basis for its broad cell tropism. *PLoS Pathog.* 2016;12:e1005411. <http://dx.doi.org/10.1371/journal.ppat.1005411>
8. Smith DB, Gaunt ER, Digard P, Templeton K, Simmonds P. Detection of influenza C virus but not influenza D virus in Scottish respiratory samples. *J Clin Virol.* 2016;74:50–3. <http://dx.doi.org/10.1016/j.jcv.2015.11.036>
9. Collin EA, Sheng Z, Lang Y, Ma W, Hause BM, Li F. Cocirculation of two distinct genetic and antigenic lineages of proposed influenza D virus in cattle. *J Virol.* 2015;89:1036–42. <http://dx.doi.org/10.1128/JVI.02718-14>
10. Tamura K, Stecher G, Peterson D, Filipowski A, Kumar S. MEGA6: molecular evolutionary genetics analysis version 6.0. *Mol Biol Evol.* 2013;30:2725–9. <http://dx.doi.org/10.1093/molbev/mst197>

Address for correspondence: Taisuke Horimoto, Laboratory of Veterinary Microbiology, Graduate School of Agricultural and Life Sciences, University of Tokyo, Yayoi 1-1-1, Bunkyo-ku, Tokyo 113-8657, Japan; email: ahorimo@mail.ecc.u-tokyo.ac.jp

Fatal Septic Meningitis in Child Caused by *Streptococcus suis* Serotype 24

Anusak Kerdsin, Marcelo Gottschalk, Rujirat Hatrongjit, Shigeyuki Hamada, Yukihiro Akeda, Kazunori Oishi

Author affiliations: Kasetsart University Chalermphrakiat Sakon Nakhon Province Campus, Sakon Nakhon, Thailand (A. Kerdsin, R. Hatrongjit); University of Montreal, Montreal, Quebec, Canada (M. Gottschalk); Osaka University, Osaka, Japan (S. Hamada, Y. Akeda); National Institute of Infectious Diseases, Tokyo, Japan (K. Oishi)

DOI: <http://dx.doi.org/10.3201/eid2208.160452>

To the Editor: *Streptococcus suis* is a zoonotic bacterium that causes invasive infections in humans and pigs (*I*). Of the 29 described serotypes, serotype 2 is the most prevalent in humans, almost exclusively affecting adults (*I*). Other serotypes occurring sporadically in humans have been reported (*I*). Here we report a rare case of *S. suis* serotype 24 infection in a child.

A 2-year-old girl with Down syndrome was admitted to a hospital in Rayong Province, eastern Thailand, in May 2015. She had a high fever of 3 days' duration, vomiting, stiff neck, rash, and purpura on her right leg and hip. The initial diagnosis was cellulitis and suspected meningococcal meningitis. Physical examination revealed a temperature of 39.5°C, pulse rate of 160 beats/min, respiratory rate of 80 breaths/min, and blood pressure of 94/55 mm Hg. Oxygen saturation was 80%, which is indicative of severe respiratory failure. An analysis of the complete blood count showed a leukocyte count of 21,460 cells/mL (83% neutrophils, 13% lymphocytes, 1% eosinophils, 3% monocytes) and platelet count of 155,000 cells/ μ L. A comprehensive metabolic panel test was not performed.

Bacteria were isolated from the patient's cerebrospinal fluid culture; however, hemoculture did not show any growth. Traditional biochemical tests and an API20Strep system assay (BioMérieux, Marcy l'Etoile, France) suggested that the organism was *S. suis*. The samples tested positive for *S. suis* serotype 24 by multiplex PCR and coagglutination testing (2,3). On the basis of these results, the condition was diagnosed as septic meningitis. Unfortunately, the patient died the day after admission, even though she had been treated with ceftriaxone on the day of admission.

The isolate from the child was susceptible to penicillin (MIC \leq 0.12 μ g/mL), ceftriaxone (MIC \leq 1 μ g/mL), erythromycin (MIC \leq 0.25 μ g/mL), levofloxacin (MIC \leq 2.0 μ g/mL), clindamycin (MIC \leq 0.25 μ g/mL), and vancomycin (MIC \leq 1.0 μ g/mL). Because breakpoints for *S. suis* are not

Influenza D Virus Infection in Herd of Cattle, Japan

Technical Appendix

Technical Appendix Table. Detection of HI antibodies to influenza D virus in a herd of cows in Ibaraki Prefecture, Japan*

Cow no.	Age (as of 2016 Jan 8)	HI test titers				Clinical sign	Detection of virus
		2016 Jan 8		2016 Feb 3			
		D/OK	D/NE	D/OK	D/NE		
7754	5 y 6 mo	<40	<40	40	40		
7761	4 y 5 mo	<40	<40	80	80		
7764	3 y 11 mo	<40	<40	160	80		
7765	3 y 10 mo	<40	<40	<40	<40		
7766	3 y 4 mo	<40	<40	160	160		
7767	3 y 4 mo	<40	<40	80	80	Respiratory illness	
7768	3 y 4 mo	<40	<40	160	160	Respiratory illness	Nasal swab RT-PCR (+)
7769	3 y 3 mo	<40	<40	320	320	Respiratory illness	
7770	3 y 1 mo	<40	<40	160	160	Respiratory illness	
7777	2 y 2 mo	<40	<40	160	80		
7779	2 y 1 mo	<40	<40	160	80		
7780	2 y 1 mo	<40	<40	80	80		
7781	2 y 0 mo	<40	<40	80	40		
7782	1 y 12 mo	<40	<40	160	80		
7784	1 y 7 mo	<40	<40	160	80		
7787	1 y 3 mo	<40	<40	160	160		
7788	1 y 0 mo	<40	<40	80	40		
7791	0 y 11 mo	<40	<40	40	40		
7795	0 y 8 mo	<40	<40	320	160		
7796	0 y 7 mo	<40	<40	80	80		
0472	7 y 9 mo	80	40	160	80		
0514	8 y 0 mo	160	40	80	40		
1336	6 y 11 mo	80	40	160	80		
1337	6 y 3 mo	640	160	320	160		
1412	7 y 10 mo	160	40	80	40		
5266	1 y 11 mo	80	40	160	80		
5270	1 y 10 mo	80	40	40	<40		
8003	9 y 0 mo	160	40	80	40		

D/NE: D/bovine/Nebraska/9-5/2012; D/OK: D/swine/Oklahoma/1334/2011; HI, hemagglutination-inhibition; RT-PCR, reverse transcription PCR.