

Acknowledgments

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References

1. Bessen RA, Marsh RF. Identification of two biologically distinct strains of transmissible mink encephalopathy in hamsters. *J Gen Virol.* 1992;73:329–34. <http://dx.doi.org/10.1099/0022-1317-73-2-329>
2. Capobianco R, Casalone C, Suardi S, Mangieri M, Miccolo C, Limido L, et al. Conversion of the BASE prion strain into the BSE strain: the origin of BSE? *PLoS Pathog.* 2007;3:e31. <http://dx.doi.org/10.1371/journal.ppat.0030031>
3. Duque Velásquez C, Kim C, Herbst A, Daude N, Garza MC, Wille H, et al. Deer prion proteins modulate the emergence and adaptation of chronic wasting disease strains. *J Virol.* 2015; 89:12362–73. <http://dx.doi.org/10.1128/JVI.02010-15>
4. Gossert AD, Bonjour S, Lysek DA, Fiorito F, Wüthrich K. Prion protein NMR structures of elk and of mouse/elk hybrids. *Proc Natl Acad Sci U S A.* 2005;102:646–50. <http://dx.doi.org/10.1073/pnas.0409008102>
5. Sigurdson CJ, Nilsson KP, Hornemann S, Heikenwalder M, Manco G, Schwarz P, et al. De novo generation of a transmissible spongiform encephalopathy by mouse transgenesis. *Proc Natl Acad Sci U S A.* 2009;106:304–9. <http://dx.doi.org/10.1073/pnas.0810680105>
6. Raymond GJ, Raymond LD, Meade-White KD, Hughson AG, Favara C, Gardner D, et al. Transmission and adaptation of chronic wasting disease to hamsters and transgenic mice: evidence for strains. *J Virol.* 2007;81:4305–14. <http://dx.doi.org/10.1128/JVI.02474-06>
7. Kurt TD, Bett C, Fernández-Borges N, Joshi-Barr S, Hornemann S, Rüllicke T, et al. Prion transmission prevented by modifying the $\beta 2$ - $\alpha 2$ loop structure of host PrP^C. *J Neurosci.* 2014;34:1022–7. <http://dx.doi.org/10.1523/JNEUROSCI.4636-13.2014>
8. Race B, Meade-White KD, Phillips K, Striebel J, Race R, Chesebro B. Chronic wasting disease agents in nonhuman primates. *Emerg Infect Dis.* 2014;20:833–7. <http://dx.doi.org/10.3201/eid2005.130778>
9. Barria MA, Telling GC, Gambetti P, Mastrianni JA, Soto C. Generation of a new form of human PrP(Sc) in vitro by interspecies transmission from cervid prions. *J Biol Chem.* 2011;286:7490–5. <http://dx.doi.org/10.1074/jbc.M110.198465>
10. Le Dur A, Béringue V, Andréoletti O, Reine F, Lai TL, Baron T, et al. A newly identified type of scrapie agent can naturally infect sheep with resistant PrP genotypes. *Proc Natl Acad Sci U S A.* 2005;102:16031–6. <http://dx.doi.org/10.1073/pnas.0502296102>

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Rabies Virus Transmission in Solid Organ Transplantation, China, 2015–2016

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We report rabies virus transmission among solid organ transplantation recipients in Changsha, China, in 2016. Two recipients were confirmed to have rabies and died. Our findings suggest that more attention should be paid to the possibility of rabies virus transmission through organ transplantation for clinical and public health reasons.

In 2016, Zhou et al. reported a case of rabies virus transmission in China that was probably a result of organ transplantation (1). We report on rabies transmission that occurred among solid organ transplant recipients in Changsha, China, during December 2015–January 2016.

In November 2015, the donor, a previously healthy boy, showed development of fever, insomnia, and agitation. On day 6 of infection, these symptoms progressed, and he was sent to a healthcare center. At this time, he experienced weariness, no desire to drink water, poor appetite, and panic. One day later, he began vomiting, and was admitted to a local hospital (hospital A), where he exhibited anemophobia, convulsions, limb rigidity, and hypersalivation. The patient was moved to hospital B (days 7–14) in Changsha. At admission, some examination findings indicated a possibility of viral encephalitis (online Technical Appendix, Table 1, <https://wwwnc.cdc.gov/EID/article/23/9/16-1704-Techapp1.pdf>). Subhypothermia hibernation therapy and assisted ventilation were administered within 72 hours of admission, and the patient's vital signs became stable. On day 10, hyponatremia was observed, and on day 11, the patient again became febrile and tachycardic, with hypertensive abdominal distention and alimentary tract hemorrhage. On day 13, viral encephalitis was diagnosed, and rabies was suspected. However, rabies virus antibody tests performed on serum samples by using ELISA yielded negative results.

On day 14, the patient was transferred to hospital C, where he became comatose and was declared brain dead. Permission was granted for organ donation, because no specific pathogen had been detected and China's organ

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transplant policy allows for organ donations in cases of viral encephalitides without laboratory-confirmed pathogens. Rabies virus–specific binding antibodies were also not detected in the serum samples. The donor had possibly been exposed to rabies but had not been vaccinated against rabies (online Technical Appendix). The patient’s kidneys and liver were removed for transplantation in hospital D in Changsha on the following day.

Kidney and liver transplantations were performed on December 10, 2015. Two female recipients in hospital D received the kidneys, and an 8-month-old girl in hospital E in Shanghai, China, received the liver. No other organs were collected or transplanted from this donor. The surgeries were all uneventful, and the recipients were discharged after transplantation. However, all 3 recipients were eventually readmitted to the hospital with complex symptoms (Figure).

On the 40th day after transplantation, the left kidney recipient (29 years of age) revisited hospital D, reporting worsening right lower quadrant abdominal pain and vomiting. In the next 3 days, she successively had anemophobia, shortness of breath, chest tightness, anxiety, insomnia, and hypersalivation. During hospitalization, her blood pressure was as high as 248/148 mm Hg. On her 7th day in the hospital, she became comatose and then died.

On the 43rd day after transplantation, the right kidney recipient (age 47 years) developed aches in her lower limbs, insomnia, and a decreased appetite. She was readmitted to hospital D the next day. On the 3rd day after admission, she exhibited hydrophobia, anemophobia, photesthesia, and fatigue. She showed agitation, dyspnea, and hypersalivation on the 4th day; she became comatose and died 1 day later.

The liver recipient was readmitted with pneumonia on the 34th day after transplantation and died of asphyxia and

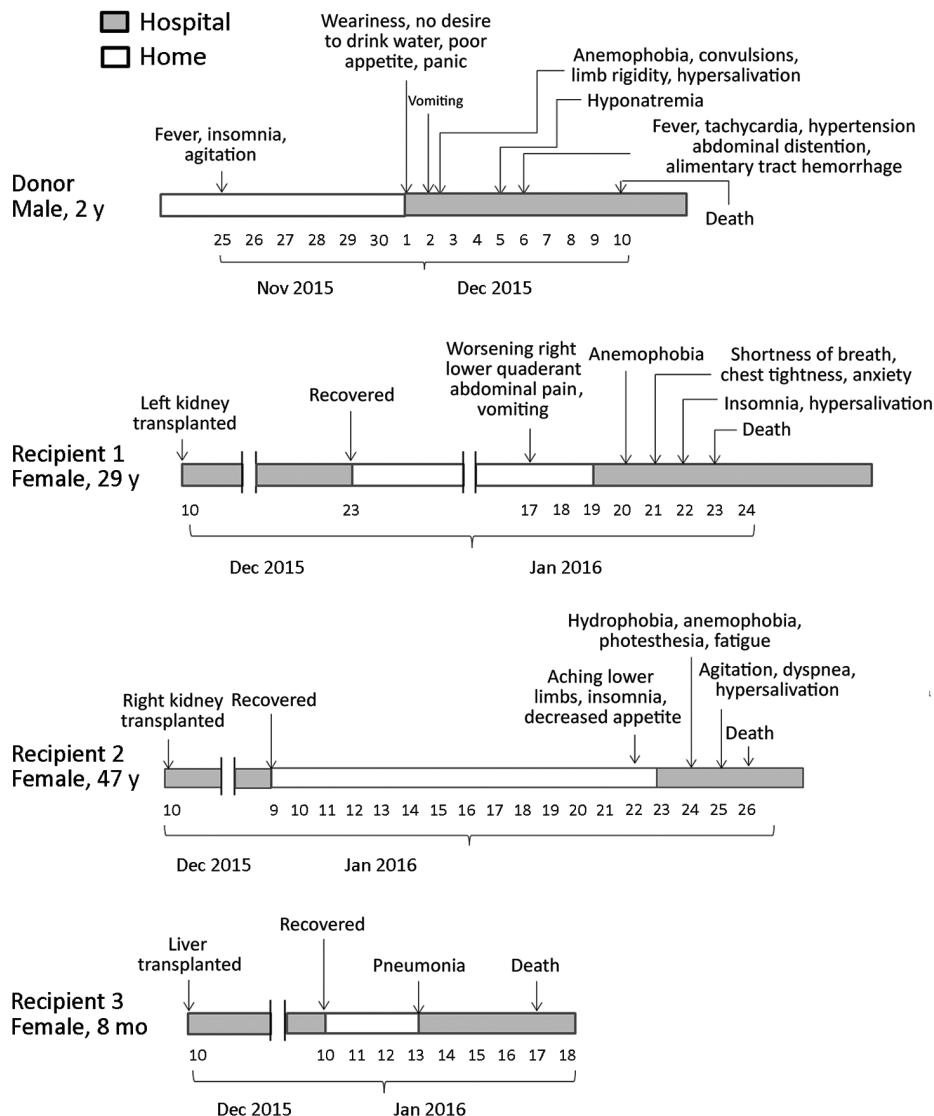


Figure. Clinical course of a donor and 3 recipients in a rabies outbreak associated with solid organ transplantation, Changsha, China, 2015–2016.

multiple organ failure within 5 days. This patient did not show any signs or symptoms of rabies or encephalitis.

None of the recipients had been exposed to potentially rabid animals or had been vaccinated previously for rabies (online Technical Appendix). Both kidney recipients tested positive for rabies virus (online Technical Appendix Table 2). The genome sequences of the rabies virus isolates from the right kidney recipient (isolate no. CCS1501H) were \approx 11 kb nucleotides in length and belonged to the China I lineage. No testing for rabies was done on the donor or on the liver recipient.

In the past 10 years, rabies transmission by solid organ transplantation has been described occasionally worldwide (2–4). Hence, rabies transmission through organ transplantation is a clinical and public health concern. To prevent future cases such as this, we recommend that patients with unexplained encephalitis or mental status changes should not be used as organ donors even if tests for some infectious causes of encephalitis are negative. In addition, if rabies is suspected in the donor after organs have been transplanted, the recipients should also not be used as organ donors. An antibody test is not the ideal choice for the diagnosis of rabies virus and by itself cannot reliably exclude rabies from the differential diagnosis. For this reason, a combination of multiple techniques, preferably direct fluorescent antibody test and reverse transcription PCR, should be used before organ transplantation, especially when the donor is suspected of having rabies or a potential exposure to rabies. In addition, if a patient has meningoencephalitis of unknown cause, a specific epidemiologic and laboratory evaluation should be performed to conclusively rule out rabies as a cause of illness before organ donation.

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References

- Zhou H, Zhu W, Zeng J, He J, Liu K, Li Y, et al. Probable rabies virus transmission through organ transplantation, China, 2015. *Emerg Infect Dis*. 2016;22:1348–52. <http://dx.doi.org/10.3201/eid2208.151993>
- Srinivasan A, Burton EC, Kuehnert MJ, Rupprecht C, Sutker WL, Ksiazek TG, et al.; Rabies in Transplant Recipients Investigation Team. Transmission of rabies virus from an organ donor to four transplant recipients. *N Engl J Med*. 2005;352:1103–11. <http://dx.doi.org/10.1056/NEJMoa043018>
- Bronnert J, Wilde H, Tepsumethanon V, Lumlertdacha B, Hemachudha T. Organ transplantations and rabies transmission. *J Travel Med*. 2007;14:177–80. <http://dx.doi.org/10.1111/j.1708-8305.2006.00095.x>
- Vora NM, Basavaraju SV, Feldman KA, Paddock CD, Orciari L, Gitterman S, et al.; Transplant-Associated Rabies Virus Transmission Investigation Team. Raccoon rabies virus variant transmission through solid organ transplantation. *JAMA*. 2013;310:398–407. <http://dx.doi.org/10.1001/jama.2013.7986>

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Identification of Clade E Avipoxvirus, Mozambique, 2016

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Analysis of scab samples collected from poultry during outbreaks of fowlpox in Mozambique in 2016 revealed the presence of clade E avipoxviruses. Infected poultry were from flocks that had been vaccinated against fowlpox virus. These findings require urgent reevaluation of the vaccine formula and control strategies in this country.

Avipoxviruses are large, enveloped DNA viruses that belong to the genus *Avipoxvirus* in the *Chordopoxvirinae* subfamily of the family *Poxviridae*. These viruses cause disease in a large number of bird species and are generally named after the species from which the virus was first isolated and characterized (*1*). Fowlpox virus (FPV) has caused substantial economic losses in domestic poultry resulting from reduced egg production and growth, blindness, and death, with a death ratio that can reach as high as 50%.

Phylogenetic analyses of the *Avipoxvirus* genus are usually conducted with the segments of the genes encoding the 4b core-like protein (P4b) and the DNA polymerase, which are both highly conserved among poxviruses (2,3). Using these loci, researchers have seen that most strains cluster into 3 major clades, namely A, B, and C, with clade A being subdivided further into subclades A1–A7 and clade B into subclades B1–B3 (3–5). Two additional clades