

Acknowledgments

We thank Luiza Vasconcellos and the residency program in the Sanitary Surveillance of INCQS/Fiocruz. We are grateful for access to the sequencing core “Plataforma Genômica de Sequenciamento de DNA/PDTIS-FIOCRUZ.”

About the Author

Ms. Volpe is an infectious diseases specialist and PhD student in infectious diseases at the Federal University of Mato Grosso do Sul, Brazil, with research interests in hospital infectious disease control.

References

1. Friedemann M. Epidemiology of invasive neonatal *Cronobacter* (*Enterobacter sakazakii*) infections. *Eur J Clin Microbiol Infect Dis*. 2009;28:1297–304. <http://dx.doi.org/10.1007/s10096-009-0779-4>
2. International Organization for Standardization, Microbiology of the food chain—horizontal method for the detection of *Cronobacter* spp. ISO 22964:2017. Geneva: The Organization; 2017.
3. Chen Y, Lampel K, Hammack T. *Cronobacter*. In: Bacteriological analytical manual. 8th ed. Revision A, 1998. Rockville (MD): US Department of Health and Human Services, Food and Drug Administration; 2012 [cited 2018 Aug 20]. <https://www.fda.gov/Food/FoodScienceResearch/LaboratoryMethods/ucm289378.htm>
4. McMullan R, Menon V, Beukers AG, Jensen SO, van Hal SJ, Davis R. *Cronobacter sakazakii* infection from expressed breast milk, Australia. *Emerg Infect Dis*. 2018;24:393–4. <http://dx.doi.org/10.3201/eid2402.171411>
5. Brandão MLL, Umeda NS, Carvalho KR, Filippis I. Investigation of an outbreak caused by *Cronobacter malonicus* in a maternity hospital in Teresina, Piauí: characterization and typing by pulsed field gel electrophoresis [in Portuguese]. *Vig Sanit Debate*. 2015;3:91–6.
6. Barreira ER, Souza ER, Gois DC, Freitas PF, Fernandes JC. *Enterobacter sakazakii* meningitis in a newborn infant: case report [in Portuguese]. *Pediatrics (São Paulo)*. 2003;25:65–70.
7. Umeda NS, de Filippis I, Forsythe SJ, Brandão MLL. Phenotypic characterization of *Cronobacter* spp. strains isolated from foods and clinical specimens in Brazil. *Food Res Int*. 2017;102:61–7. <http://dx.doi.org/10.1016/j.foodres.2017.09.083>
8. Forsythe SJ, Dickins B, Jolley KA. *Cronobacter*, the emergent bacterial pathogen *Enterobacter sakazakii* comes of age; MLST and whole genome sequence analysis. *BMC Genomics*. 2014;15:1121. <http://dx.doi.org/10.1186/1471-2164-15-1121>
9. Hariri S, Joseph S, Forsythe SJ. *Cronobacter sakazakii* ST4 strains and neonatal meningitis, United States. *Emerg Infect Dis*. 2013;19:175–7. <http://dx.doi.org/10.3201/eid1901.120649>

Address for correspondence: Marcelo Luiz Lima Brandão, INCQS/Fiocruz—Immunology, Av. Brasil, 4365 Manguinhos, Rio de Janeiro, RJ, CEP 21040-900, Brazil; email: marcelo.brandao@incqs.fiocruz.br

Introduction of Eurasian-Origin Influenza A(H8N4) Virus into North America by Migratory Birds

Andrew M. Ramey, Andrew B. Reeves, Tyrone Donnelly, Rebecca L. Poulson, David E. Stallknecht

Author affiliations: US Geological Survey Alaska Science Center, Anchorage, Alaska, USA (A.M. Ramey, A.B. Reeves, T. Donnelly); University of Georgia, Athens, Georgia, USA (R.L. Poulson, D.E. Stallknecht)

DOI: <https://doi.org/10.3201/eid2410.180447>

We identified a Eurasian-origin influenza A(H8N4) virus in North America by sampling wild birds in western Alaska, USA. Evidence for repeated introductions of influenza A viruses into North America by migratory birds suggests that intercontinental dispersal might not be exceedingly rare and that our understanding of viral establishment is incomplete.

Research of and surveillance for influenza A viruses in wild birds inhabiting western Alaska have consistently provided support for the exchange of viruses between East Asia and North America via Beringia (1,2). Sampling of wild birds inhabiting Izembek National Wildlife Refuge (NWR) and surrounding areas in Alaska (≈55°N, 163°W) conducted during 2011–2015 has been used in recent research to identify the dispersal of influenza A(H9N2) viruses among China, South Korea, and Alaska (3); provide inference about the evolutionary pathways of economically important foreign-origin poultry pathogens introduced into North America (4); and identify sampling efficiencies for optimizing the detection of evidence for intercontinental virus exchange (5).

During September–October 2016, we collected 541 combined oral-pharyngeal and cloacal swab samples from hunter-harvested waterfowl (*Anseriformes* spp.) and 401 environmental fecal samples from monospecific flocks of either emperor geese (*Chen canagica*) or glaucous-winged gulls (*Larus glaucescens*) within and around Izembek NWR. Samples were deposited into viral transport media, placed in dry shippers charged with liquid nitrogen within 24 h, shipped, and stored frozen at –80°C before laboratory analysis. We screened samples for the influenza A virus matrix gene and subjected them to virus isolation; resultant isolates were genomically sequenced in accordance with previously reported methods (5). A total of 116 samples tested positive for the matrix gene, and 38 isolates were recovered of the following combined subtypes: H1N2, H3N2, H3N2/N6 (mixed infection), H3N8, H4N6, H5N2, H6N2, H7N3,

H8N4, and H12N2. We selected the single H8N4 isolate, A/northern pintail/Alaska/UGA116-3997/2016(H8N4) (GenBank accession nos. MG976689–96), for genomic characterization as part of this investigation.

We queried sequence information for the complete coding region of each gene segment of A/northern pintail/Alaska/UGA116-3997/2016(H8N4) against the GenBank database to identify strains sharing $\geq 99\%$ nt identity. We then reconstructed maximum-likelihood phylogenetic trees for each gene segment in MEGA 7.0.21 (<https://www.megasoftware.net/>) by incorporating sequence information for representative reference sequences from avian-origin influenza A virus isolates from Eurasia and North America using the general time-reversible plus invariant sites (G+I) model with 1,000 bootstrap replications.

Gene segments for A/northern pintail/Alaska/UGA116-3997/2016(H8N4), isolated from a sample collected from a hunter-harvested duck on September 6, 2016, shared $\geq 99\%$ nt identity to those of ≥ 1 isolates recovered from wild and domestic birds sampled in East Asia during 2006–2016 (online Technical Appendix Table, <https://wwwnc.cdc.gov/EID/article/24/10/18-0447-Techap1.pdf>). This isolate also shared $\geq 99\%$ nt identity with 1–4 isolates recovered from wild bird samples collected at Izembek NWR during 2012–2015 at the polymerase acidic and polymerase basic 2 gene segments (online Technical Appendix Table). A/northern pintail/Alaska/UGA116-3997/2016(H8N4) did not, however, share $\geq 99\%$ nt identity at all 8 gene segments with any other influenza A virus isolate for which genomic information was available, indicating that this H8N4 isolate might

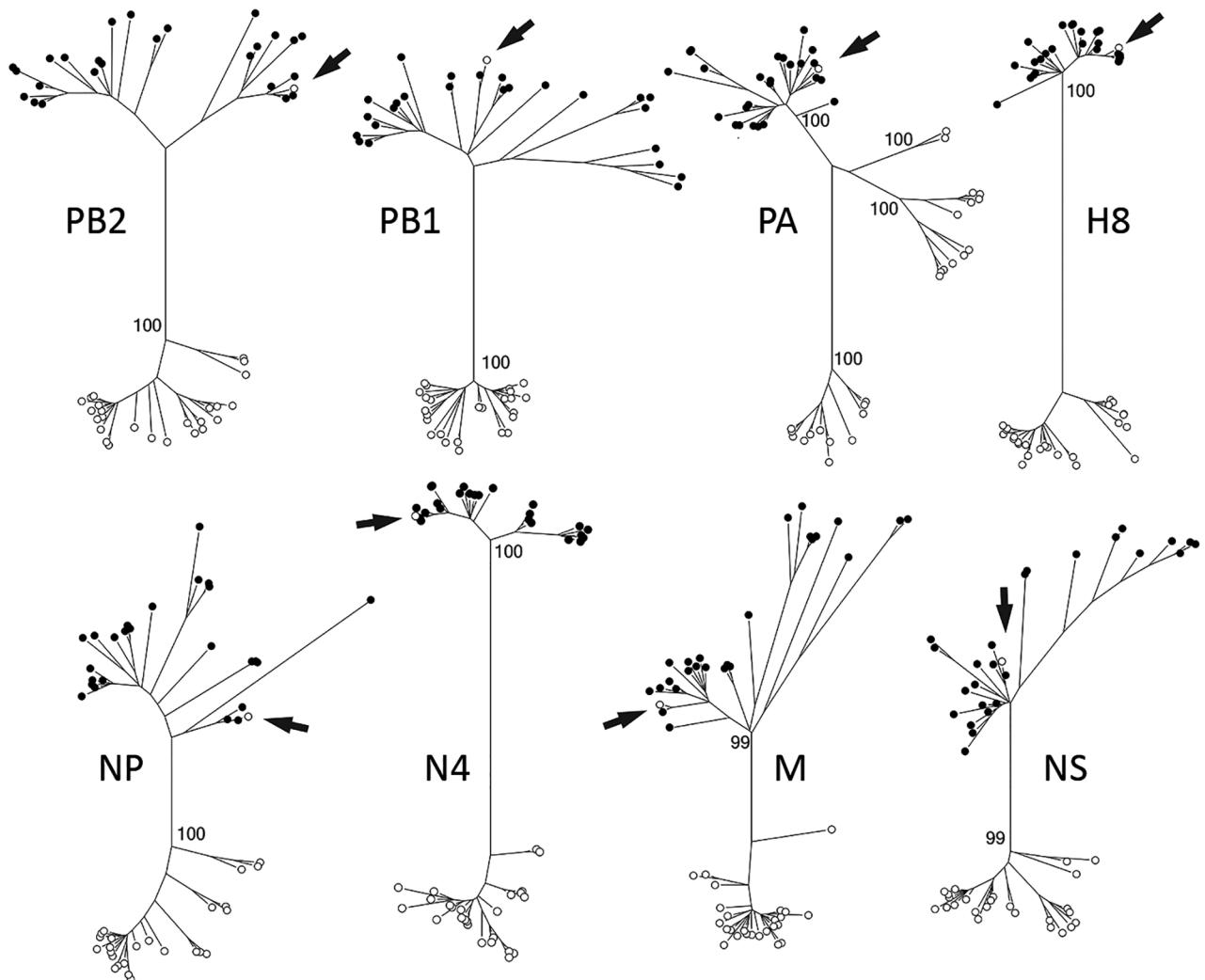


Figure. Maximum-likelihood phylogenetic trees showing inferred relationships among nucleotide sequences for the complete coding regions of gene segments for influenza A virus strain A/northern pintail/Alaska/UGA116-3997/2016(H8N4) (white circle indicated with an arrow) and reference sequences from viruses isolated from birds in Eurasia (black circles) and North America (white circles). Bootstrap support values for continentally affiliated clades are shown. Phylogenetic trees with complete strain names as tip labels are provided in the online Technical Appendix Figure (<https://wwwnc.cdc.gov/EID/article/24/10/18-0447-Techap1.pdf>). H, hemagglutinin; M, matrix; N, neuraminidase; NP, nucleoprotein; NS, nonstructural; PA, polymerase acidic; PB, polymerase basic.

represent a previously unidentified or unreported genome constellation (online Technical Appendix Table).

Phylogenetic analyses strongly supported structuring of tree topologies into major clades by continental affiliation of reference sequences (bootstrap values ≥ 99 ; online Technical Appendix Figure). Sequence information for all 8 gene segments of A/northern pintail/Alaska/UGA116-3997/2016(H8N4) clustered within clades composed of reference sequences for influenza A viruses originating from samples collected in Eurasia (Figure; online Technical Appendix Figure). Therefore, phylogenetic analyses provided support for Eurasian ancestry of this genomic constellation. We inferred our results to provide evidence for the introduction of this foreign-origin H8N4 virus into North America by migratory birds given previous support for intercontinental viral dispersal derived through genetic characterization of avian influenza A viruses originating from western Alaska (1–3,5), the intercontinental migratory tendencies of northern pintails (6,7) and other species inhabiting Izembek NWR at the time of sampling (8), the paucity of domestic poultry in this region, and the proximity of Izembek NWR to East Asia.

During 2010–2016, research and surveillance for influenza A viruses in wild birds inhabiting North America have provided evidence for the intercontinental dispersal of the following 4 viral genome constellations between Eurasia and North America: H16N3 (9), H9N2 (3), highly pathogenic clade 2.3.4.4 H5N8 (10), and H8N4 (this study). Four reports of independent purported intercontinental dispersal events for influenza A viruses via migratory birds during 7 years of sampling do not disprove the paradigm of restricted viral dispersal between Eurasia and North America. However, repeated detections of these viruses crossing the Bering Strait (3,10; this study) suggest that viral dispersal between East Asia and North America might not be exceedingly rare. Thus, a lack of selective advantage for comparatively rare foreign-origin influenza A viruses, purifying selection for endemic viruses, or both might be important mechanisms regulating the establishment of these viruses within the wild bird reservoir. Therefore, additional research directed toward understanding selection pressures regulating the establishment of these viruses might provide useful inference for informing surveillance and response activities for economically costly or potentially pandemic foreign-origin viruses in wild birds inhabiting North America.

Acknowledgments

We thank G. Risdahl, L. Melendez, and other US Fish and Wildlife Service staff at Izembek National Wildlife Refuge for logistical support. We appreciate laboratory support provided by N. Davis-Fields and C. Kienzle. We thank G. Hilderbrand, M. Wille, and 2 anonymous reviewers for constructive feedback on previous versions of this manuscript.

This work was funded by the US Geological Survey through the Wildlife Program of the Ecosystems Mission area and by the National Institute of Allergy and Infectious Diseases, National Institutes of Health, under contract HHSN272201400006C.

Data that support the findings of this publication can be found at <https://doi.org/10.5066/F7JD4W2W>.

About the Author

Dr. Ramey is a research scientist at the US Geological Survey Alaska Science Center, Anchorage, Alaska. His primary research interests include the maintenance and dispersal of infectious agents by wildlife.

References

1. Ramey AM, Pearce JM, Flint PL, Ip HS, Derksen DV, Franson JC, et al. Intercontinental reassortment and genomic variation of low pathogenic avian influenza viruses isolated from northern pintails (*Anas acuta*) in Alaska: examining the evidence through space and time. *Virology*. 2010;401:179–89. <http://dx.doi.org/10.1016/j.virol.2010.02.006>
2. Reeves AB, Pearce JM, Ramey AM, Ely CR, Schmutz JA, Flint PL, et al. Genomic analysis of avian influenza viruses from waterfowl in western Alaska, USA. *J Wildl Dis*. 2013;49:600–10. <http://dx.doi.org/10.7589/2012-04-108>
3. Ramey AM, Reeves AB, Sonsthagen SA, TeSlaa JL, Nashold S, Donnelly T, et al. Dispersal of H9N2 influenza A viruses between East Asia and North America by wild birds. *Virology*. 2015;482:79–83. <http://dx.doi.org/10.1016/j.virol.2015.03.028>
4. Ramey AM, Reeves AB, TeSlaa JL, Nashold S, Donnelly T, Bahl J, et al. Evidence for common ancestry among viruses isolated from wild birds in Beringia and highly pathogenic intercontinental reassortant H5N1 and H5N2 influenza A viruses. *Infect Genet Evol*. 2016;40:176–85. <http://dx.doi.org/10.1016/j.meegid.2016.02.035>
5. Reeves AB, Hall JS, Poulson RL, Donnelly T, Stallknecht DE, Ramey AM. Influenza A virus recovery, diversity, and intercontinental exchange: a multi-year assessment of wild bird sampling at Izembek National Wildlife Refuge, Alaska. *PLoS One*. 2018;13:e0195327. <http://dx.doi.org/10.1371/journal.pone.0195327>
6. Miller MR, Takekawa JY, Fleskes JP, Orthmeyer DL, Casazza ML, Perry WM. Spring migration of northern pintails from California's Central Valley wintering area tracked with satellite telemetry: routes, timing, and destinations. *Canadian Journal of Zoology*. 2005;83:1314–32. <http://dx.doi.org/10.1139/z05-125>
7. Hupp JW, Yamaguchi N, Flint PL, Pearce JM, Tokita K, Shimada T, et al. Variation in spring migration routes and breeding distribution of northern pintails *Anas acuta* that winter in Japan. *J Avian Biol*. 2011;42:289–300. <http://dx.doi.org/10.1111/j.1600-048X.2011.05320.x>
8. Hupp JW, Schmutz JA, Ely CR, Syroechkovskiy EE Jr, Kondratyev AV, Eldridge WD, et al. Molt migration of emperor geese *Chen canagica* between Alaska and Russia. *J Avian Biol*. 2007;38:462–70. <http://dx.doi.org/10.1111/j.0908-8857.2007.03969.x>
9. Huang Y, Wille M, Benkaroun J, Munro H, Bond AL, Fifield DA, et al. Perpetuation and reassortment of gull influenza A viruses in Atlantic North America. *Virology*. 2014;456-457:353–63. <http://dx.doi.org/10.1016/j.virol.2014.04.009>
10. Lee DH, Torchetti MK, Winker K, Ip HS, Song CS, Swayne DE. Intercontinental spread of Asian-origin H5N8 to North America through Beringia by migratory birds. *J Virol*. 2015;89:6521–4. <http://dx.doi.org/10.1128/JVI.00728-15>

Address for correspondence: Andrew M. Ramey, US Geological Survey Alaska Science Center, 4210 University Dr, Anchorage, AK 99508, USA; email: aramey@usgs.gov

New Reassortant Clade 2.3.4.4b Avian Influenza A(H5N6) Virus in Wild Birds, South Korea, 2017–2018

Jung-Hoon Kwon,¹ Sol Jeong,¹ Dong-Hun Lee,
David E. Swayne, Yu-jin Kim, Sun-hak Lee,
Jin-Yong Noh, Tseren-Ochir Erdene-Ochir,
Jei-Hyun Jeong, Chang-Seon Song

Author affiliations: Konkuk University, Seoul, South Korea (J.-H. Kwon, S. Jeong, Y.-J. Kim, S.-H. Lee, J.-Y. Noh, T.-O. Erdene-Ochir, J.-H. Jeong, C.-S. Song); US Department of Agriculture, Athens, Georgia, USA (D.-H. Lee, D.E. Swayne)

DOI: <https://doi.org/10.3201/eid2410.180461>

We isolated new reassortant avian influenza A(H5N6) viruses from feces of wild waterfowl in South Korea during 2017–18. Phylogenetic analysis suggested that reassortment occurred between clade 2.3.4.4b H5N8 and Eurasian low pathogenicity avian influenza viruses circulating in wild birds. Dissemination to South Korea during the 2017 fall migratory season followed.

Clade 2.3.4.4 H5 highly pathogenic avian influenza viruses (HPAIVs) have evolved by reassortment with different neuraminidase (NA) and internal genes of prevailing low pathogenicity avian influenza viruses (LPAIVs) and other HPAIVs to generate new genotypes and further evolved into genetic subgroups A–D since 2014 (1). Among these, subgroups A and B viruses were disseminated over vast geographic regions by migratory wild birds (2,3). Subgroup B influenza A(H5N8) viruses were detected in Qinghai Lake, China, and Uvs-Nuur Lake, Russia, during May–June 2016 (Qinghai/Uvs-like), followed by the identification of reassortant viruses in multiple Eurasian countries (4–6). Recently, subgroup B H5N6 viruses were isolated from birds in Greece during February 2017 and England, Germany, the Netherlands, Japan, and Taiwan during winter 2017–18 (7,8).

¹These authors contributed equally to this article.

During December 2017–January 2018 in South Korea, we isolated 6 H5N6 HPAIVs from 231 fecal samples of wild birds collected from the banks of the Cheongmi-cheon River (37°06'56.9"N, 127°25'18.3"E) and 34 from 222 fecal samples collected from the banks of the Gokgyo-cheon River (36°45'12.3"N, 127°07'12.7"E) (online Technical Appendix 1, <https://wwwnc.cdc.gov/EID/article/24/10/18-0461-Techapp1.pdf>). These wild bird habitats are wintering sites of migratory waterfowl, including mallard (*Anas platyrhynchos*), spot-billed duck (*Anas poecilorhyncha*), Mandarin duck (*Aix galericulata*), and common teal (*Anas crecca*). The Gokgyo-cheon River is a major habitat site for Mandarin ducks, and numerous HPAIVs were detected in fecal samples from Mandarin ducks during 2011, 2015, and 2016 (9). We identified avian influenza virus–positive fecal samples from 38 Mandarin ducks and 2 mallards, based on DNA barcoding technique (10). We performed full-length genome sequencing and comparative phylogenetic analysis on 19 of the 40 isolates (online Technical Appendix 1; online Technical Appendix 2, <https://wwwnc.cdc.gov/EID/article/24/10/18-0461-Techapp2.xlsx>).

All H5N6 isolates shared high nucleotide sequence identities in all 8 gene segments (99.58%–100%) and were identified as HPAIVs based on the presence of multiple basic amino acids at the HA proteolytic cleavage site (PLREKRRKR/G). Searches of the GISAID (<https://www.gisaid.org>) and BLAST (<https://blast.ncbi.nlm.nih.gov/Blast.cgi>) databases indicated that all 8 genomes had the highest nucleotide identity with A/Great_Black-backed_Gull/Netherlands/1/2017 (Netherlands/1) clade 2.3.4.4 subgroup B H5N6 strain from December 2017 (99.17%–99.79%), rather than subgroup B H5N6 viruses from Japan and Taiwan collected during December 2017 (97.18%–99.27%).

In phylogenetic analysis, we identified 2 genotypes of subgroup B H5N6 viruses (online Technical Appendix 1 Figures 1, 2): genotypes B.N6.1 and B.N6.2. The genotype B.N6.1 viruses were identified from South Korea, Japan, Taiwan, Greece, and the Netherlands (Netherlands/1 strain), and the genotype B.N6.2 viruses were detected from England, Germany, and the Netherlands. For genotype B.N6.1, all genes except NA clustered with H5N8 HPAIV of previously reported genotypes, H5N8-NL cluster I in the Netherlands (6), Ger-11-16 in Germany (5), and Duck/Poland/82a/16-like in Italy (4). The NA gene clustered with LPAIVs circulating in wild birds in Eurasia and separated into 2 clusters, suggesting the potential for >2 independent reassortment events between H5N8 virus and unidentified wild bird origin N6 segments. Consistent clustering of South Korea isolates with the Netherlands/1 strain in maximum-likelihood (ML) phylogenies for each gene supported by high ML bootstrap values (86–100) suggests their close relationship. The genotype B.N6.2 viruses