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## References

- Normand A-C, Cassagne C, Gautier M, Becker P, Ranque S, Hendrickx M, et al. Decision criteria for MALDI-TOF MS-based identification of filamentous fungi using commercial and in-house reference databases. BMC Microbiol. 2017;17:25. http://dx.doi.org/ 10.1186/s12866-017-0937-2
- Bhandary S, Karki P, Sinha BK. Malignant otitis externa: a review. Pac Health Dialog. 2002;9:64–7.
- Parize P, Chandesris M-O, Lanternier F, Poirée S, Viard J-P, Bienvenu B, et al. Antifungal therapy of *Aspergillus* invasive otitis externa: efficacy of voriconazole and review. Antimicrob Agents Chemother. 2009;53:1048–53. http://dx.doi.org/10.1128/ AAC.01220-08
- Gabrielli E, Fothergill AW, Brescini L, Sutton DA, Marchionni E, Orsetti E, et al. Osteomyelitis caused by *Aspergillus* species: a review of 310 reported cases. Clin Microbiol Infect. 2014;20: 559–65. http://dx.doi.org/10.1111/1469-0691.12389
- Nawas MT, Daruwalla VJ, Spirer D, Micco AG, Nemeth AJ. Complicated necrotizing otitis externa. Am J Otolaryngol. 2013;34:706–9. http://dx.doi.org/10.1016/j.amjoto.2013.07.003
- Matt BH, Lusk RP. Delineation of a deep neck abscess with magnetic resonance imaging. Ann Otol Rhinol Laryngol. 1987;96: 615–7. http://dx.doi.org/10.1177/000348948709600526
- Aribandi M, McCoy VA, Bazan C III. Imaging features of invasive and noninvasive fungal sinusitis: a review. Radiographics. 2007;27:1283–96. http://dx.doi.org/10.1148/rg.275065189
- Jain V, Makwana GE, Bahri N, Mathur MK. The "dot in circle" sign on MRI in maduramycosis: a characteristic finding. J Clin Imaging Sci. 2012;2:66–66. http://dx.doi.org/10.4103/ 2156-7514.103056
- Yamada K, Zoarski GH, Rothman MI, Zagardo MT, Nishimura T, Sun CCJ. An intracranial aspergilloma with low signal on T2-weighted images corresponding to iron accumulation. Neuroradiology. 2001;43:559–61. http://dx.doi.org/10.1007/ s002340000535
- Stodulski D, Kowalska B, Stankiewicz C. Otogenic skull base osteomyelitis caused by invasive fungal infection. Eur Arch Otorhinolaryngol. 2006;263:1070–6. http://dx.doi.org/10.1007/ s00405-006-0118-7

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# Epizootic Hemorrhagic Disease in White-Tailed Deer, Canada

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Epizootic hemorrhagic disease affects wild and domestic ruminants and has recently spread northward within the United States. In September 2017, we detected epizootic hemorrhagic disease virus in wild white-tailed deer, *Odocoileus virginianus*, in east-central Canada. *Culicoides* spp. midges of the subgenus *Avaritia* were the most common potential vectors identified on site.

Epizootic hemorrhagic disease viruses (EHDVs) and bluetongue viruses (BTVs) are *Culicoides* spp. midge– transmitted orbiviruses that represent an imminent threat to the health of ruminant livestock and wildlife. For susceptible ruminants, EHDV and BTV infections can result in high rates of illness and death, leading to severe economic hardship to the agricultural sector (1). These viruses have a historical geographic range of  $40^{\circ}N-50^{\circ}N$  and  $35^{\circ}S$ , following the distribution of the *Culicoides* vectors. However, the epidemiology of these pathogens is changing, with decades of northward expansion into areas of Europe and North America with immunologically naive hosts (1–3).

In Canada, EHDV has rarely and sporadically been detected in the southern portions of British Columbia, Alberta, and Saskatchewan (4). We report the detection of EHDV in white-tailed deer, *Odocoileus virginianus*, in east-central Canada, providing further evidence of the northern range expansion of orbiviruses within North America.

On September 7, 2017, two wild white-tailed deer carcasses were found in a seminatural area adjacent to a

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river and ravine in London, Ontario, Canada (42.9849°N, 81.2453°W). The carcasses were submitted for diagnostic investigation to the Ontario-Nunavut region of the Canadian Wildlife Health Cooperative. Both deer had gross lesions consistent with epizootic hemorrhagic disease, including multiorgan petechial and ecchymotic hemorrhages on serosal surfaces (5). Spleen, lung, and liver samples from both deer were submitted to the National Centre for Foreign Animal Diseases in Winnipeg. All samples were positive for EHDV genomes by reverse transcription PCR (RT-PCR) (6). The serotype of the virus was identified by serotype-specific conventional RT-PCR followed by Sanger sequencing. EHDV-2 was isolated from spleen samples of both deer. Two additional deer carcasses were identified in the area but were not submitted for diagnostic testing. Tissues from an additional 14 deer carcasses that originated in southern Ontario during September 23-November 28, 2017, showed negative results for EHDV by RT-PCR, as part of enhanced surveillance conducted by the Ontario Ministry of Natural Resources and Forestry and the Canadian Wildlife Health Cooperative.

In addition, we conducted serosurveillance across southern Ontario in the fall of 2017 with samples collected by private veterinarians. All live animal samples were collected by using protocols approved by the Animal Care Committee, University of Guelph (approval no. 3529). Cattle were from Chatham-Kent County, adjacent to Middlesex County, and were born and had lived their whole lives in Canada. Results revealed 15 cattle with antibodies to EHDV-2 by ELISA and serum neutralization tests (S.E. Allen, unpub. data).

We also conducted a targeted survey for adult *Culicoides* midges in the area of the white-tailed deer deaths by using four 2770 UV LED CDC light traps (BioQuip Products, http://www.bioquip.com) for 16 trap-nights during September 9–13, 2017. We taxonomically identified 31 individual *Culicoides* spp. midges (7), including subgenus *Avaritia* (n = 22), *C. haematopotus* (n = 5), *C. stellifer* (n = 2), *C. venustus* (n = 1), and *C. crepuscularis* (n = 1) midges.

The northern expansion of EHDV and BTV in the midwesterm and northeastern regions of the United States in recent years (1,8) has increased the likelihood of incursion into eastern Canada. The EHDV occurrence in white-tailed deer in Ontario we report supports the need to maintain vigilance. Although this initial occurrence was limited spatially and temporally, future cases may be more widespread in wildlife and livestock. Furthermore, this localized EHDV-2 occurrence was within 72 km of the US border of Michigan and coincided with concurrent and widespread EHDV-2 and EHDV-6 outbreaks across much of the eastern United States during July–October, 2017 (D.E. Stallknecht, University of Georgia, pers. comm, 2018 Jan 10).

EHDV may have been introduced to Ontario through the movement of infected vectors or ruminants, and its detection in Ontario may represent the northern edge of the EHDV outbreak in the eastern United States. Alternatively, low levels of previously undetected EHDV transmission may have been transient or ongoing across a wider region, including Ontario, for an unknown period. In either case, this event may preclude a much more widespread and higher impact EHDV outbreak, such as occurred with BTV in Europe and with EHDV in the United States (2,8).

We identified numerous Culicoides spp. midges at the site where the EHDV-infected white-tailed deer carcasses were recovered. However, C. sonorensis midges, the only confirmed EHDV vector species in North America (1,9), was not among these. This species was recently identified in Ontario (10), but the density and distribution of this species in the province, as well as its role in regional EHDV transmission, is currently unknown. Culicoides spp. composition within areas of past EHDV outbreaks varies widely; therefore, additional Culicoides spp. midges may serve as competent EHDV vectors. However, this possibility has not been confirmed (1,9). In addition to vector competence, the climatic limitations for the survival and successful breeding and establishment of current Culicoides spp. populations in the region remain to be investigated (1,8). These factors indicate the need for ongoing Culicoides spp. midge and deer mortality surveillance in the region.

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#### References

- Ruder MG, Lysyk TJ, Stallknecht DE, Foil LD, Johnson DJ, Chase CC, et al. Transmission and epidemiology of bluetongue and epizootic hemorrhagic disease in North America: current perspectives, research gaps, and future directions. Vector Borne Zoonotic Dis. 2015;15:348–63. http://dx.doi.org/10.1089/ vbz.2014.1703
- Maclachlan NJ. Global implications of the recent emergence of bluetongue virus in Europe. Vet Clin North Am Food Anim Pract. 2010;26:163–71. http://dx.doi.org/10.1016/j.cvfa.2009.10.012

### RESEARCH LETTERS

- McVey DS, Drolet BS, Ruder MG, Wilson WC, Nayduch D, Pfannenstiel R, et al. Orbiviruses: a North American perspective. Vector Borne Zoonotic Dis. 2015;15:335–8. http://dx.doi.org/10.1089/vbz.2014.1699
- Pybus MJ, Ravi M, Pollock C. Epizootic hemorrhagic disease in Alberta, Canada. J Wildl Dis. 2014;50:720–2. http://dx.doi.org/ 10.7589/2014-02-024
- 5 Howerth EW, Stallknecht DE, Kirkland PD. Bluetongue, epizootic hemorrhagic disease, and other orbivirus-related diseases. In: Williams ES, Barker IK, editors. Infectious diseases of wild mammals. Ames (IA): Iowa State University Press; 2001. p. 82–9.
- Pasick J, Handel K, Zhou EM, Clavijo A, Coates J, Robinson Y, et al. Incursion of epizootic hemorrhagic disease into the Okanagan Valley, British Columbia in 1999. Can Vet J. 2001;42:207–9.
- Wirth WW, Dyce AL, Peterson BV. An atlas of wing photographs with a summary of the numerical characters of the Nearctic species of *Culicoides* (Diptera: Ceratopogonidae). Contrib Am Entomol Inst. 1985;22:1–46.
- Stallknecht DE, Allison AB, Park AW, Phillips JE, Goekjian VH, Nettles VF, et al. Apparent increase of reported hemorrhagic disease in the midwestern and northeastern USA. J Wildl Dis. 2015;51:348–61. http://dx.doi.org/10.7589/2013-12-330
- Pfannenstiel RS, Mullens BA, Ruder MG, Zurek L, Cohnstaedt LW, Nayduch D. Management of North American *Culicoides* biting midges: current knowledge and research needs. Vector Borne Zoonotic Dis. 2015;15:374–84. http://dx.doi.org/10.1089/ vbz.2014.1705
- Jewiss-Gaines A, Barelli L, Hunter FF. First records of *Culicoides* sonorensis (Diptera: Ceratopogonidae), a known vector of bluetongue virus, in southern Ontario. J Med Entomol. 2017; 54:757–62.

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# Effects of Political Instability in Venezuela on Malaria Resurgence at Ecuador-Peru Border, 2018

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Mass migration from Venezuela has increased malaria resurgence risk across South America. During 2018, migrants from Venezuela constituted 96% of imported malaria cases along the Ecuador–Peru border. *Plasmodium vivax* predominated (96%). Autochthonous malaria cases emerged in areas previously malaria-free. Heightened malaria control and a response to this humanitarian crisis are imperative.

Malaria is a vectorborne parasitic infection caused by *Plasmodium* spp. and transmitted by *Anopheles* mosquitoes, characterized by fever and hemolysis with chronic and fatal potential (1). Despite substantial strides toward elimination in the Americas, malaria remains a major concern;  $\approx$ 975,700 cases occurred and 138 million persons were at risk in 2017 (2). Most malaria cases in South America occur in the Amazon region, and *P. vivax* is more common than *P. falciparum* (3).

*P. vivax* and *P. falciparum* malaria were historically endemic to the Ecuador–Peru coastal border region. During 1990–2012, a total of 62,000 malaria cases were reported from El Oro Province, Ecuador, and 85,605 from Tumbes Region, Peru (4). Through vector control and active case surveillance and response, malaria was eliminated from El Oro Province in 2011 and Tumbes Region in 2012 (4). However, malaria cases elsewhere in Ecuador increased from 378 in 2013 (5) to 1,279 in 2017 (6). Peru and other countries in the region also reported increased malaria in 2017, indicating a major risk for reintroduction to elimination areas (2). In 2017, Venezuela alone accounted for more than half of all malaria cases in the Americas (2).

The public health sector in Venezuela is struggling with infectious disease epidemics, including malaria (7), despite a historically successful malaria control program (3). The worsening social and economic crisis has led to large-scale migration from and within Venezuela. The shortage of antimalarial drugs and lax in-country control efforts have exacerbated the situation, affecting countries throughout South America (8). Many people from Venezuela are migrating through Colombia and Ecuador to reach Peru and the southern cone of South America, stopping at various locations along the way (Figure). We report a series of imported malaria cases in migrants from Venezuela and the first autochthonous cases of malaria in the Ecuador–Peru border region since local elimination.

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