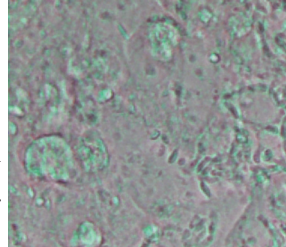


etymologia

Neospora caninum [ne-os'pə-rə ca-nin'um]

Ronnie Henry

From the *neo-* (Latin, “new”) + *spora* (Greek, “seed”) and *canis* (Latin, “dog”), *Neospora caninum* is a sporozoan parasite that was first described in 1984. It is a major pathogen of cattle and dogs but can also infect horses, goats, sheep, and deer. Antibodies to *N. caninum* have been found in humans, predominantly in those with HIV infection, although the role of this parasite in causing or exacerbating illness is unclear.



Neospora caninum, a coccidian parasite, which identified as a species in 1988. It is a major cause of spontaneous abortion in infected livestock. Image from Wikipedia ([https://en.wikipedia.org/wiki/Neospora_caninum#/media/File:Neospora_caninum_\(5256961091\).jpg](https://en.wikipedia.org/wiki/Neospora_caninum#/media/File:Neospora_caninum_(5256961091).jpg)).

Sources

1. Bjerkås I, Mohn SF, Presthus J. Unidentified cyst-forming sporozoan causing encephalomyelitis and myositis in dogs. *Z Parasitenkd.* 1984;70:271–4. <http://dx.doi.org/10.1007/BF00942230>
2. Dubey JP. Review of *Neospora caninum* and neosporosis in animals. *Korean J Parasitol.* 2003; 41:1–16. <http://dx.doi.org/10.3347/kjp.2003.41.1.1>
3. Lobato J, Silva DA, Mineo TW, Amaral JD, Segundo GR, Costa-Cruz JM, et al. Detection of immunoglobulin G antibodies to *Neospora caninum* in humans: high seropositivity rates in patients who are infected by human immunodeficiency virus or have neurological disorders. *Clin Vaccine Immunol.* 2006;13:84–9. <http://dx.doi.org/10.1128/CVI.13.1.84-89.2006>

Address for correspondence: Ronnie Henry, Centers for Disease Control and Prevention, 1600 Clifton Rd NE, Mailstop E28, Atlanta, GA 30329-4027, USA; email: boq3@cdc.gov

DOI: <https://doi.org/10.3201/eid2506.ET2506>

Reemergence of Classical Swine Fever, Japan, 2018

Alexander Postel,¹ Tatsuya Nishi,¹ Ken-ichiro Kameyama, Denise Meyer, Oliver Suckstorff, Katsuhiko Fukai, Paul Becher

Author affiliations: University of Veterinary Medicine, Hannover, Germany (A. Postel, D. Meyer, O. Suckstorff, P. Becher); National Agriculture and Food Research Organization, Tokyo, Japan (T. Nishi, K. Kameyama, K. Fukai)

DOI: <https://doi.org/10.3201/eid2506.181578>

In September 2018, classical swine fever reemerged in Japan after 26 years, affecting domestic pigs and wild boars. The causative virus belongs to the 2.1 subgenotype, which caused repeated outbreaks in eastern and Southeast Asia. Intensive surveillance of swine and vaccination of wild boars will help control and eradicate this disease in Japan.

Classical swine fever (CSF) is one of the economically most devastating diseases worldwide and is notifiable to

the World Organisation for Animal Health (OIE). The presence of CSF in a pig population results in severe restrictions on international trade of pigs and pork products. Many countries have implemented compulsory eradication programs and perform intensive surveillance. Most countries with industrialized pig production and high biosecurity standards have achieved the OIE status of being CSF free, including Japan in 2015 (1). Nevertheless, CSF is endemic to many countries that have a high number of backyard pigs. Because wild boars are as susceptible to CSF virus (CSFV) as domestic pigs, eradication of CSF in wild boars is of epidemiologic value (2).

CSFV, a positive-sense RNA virus (family *Flaviviridae*, genus *Pestivirus*) is divided into 3 major genotypes (1–3) and several subgenotypes (3,4). In Europe, the more recent outbreaks were caused by genotype 2.1 (Lithuania, 2009 and 2011) and genotype 2.3 (Latvia, 2013–2015) (5). In Asia, recent outbreaks were caused mainly by CSFV genotypes 1.1, 2.1, 2.2, and 2.3.

The spread of African swine fever (ASF) across China in 2018 has increased awareness of ASF and CSF in Southeast Asia. During August 16–September 3, 2018, at a pig farm in Gifu city, Gifu Prefecture, Japan, ~20 fattening pigs died. A veterinarian recognized that the pigs were weakened and inappetent; no clinical signs were detected before

¹These authors contributed equally to this article.

August 20. Staff from the Gifu prefectural animal hygiene service center collected and sent samples from the following animals to the National Institute of Animal Health (Tokyo, Japan) to test for ASF and CSF viruses: 6 live pigs on August 24, 1 dead pig on September 3, and 11 live pigs and 1 dead pig on September 8. The CSFV genome was detected by reverse transcription PCR and confirmed by

nucleotide sequencing. Control measures comprised culling of ≈ 600 pigs from the infected farm, movement restrictions, disinfection, epidemiologic investigations, clinical and laboratory investigations of 13 farms with epidemiologic links, and intensified surveillance. On September 13, a dead wild boar was found in the restriction zone of the initial outbreak and was CSFV positive. By March 7, 2019,

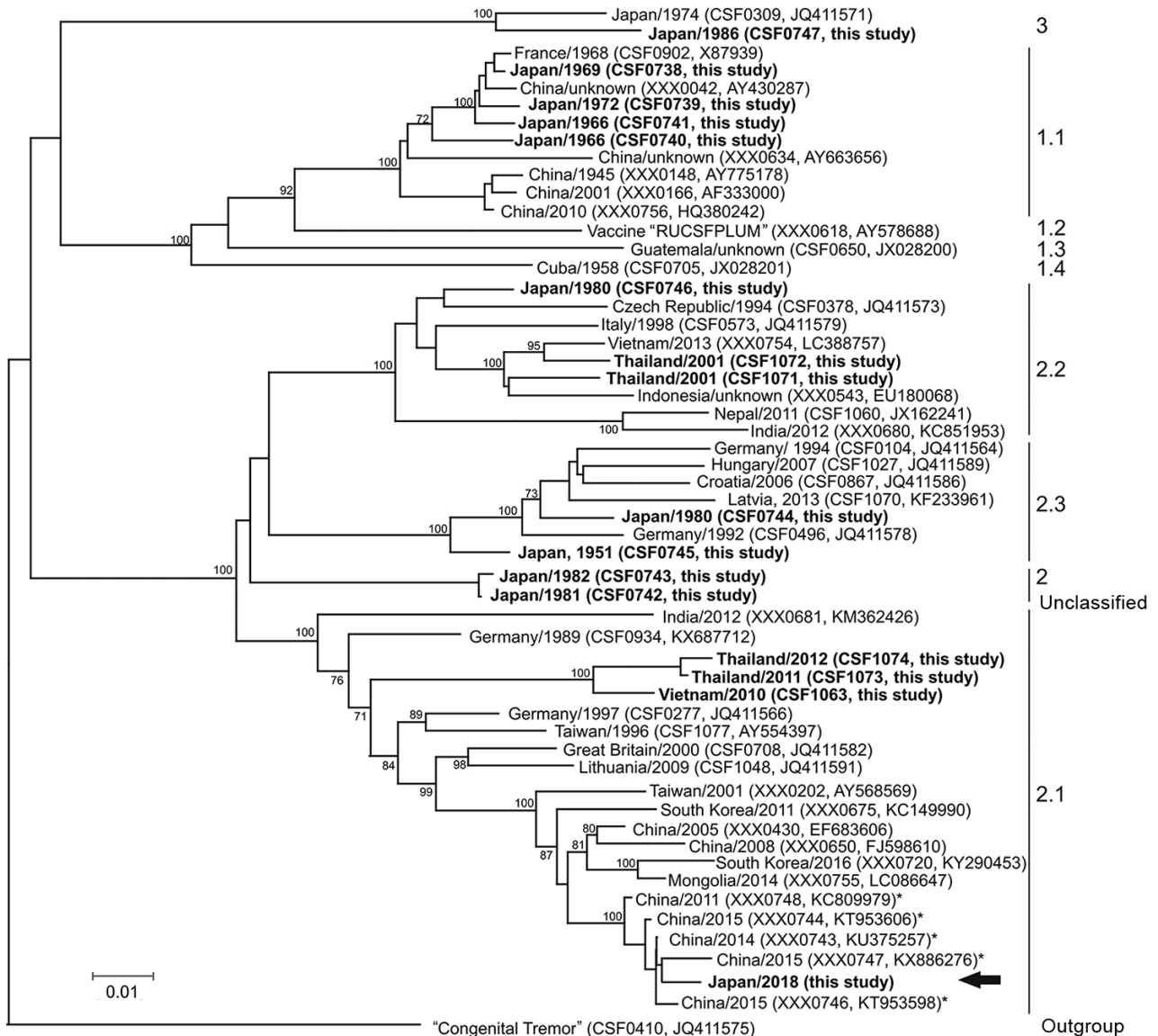


Figure. Phylogenetic tree displaying the genetic relatedness of the classical swine fever virus (CSFV) isolate obtained from the 2018 classical swine fever (CSF) outbreak in Japan to other CSFV isolates. Phylogenetic analyses were performed by using the neighbor-joining method with complete E2 (1,119 nt) sequences, including 1,000 iterations for bootstrap analysis, and were generated by the genetic typing module of the CSF database at the European Union and OIE Reference Laboratory for Classical Swine Fever (6). Only bootstrap values $\geq 70\%$ are indicated. For each isolate, country and year are given, along with catalog number of the CSF database and the GenBank accession number in parentheses. The arrow indicates the sequence of the virus isolate obtained from a domestic pig during the 2018 CSF outbreak in Japan (Japan/2018) (GenBank accession no. LC425432). Boldface indicates additional newly generated sequences (GenBank accession nos. MK026451–65). Two ancient sequences from Japan (CSF0742, CSF0743) were identified as belonging to genotype 2 with no clear affiliation to any of the 3 established subgenotypes 2.1, 2.2, and 2.3 (indicated as genotype 2, unclassified). Asterisks indicate 5 E2 sequences belonging to a group of sequences that are most closely related to the E2 sequence of the CSFV isolate Japan/2018. Scale bar indicates nucleotide substitutions per site.

a total of 68 dead and 153 live wild boars in Gifu and Aichi Prefectures had been found to be CSFV positive.

The last CSF outbreak in Japan (Kumamoto Prefecture) occurred in 1992; since 2006, vaccination against CSF has been banned. The absence of CSF in Japan for 26 years strongly suggests reintroduction of the virus from outside Japan. To support epidemiologic investigations, we performed molecular typing based on the partial 5' untranslated region (UTR) (150 nt) and on the complete E2 gene (1,119 nt) by using the CSF sequence database and the integrated tool for phylogenetic analysis (3,6). Most similar sequences identified by database search (GenBank, <https://www.ncbi.nlm.nih.gov/nucleotide>; BLAST, <https://blast.ncbi.nlm.nih.gov/Blast.cgi>) were included in the analysis together with 15 complete E2 encoding sequences (GenBank accession nos. MK026451–65) newly generated from isolates originating from Japan (10 sequences in 1951–1986), Thailand (4 sequences in 2001, 2011, and 2012), and Vietnam (1 sequence in 2010) (Figure). Phylogenetic analyses revealed that the 2018 isolate from Japan belongs to genotype 2.1; the E2 (Figure) and 5'-UTR sequences (Appendix, <https://wwwnc.cdc.gov/EID/article/25/6/18-1578-App1.pdf>) were most closely related to CSFV detected in China during 2011–2015 (98%–99% identity in E2 sequences; Figure) and China and Mongolia during 2014–2015 (98%–99% identity in partial 5'-UTR; Appendix). Subsequently, a complete genome sequence of the index isolate was determined (GenBank accession no. LC425854); the closest genetic relationship (98.9% identity) was with 2 recent isolates (GenBank accession nos. MG387217–8) from Beijing, China (7). Members of this phylogenetic clade reportedly form an emerging group of moderately virulent CSFV that is becoming more prevalent in China (8,9). Despite good availability of sequence data from China, much less information is available from other countries in the region. Therefore, similar viruses may be in other countries in eastern and Southeast Asia. Additional CSFV sequences from previous outbreaks in Japan, Thailand, and Vietnam were only distantly related to the sequence of the isolate from Japan. Partial E2 and 5'UTR sequences (GenBank accession nos. LC425434–5) obtained from the first positive wild boar (index case) revealed 100% identity to the index isolate.

Japan is among the top 10 pork-producing countries worldwide; in 2017, an estimated 16.3 million pigs were slaughtered in Japan. Presence of CSFV in wild boars remains a serious threat for domestic pigs. By February 2019, the virus had further spread from Gifu Prefecture into other prefectures in Japan, emphasizing the need for defined strategies to control the outbreak, including vaccination of wild boar, in addition to the standard policy of culling. Moreover, intensive surveillance is needed to monitor the situation carefully and will contribute to the control and eradication of CSF in Japan.

Acknowledgments

We are grateful to the regional and national authorities in Japan. We thank all providers of sequence information, diagnostic materials, and virus isolates. In particular, we thank Regional Animal Health Office No. 6, Ho Chi Minh City, Vietnam, and the National Institute of Animal Health, Bangkok, Thailand. We are thankful to the team of the European Union and OIE Reference Laboratory for CSF and all colleagues who actively support the CSF sequence database and virus bank in Hannover as well as the work of the OIE Reference Laboratories for CSF. We thank Yoshihiro Sakoda for his valuable support and discussions.

This work in part was financially supported by the Directorate-General for Health and Consumers of the European Commission and the Ministry of Agriculture, Forestry and Fisheries of Japan. The funding sources had no influence on the study design or data presented.

About the Author

Dr. Postel is a veterinarian and head of the Laboratory for Molecular Biology of the European Union and OIE Reference Laboratory for Classical Swine Fever at the Institute of Virology of the University of Veterinary Medicine in Hannover, Germany. His research interests are molecular evolution and pathogenesis of pestiviruses, characterization of novel pestivirus isolates, and diagnosis and control of classical swine fever. Dr. Nishi is a veterinarian and a researcher at the National and OIE Reference Laboratory for Classical Swine Fever at the National Institute of Animal Health, Japan. His research interests are molecular characterization, diagnosis, and control of transboundary infectious disease viruses.

References

1. Postel A, Austermann-Busch S, Petrov A, Moennig V, Becher P. Epidemiology, diagnosis and control of classical swine fever: recent developments and future challenges. *Transbound Emerg Dis*. 2017; 2018;65(Suppl 1):248–61.
2. Moennig V. The control of classical swine fever in wild boar. *Front Microbiol*. 2015;6:1211. <http://dx.doi.org/10.3389/fmicb.2015.01211>
3. Postel A, Schmeiser S, Bernau J, Meindl-Boehmer A, Pridotkas G, Dirbakova Z, et al. Improved strategy for phylogenetic analysis of classical swine fever virus based on full-length E2 encoding sequences. *Vet Res (Faisalabad)*. 2012;43:50. <http://dx.doi.org/10.1186/1297-9716-43-50>
4. Lowings P, Ibata G, Needham J, Paton D. Classical swine fever virus diversity and evolution. *J Gen Virol*. 1996;77:1311–21. <http://dx.doi.org/10.1099/0022-1317-77-6-1311>
5. Postel A, Moennig V, Becher P. Classical swine fever in Europe—the current situation. *Berl Munch Tierarztl Wochenschr*. 2013;126:468–75.
6. Postel A, Schmeiser S, Zimmermann B, Becher P. The European classical swine fever virus database: blueprint for a pathogen-specific sequence database with integrated sequence analysis tools. *Viruses*. 2016;8: pii: E302. <http://dx.doi.org/10.3390/v8110302>
7. Nishi T, Kameyama KI, Kato T, Fukai K. Genome sequence of a classical swine fever virus of subgenotype 2.1, isolated from a pig

in Japan in 2018. *Microbiol Resour Announc*. 2019;8:pii: e01362-18. <http://dx.doi.org/10.1128/MRA.01362-18>

8. Hu D, Lv L, Gu J, Chen T, Xiao Y, Liu S. Genetic diversity and positive selection analysis of classical swine fever virus envelope protein gene E2 in east China under C-strain vaccination. *Front Microbiol*. 2016;7:85. <http://dx.doi.org/10.3389/fmicb.2016.00085>
9. Luo Y, Ji S, Liu Y, Lei JL, Xia SL, Wang Y, et al. Isolation and characterization of a moderately virulent classical swine fever virus emerging in China. *Transbound Emerg Dis*. 2017;64:1848–57. <http://dx.doi.org/10.1111/tbed.12581>

Address for correspondence: Paul Becher, EU and OIE Reference Laboratory for Classical Swine Fever, Institute for Virology, University of Veterinary Medicine, Buenteweg 17, 30559 Hannover, Germany; email: paul.becher@tiho-hannover.de; Katsuhiko Fukai, National and OIE Reference Laboratory for Classical Swine Fever, National Institute of Animal Health, National Agriculture and Food Research Organization, 6-20-1, Josui-honcho, Kodaira, Tokyo 187-0022, Japan; email: fukai@affrc.go.jp

African Swine Fever Virus in Pork Brought into South Korea by Travelers from China, August 2018

Hyun-Joo Kim, Min-Jung Lee, Soo-Kyoung Lee, Da-young Kim, Sang-Ji Seo, Hae-Eun Kang, Hyang-Mi Nam

Author affiliation: Animal and Plant Quarantine Agency, Gimcheon, South Korea

DOI: <https://doi.org/10.3201/eid2506.181684>

We tested samples of pork products confiscated from travelers to South Korea for African swine fever virus (ASFV). We detected ASFV in 4 food items confiscated from travelers from Shenyang, China, in August 2018. Surveillance of pork products at country entry points is needed to mitigate the risk for ASFV introduction.

African swine fever (ASF) is a fatal viral disease that affects pigs of all ages and breeds. ASF virus (ASFV) is highly virulent and remains a global threat because of the lack of a vaccine and the ability of the virus to survive in various environmental conditions. Since 2007, ASFV has been spreading across Europe and Russia. In August 2018,

China reported the first outbreak of ASF in Asia (1). Since then, ASFV has been reported in numerous provinces and continues to spread across China (2).

Although ASF has never occurred in the Republic of Korea (hereafter referred to as South Korea), ASFV could be introduced into this country through various routes. The risk for ASF introduction into South Korea increases with the continuous spread of the disease across China. Pork products contaminated with ASFV are among the main risk factors for spreading the disease (3). Hence, since 2015, we have been conducting surveillance on pork products confiscated at airports and ports from travelers coming from countries affected by ASF. Since the program started in 2015, an average of 6,200 products have been seized per month, and we tested an average of 10 (0.16%) products per month.

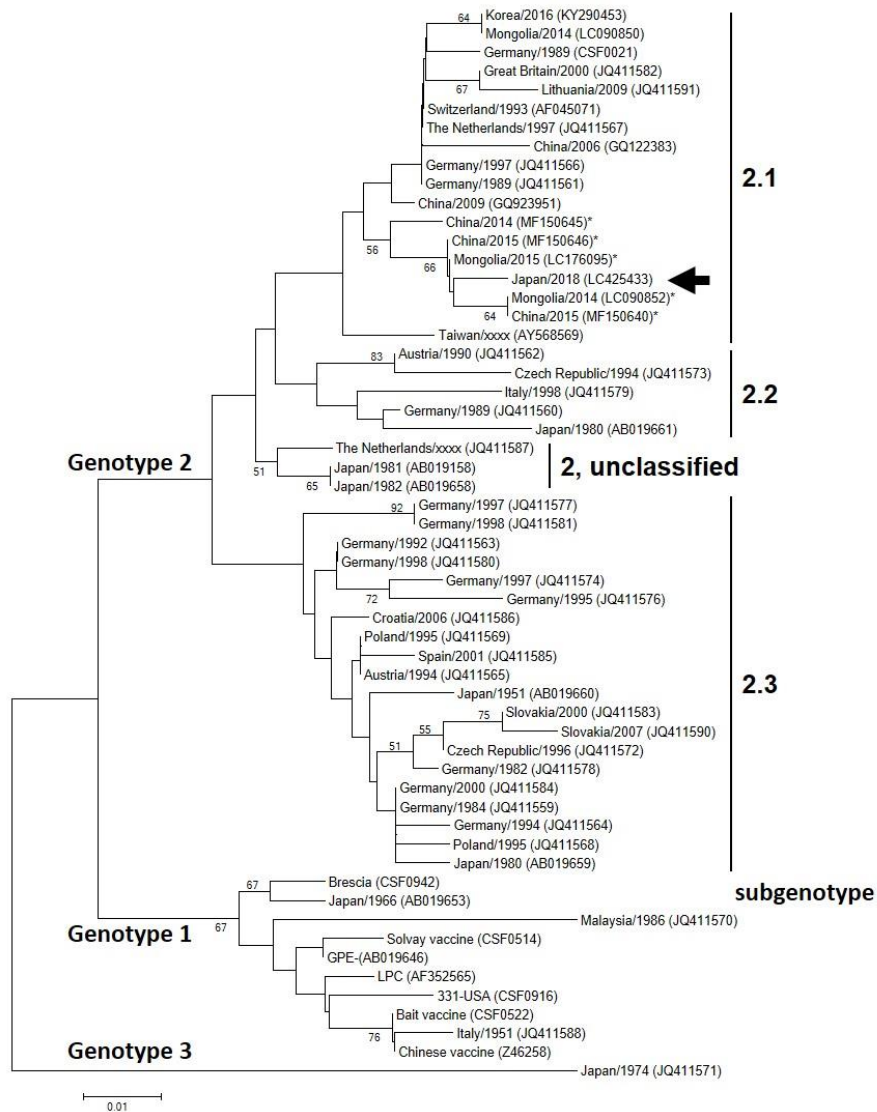
After the first ASF outbreak in China, South Korea enhanced quarantine inspections of travelers, especially those coming from China. A total of 4,064 pork products (3,374 sausages, 12 hams, and 678 other products containing pork) were seized from travelers from China in August 2018. Among these products, we randomly selected samples from 52 (1.28%) products for real-time PCR testing. We homogenized these samples and extracted nucleic acids using High Pure PCR Template Preparation Kit (Roche, <https://www.roche.com>) in a Biosafety Level 3 laboratory at the Animal and Plant Quarantine Agency in Gimcheon, South Korea. We used ASFV OIRT88/3 virus as a positive control. To amplify the ASFV *B646L* gene, we performed TaqMan real-time PCR (Applied Biosystems, <https://www.thermofisher.com>) as recommended (4).

In total, 4 samples from China tested positive for ASFV: 2 blood sausages (identification [ID] no. 18083111, seized August 16, 2018, and ID no. 18081148, seized August 20, 2018), 1 dumpling (ID no. 18082721, seized August 18, 2018), and 1 commercial sausage product (ID no. JI 18080406, seized August 26, 2018). All ASFV-positive samples were from products seized at the Incheon and Jeju International Airports from passengers flying from Shenyang, China, where the first ASFV outbreak in China was reported.

We performed conventional PCR to further analyze the ASFV isolates detected. We amplified 3 independent regions of the ASFV genome: the *B646L* gene encoding p72, the *E183L* gene encoding p54, and a tandem-repeat sequence located between the *I73R* and *I329L* genes (5–7). All genes detected were ASFV genotype II (Figure). All positive samples had an intergenic region II variant with an additional tandem-repeat sequence (5'-GGAATATATA-3') between the *I73R* and *I329L* genes (5). The intergenic region II variant we observed was identical to those reported in isolates Ukr12/Zapo, Bel13/Grodno, Lt14/1490, Lt14/1482, Pol14/Sz, and Pol14/Krus (6). The same tandem-repeat sequence insertion was also observed in China isolates ASFV SY18 and CN201801 (1,2).

Reemergence of Classical Swine Fever, Japan, 2018

Appendix



Appendix Figure. Phylogenetic tree based on partial 5'UTR sequences (150 nt). Analysis was performed by the neighbor-joining method including 1,000 iterations. Only bootstrap values $\geq 50\%$ are indicated. For each isolate, country and year are given (GenBank accession number in paranthesis). The sequence of the virus isolate obtained from a domestic pig during the outbreak of CSF in Japan in 2018 (Japan/2018) is indicated by an arrow (GenBank LC425433). Two ancient sequences from Japan

(CSF0742, CSF0743) together with an ancient sequence from The Netherlands (JQ411587) were identified to belong to genotype 2, without clear affiliation to any of the three established subgenotypes 2.1, 2.2 and 2.3 (indicated as genotype 2, unclassified). Five sequences belonging to a group of sequences that are most closely related to the partial 5'UTR sequence of the CSFV isolate Japan/2018 (BLAST search, GenBank) are indicated by asterisks. In addition to three sequences from China, these comprise two sequences from Mongolia (Mongolia/2014 (LC090852) and Mongolia/2015 (LC176095) for which complete E2 encoding sequences were not available.