

7. Azevedo MC, Ramuno NM, Fachin LR, Tassa M, Rosa PS, Belone AF, et al. qPCR detection of *Mycobacterium leprae* in biopsies and slit skin smear of different leprosy clinical forms. *Braz J Infect Dis*. 2017;21:71–8. <https://doi.org/10.1016/j.bjid.2016.09.017>
8. Lugton I. Mucosa-associated lymphoid tissues as sites for uptake, carriage and excretion of tubercle bacilli and other pathogenic mycobacteria. *Immunol Cell Biol*. 1999;77:364–72. <https://doi.org/10.1046/j.1440-1711.1999.00836.x>

Address for correspondence: Marilda A.M. Morgado de Abreu, rua São Paulo, 1949, centro, CEP 17900-000, Dracena, SP, Brazil; email: marilda@morgadoabreu.com.br

Fatal *Chlamydia avium* Infection in Captive Picazuro Pigeons, the Netherlands

Marja Kik, Marloes Heijne, Jooske IJzer, Guy Grinwis, Yvonne Pannekoek, Andrea Gröne

Author affiliations: Utrecht University, Utrecht, the Netherlands (M. Kik, J. IJzer, G. Grinwis, A. Gröne); Wageningen Bioveterinary Research, Lelystad, the Netherlands (M. Heijne); University of Amsterdam, Amsterdam, the Netherlands (Y. Pannekoek)

DOI: <https://doi.org/10.3201/eid2610.191412>

In 2016, an outbreak of *Chlamydia avium* infection occurred among Picazuro pigeons (*Patagioenas picazuro*) living in an aviary in the Netherlands. Molecular typing revealed a unique strain of *C. avium*. Our findings show that *C. avium* infection, which usually causes subclinical infection, can cause fatal disease in pigeons.

Until approximately 2014, *Chlamydia psittaci* was the only *Chlamydia* species detected in birds. Researchers have catalogued ≈465 bird species affected by this pathogen, which mainly causes subclinical infections but sometimes results in acute disease and death (1). In humans, *C. psittaci* is highly infectious and can cause severe pneumonia. *Chlamydia* bacteria, which are present in (dried) excreta or feather dust, are transmitted through direct contact or inhalation. In 2014, researchers proposed 2 new members of *Chlamydiaceae*: *C. avium* and *C. gallinacea* (2). *C. avium* affects pigeons and psittacine birds, whereas

C. gallinacea affects poultry. Most *C. avium* and *C. gallinacea* infections in birds are subclinical, and the zoonotic potential of these species is unknown (3).

In 2016, an outbreak of *C. avium* infection occurred among 11 Picazuro pigeons (*Patagioenas picazuro*) housed in an aviary with other bird species in the Netherlands. The birds lost weight, had ruffled feathers, and were anorexic. Despite treatment with fluids, force-feeding, and in 1 bird, doxycycline treatment (50 mg/kg 1×/d), all 11 animals died or were euthanized. Necropsy revealed that 9 of these birds were in poor physical condition, lacking fat and pectoral muscle mass. The livers and spleens were enlarged; the livers extended an average of 0.5 cm beyond the rear edge of the sternum, whereas the mean diameter of the spleens was 1.0 cm, approximately twice as large as the normal size. We suspected *Chlamydia* infection because of intracellular inclusions in Stamp (modified Ziehl Neelsen)–stained cytology of liver and spleen. We found multifocal heterophilic and lymphoplasmacytic infiltrates with necrosis in the liver and lymphoid depletion with necrosis and heterophilic infiltrates in the spleen. We stained slides with polyclonal antibodies against *Chlamydia* (bioMérieux, <https://www.biomerieux.com>) after a standard Avidin Biotin Complex protocol (4); liver and kidney tissues from 7 birds tested positive for *Chlamydia*. We did not observe any histologic changes consistent with viral inclusions or bacterial infection.

Because psittacosis in birds is a notifiable disease in the Netherlands, we informed public health authorities of our results. We forwarded frozen tissue samples to the Wageningen Bioveterinary Research institute to confirm *C. psittaci* infection. We also collected and forwarded 2 Picazuro pigeon carcasses and 3 pooled fecal samples from contact birds (i.e., Roseate spoonbill [*Platalea ajaja*], Puna ibis [*Plegadis ridgwayi*], and Scarlet ibis [*Eudocimus ruber*]), from the aviary. Two liver samples, 2 conjunctival and cloacal swabs, and 3 pooled fecal samples initially tested negative for *C. psittaci*, *C. abortus*, *C. felis*, and *C. caviae* in a PCR selective for the *ompA* gene. Because the liver and kidney samples of 7 pigeons tested positive for antibodies against *Chlamydia*, we submitted samples from all 11 pigeons and the 3 pooled fecal samples for further testing with real-time PCR selective for the 23S gene of *Chlamydiaceae* (5) and a duplex real-time PCR selective for *C. gallinacea* and *C. avium* (3,6). All 11 pigeons tested positive for *C. avium* in ≥1 samples of conjunctiva, cloaca, liver or intestines. The pooled fecal samples of contact birds tested negative in a PCR for *Chlamydiaceae* (Appendix, <https://wwwnc.cdc.gov/EID/article/26/10/20-0086-App1.pdf>).

We used Buffalo green monkey cells to isolate *Chlamydia* from the spleen of 1 of the pigeons that tested positive. Multilocus sequence typing using the concatenated sequences of 7 housekeeping genes revealed that this isolate is a unique sequence type, 254, that is closely related to the other 3 *C. avium* strains previously described (2) (Figure).

The clinical signs, histopathologic results, and positive intralésional immunohistochemistry findings

(Appendix) showed that the birds had generalized disease consistent with a *Chlamydia* infection. Real-time PCR revealed an infection with *C. avium*. Further analysis with multilocus sequence typing showed the isolated strain is unique, but most closely related to other reported *C. avium* strains. *C. avium* has been detected mainly in urban or feral pigeons without clinical signs and in co-infections of feral pigeons with *C. psittaci* (2).

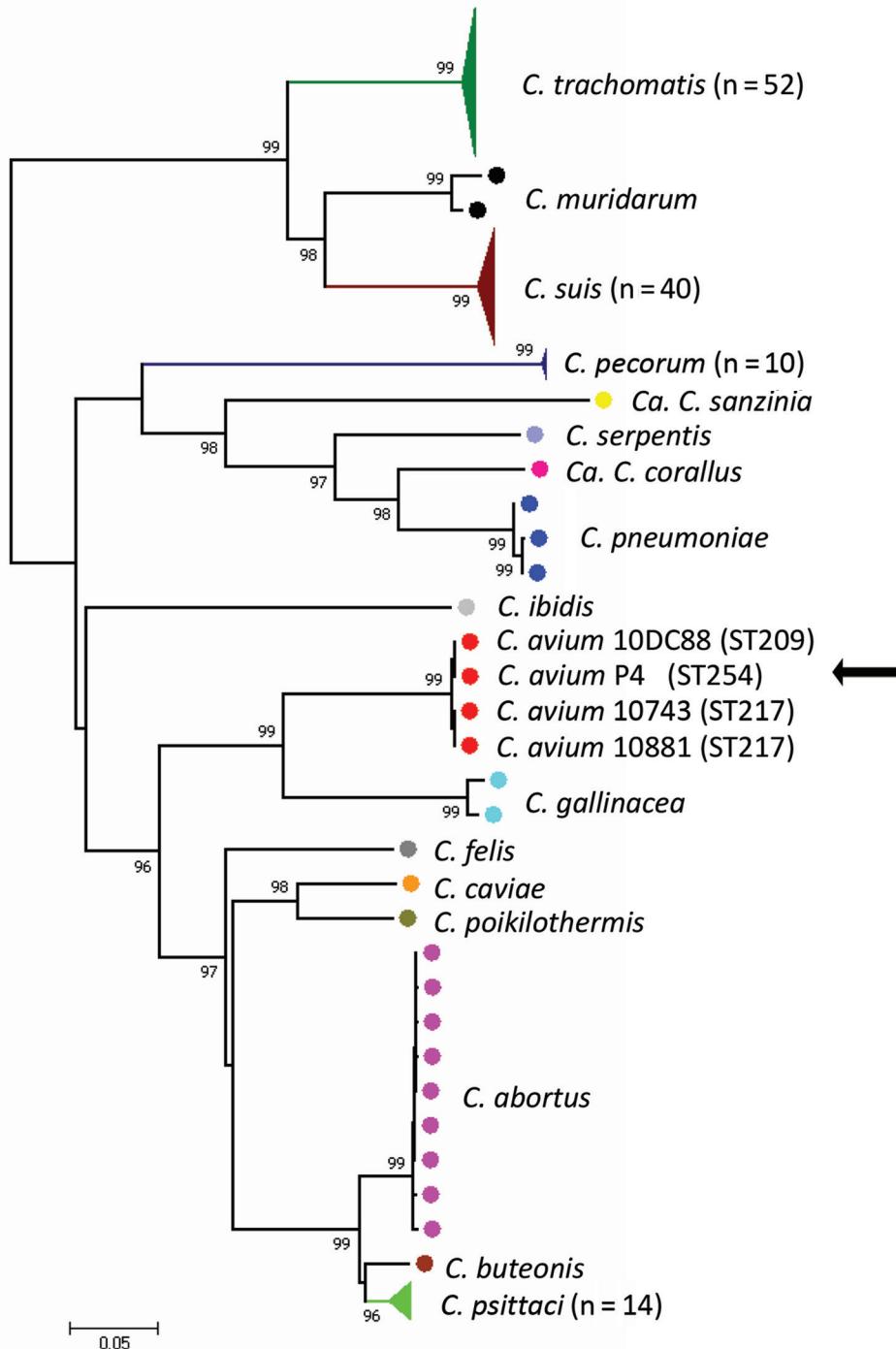


Figure. Phylogenetic analyses of concatenated sequences of 7 housekeeping gene fragments of *Chlamydiaceae*, the Netherlands, 2016. Numbers indicate bootstrap values >90%. Filled circles represent isolates, colored by species. Filled colored triangles represent >9 isolates of the same species; total number of isolates used for the analyses is indicated. The sequence types of the *C. avium* isolates are labeled. *C. avium* isolate P4 is indicated by the arrow. Scale bar indicates sequence divergence. ST, sequence type.

Our results show that *C. avium* strains might also cause severe, potentially fatal infections in birds. Data on *C. avium* are limited, but several factors might explain the severity of the clinical signs. Unlike previously reported cases, these pigeons were held in captivity. Furthermore, we cannot exclude possible differences in virulence between sequence types of *C. avium*. No human cases were reported during this outbreak, so the zoonotic potential of *C. avium* remains unknown.

Acknowledgments

We thank Rachel Thomas for proofreading. We also thank Frank Harders and Annemieke Dinkla for their technical assistance in DNA isolation and sequencing.

This work was partly funded by the Dutch Ministry of Agriculture, Nature and Food Quality (grant no. WOT-01-002-005.02).

About the Author

Dr. Kik is a veterinary pathologist at Utrecht University. Her research interests include the pathology of exotic animals and wildlife.

References

1. Kaleta EF, Taday EM. Avian host range of *Chlamydophila* spp. based on isolation, antigen detection and serology. *Avian Pathol.* 2003;32:435–62. <https://doi.org/10.1080/03079450310001593613>
2. Sachse K, Laroucau K, Riege K, Wehner S, Dilcher M, Creasy HH, et al. Evidence for the existence of two new members of the family *Chlamydiaceae* and proposal of *Chlamydia avium* sp. nov. and *Chlamydia gallinacea* sp. nov. *Syst Appl Microbiol.* 2014;37:79–88. <https://doi.org/10.1016/j.syapm.2013.12.004>
3. Sachse K, Laroucau K. Two more species of *Chlamydia*—does it make a difference? *Pathog Dis.* 2015;73:1–3. <https://doi.org/10.1093/femspd/ftu008>
4. Key M. Immunohistochemical staining methods. In: Kumar GL, Rudbeck L, editors. *Immunohistochemical staining methods*. 5th ed. Carpinteria (CA): Dako Corporation; 2009. p. 57–60.
5. Zocevic A, Vorimore F, Vicari N, Gasparini J, Jacquin L, Sachse K, et al. A real-time PCR assay for the detection of atypical strains of *Chlamydiaceae* from pigeons. *PLoS One.* 2013;8:e58741. <https://doi.org/10.1371/journal.pone.0058741>
6. Pannekoek Y, Morelli G, Kusecek B, Morré SA, Ossewaarde JM, Langerak AA, et al. Multi locus sequence typing of Chlamydiales: clonal groupings within the obligate intracellular bacteria *Chlamydia trachomatis*. *BMC Microbiol.* 2008;8:42. <https://doi.org/10.1186/1471-2180-8-42>

Address for correspondence: Marja Kik, Faculty of Veterinary Medicine, Pathobiology, Utrecht University, Yalelaan 1, 3584 CL Utrecht, the Netherlands; email: info@kikdierenarts.nl

***Streptococcus equi* Subspecies *zooepidemicus* and Sudden Deaths in Swine, Canada**

Matheus de O. Costa, Brad Lage

Author affiliations: University of Minnesota, St. Paul, Minnesota, USA (M.O. Costa); University of Saskatchewan, Saskatoon, Saskatchewan, Canada (M.O. Costa); Utrecht University, Utrecht, the Netherlands (M.O. Costa); Maple Leaf Agri-Farms, Landmark, Manitoba, Canada (B. Lage)

DOI: <https://doi.org/10.3201/eid2610.191485>

Historically described as a commensal of the swine upper respiratory tract, *Streptococcus equi* subspecies *zooepidemicus* was previously reported as an important swine pathogen only in Asia. Here we report the isolation and whole genome characterization of *S. equi* subsp. *zooepidemicus* associated with a sudden death outbreak in pigs in Canada.

Streptococcus equi subspecies *zooepidemicus* is considered a commensal and opportunistic pathogen of several warm-blooded hosts, including humans, horses, canines, and swine. It is a gram-positive, β -hemolytic coccus belonging to the Lancefield group C and can cause severe disease characterized by pneumonia, septicemia, and meningitis (1,2). *S. equi* subsp. *zooepidemicus* has been suggested as a normal inhabitant of the palatine tonsils of pigs, being detected by both culture and high-throughput sequencing in samples collected from healthy animals (3). However, strains virulent to pigs have also been reported, particularly associated with high-mortality outbreaks of sudden death and respiratory disease in China (4). No vaccines are available for this pathogen, and control and prevention methods are rarely applied because of its normally harmless commensal nature in swine. Here, we report an outbreak of sudden death associated with *S. equi* subsp. *zooepidemicus* in pigs housed in intensive commercial rearing facilities in Canada.

In April 2019, an outbreak of sudden deaths and abortions occurred in 4 loose-housed, commercial sow farms (\approx 9,000 sows) in a large vertically integrated swine system in Manitoba, Canada. This outbreak increased the cumulative death in the 3 affected sow herds by $>1,000$ animals over a 12-week period. The abortion rate during this time was $\approx 11\times$ normal.

The sows were often described as apparently healthy during morning checks. However, over the course of hours, infected sows would become