RESEARCH LETTERS

Fatal Fungicide-Associated Triazole-Resistant Aspergillus fumigatus Infection, Pennsylvania, USA

Kennedy Bradley,¹ Audrey Le-Mahajan,¹ Beth Morris, Tiina Peritz, Tom Chiller, Kaitlin Forsberg, Natalie S. Nunnally, Shawn R. Lockhart, Jeremy A.W. Gold,² Jane M. Gould²

Author affiliations: Department of Public Health, Philadelphia, Pennsylvania, USA (K. Bradley, B. Morris, T. Peritz, J.M. Gould); Hospital of the University of Pennsylvania, Philadelphia (A. Le-Mahajan); Centers for Disease Control and Prevention, Atlanta, Georgia, USA (T. Chiller, K. Forsberg, N.S. Nunnally, S.R. Lockhart, J.A.W. Gold)

DOI: https://doi.org/10.3201/eid2809.220517

We report a fatal infection in a 65-year-old immunocompromised male patient caused by pan-triazole–resistant *Aspergillus fumigatus* containing a $TR_{34}/L98H$ genetic mutation linked to agricultural fungicide use. Clinical and environmental surveillance of triazole-resistant *A. fumigatus* is needed in the United States to prevent spread and guide healthcare and agricultural practices.

A spergillus fumigatus is the most common cause of invasive aspergillosis, a life-threatening fungal infection that primarily affects immunocompromised persons, including those with hematologic malignancies or stem cell or solid organ transplants or those receiving immunosuppressive medications (1). Patients are infected by inhaling *A. fumigatus* spores found in the environment. Each year, invasive aspergillosis accounts for >14,000 hospitalizations and imposes >\$1.2 billion in direct costs on the US healthcare system (2).

Voriconazole belongs to the triazole class of antifungal drugs and is a first-line treatment for invasive aspergillosis (1). Triazole drugs have improved patient survival; however, the emergence of triazoleresistant *A. fumigatus* threatens the effectiveness of triazoles in clinical practice (3). Patients with invasive aspergillosis caused by voriconazole-resistant *A. fumigatus* had a mortality rate of ~60%, which was ~2 times the mortality rate associated with voriconazolesusceptible infection (4). Patients can acquire triazoleresistant *A. fumigatus* infections because of exposure to long-term triazole therapy for chronic aspergillosis or by directly inhaling environmental spores that are already triazole-resistant (3). The agricultural use of triazole fungicides, a practice that recently increased 4-fold in the United States, can select for *A. fumigatus* strains harboring unique *CYP51A* gene mutations, such as $TR_{34}/L98H$ and $TR_{46}/Y121F/T289A$, that can cause pan-triazole resistance in patients (5,6).

Reports of environmentally acquired triazoleresistant *A. fumigatus* infections are increasing worldwide; however, data on these infections and their clinical implications are lacking in the United States (3). We report a patient who died from an invasive infection caused by a pan-triazole-resistant *A. fumigatus* strain containing an environmentally acquired TR₄₄/L98H mutation in *CYP51A*.

The male patient was 65 years of age and previously underwent chimeric antigen receptor T-cell therapy for acute myeloid leukemia. One month before hospital admission, the patient received an allogeneic stem cell transplant that was complicated by cutaneous graftversus-host disease. Despite topical therapy, he was admitted to the hospital because of worsening rashes, fever, and lethargy. The patient received broad-spectrum antibacterial drugs and systemic corticosteroid therapy for progressive graft-versus-host disease involving the gastrointestinal tract and eyes and continued receiving transplant-related fluconazole prophylaxis.

On hospital day 3, the patient was transferred to the intensive care unit for wound management and treated for hypovolemic shock; his antifungal prophylaxis was changed from fluconazole to posaconazole. After 6 days, posaconazole was replaced with caspofungin because the posaconazole was potentially exacerbating the patient's rash. The patient improved and remained hemodynamically stable for \approx 2 weeks, after which clinicians deescalated antibacterial therapy.

On hospital day 23, acute-onset shock and hypoxemic respiratory failure developed in the patient; he was intubated and placed on mechanical ventilation. Chest computed tomography imaging showed multifocal pneumonia; bronchial cultures were positive for A. *fumigatus*. Clinicians initiated voriconazole therapy for probable invasive aspergillosis and continued caspofungin. On hospital day 27, progressive acidemia, refractory hypotension, and focal neurologic deficits developed in the patient. Rhizopus spp. was identified from the patient's skin culture, but the patient was not treated for this pathogen because his family had decided to focus on comfort care. The patient died on hospital day 28. An autopsy determined that the cause of death was sepsis from disseminated A. fu*migatus* and *Rhizopus* spp. infections.

Although most US clinical laboratories do not perform antifungal susceptibility testing, triazole

¹These first authors contributed equally to this article.

²These senior authors contributed equally to this article.

susceptibility testing for A. fumigatus isolates is available through the Centers for Disease Control and Prevention (CDC) Antibiotic Resistance Laboratory Network (https://www.cdc.gov/drugresistance/laboratories. html). Clinicians sent an isolate from the patient's bronchial washings to CDC as part of an ongoing passive surveillance for triazole-resistant A. fumigatus. Using previously described methods (7), CDC performed broth microdilution to determine the MICs of itraconazole (>16 μ g/mL) and voriconazole (2 μ g/mL) for the isolate. The isolate was classified as voriconazoleresistant in accordance with Clinical and Laboratory Standards Institute MIC breakpoints (8). The MIC of itraconazole for the isolate was considered non-wildtype on the basis of proposed epidemiologic cutoff values (9). CDC performed DNA sequence analysis of the CYP51A gene and determined that the isolate contained the TR₃₄/L98H mutation (7).

In summary, we report a fatal disseminated fungal infection in an immunocompromised patient in the United States involving pan-triazole-resistant A. fumigatus with an environmentally acquired TR₃₄/ L98H mutation. This report underscores the potential severity of triazole-resistant A. fumigatus infections in immunocompromised persons. Furthermore, clinicians should consider the possible presence of drugresistant A. fumigatus in patients with invasive aspergillosis who do not improve with first-line therapy. In Europe, the emergence of environmentally acquired triazole resistance is well documented, and voriconazole monotherapy is no longer recommended as a first-line invasive aspergillosis treatment for patients in regions with environmental resistance rates of >10% (10). In the United States, systematic clinical and environmental surveillance for triazole-resistant A. *fumigatus* is needed to determine the spread of this fungus and guide clinical and agricultural practices.

Acknowledgments

We thank Elizabeth Berkow for establishing laboratory surveillance protocols for *A fumigatus* infections; Brendan Jackson, Megan Lyman, and Mitsuru Toda for assistance with manuscript editing; laboratory staff from the Mycotic Diseases Branch, Division of Foodborne, Waterborne, and Environmental Diseases, National Center for Emerging and Zoonotic Infectious Diseases, CDC; and the Medical Examiner's Office, Philadelphia, Pennsylvania, USA, for performing the autopsy.

This activity was reviewed by CDC and conducted consistently with applicable federal laws and CDC policy (see e.g., 45 C.F.R. part 46.102(l)(2); 21 C.F.R. part 56; 42 U.S.C. §241(d); 5 U.S.C. §552a; 44 U.S.C. §3501 et seq).

About the Author

Ms. Bradley is an epidemiology fellow in the Healthcare Associated Infections and Antimicrobial Resistance Program of the Department of Public Health, Philadelphia, Pennsylvania. Her primary research interests are the epidemiology of infectious diseases and antimicrobial stewardship.

References

- Patterson TF, Thompson GR III, Denning DW, Fishman JA, Hadley S, Herbrecht R, et al. Practice guidelines for the diagnosis and management of aspergillosis: 2016 update by the Infectious Diseases Society of America. Clin Infect Dis. 2016;63:e1-60. https://doi.org/10.1093/cid/ciw326
- Benedict K, Jackson BR, Chiller T, Beer KD. Estimation of direct healthcare costs of fungal diseases in the United States. Clin Infect Dis. 2019;68:1791–7. https://doi.org/10.1093/ cid/ciy776
- Beer KD, Farnon EC, Jain S, Jamerson C, Lineberger S, Miller J, et al. Multidrug-resistant *Aspergillus fumigatus* carrying mutations linked to environmental fungicide exposure – three states, 2010–2017. MMWR Morb Mortal Wkly Rep. 2018;67:1064–7. https://doi.org/10.15585/mmwr.mm6738a5
- Lestrade PP, Bentvelsen RG, Schauwvlieghe AFAD, Schalekamp S, van der Velden WJFM, Kuiper EJ, et al. Voriconazole resistance and mortality in invasive aspergillosis: a multicenter retrospective cohort study. Clin Infect Dis. 2019;68:1463–71. https://doi.org/10.1093/cid/ciy859
- Toda M, Beer KD, Kuivila KM, Chiller TM, Jackson BR. Trends in agricultural triazole fungicide use in the United States, 1992–2016 and possible implications for antifungalresistant fungi in human disease. Environ Health Perspect. 2021;129:55001. https://doi.org/10.1289/EHP7484
- Kang SE, Sumabat LG, Melie T, Mangum B, Momany M, Brewer MT. Evidence for the agricultural origin of resistance to multiple antimicrobials in *Aspergillus fumigatus*, a fungal pathogen of humans. G3 (Bethesda). 2022;12:jkab427. https://doi.org/10.1093/g3journal/jkab427
- Berkow EL, Nunnally NS, Bandea A, Kuykendall R, Beer K, Lockhart SR. Detection of TR₃₄/L98H CYP51A mutation through passive surveillance for azole-resistant *Aspergillus fumigatus* in the United States from 2015 to 2017. Antimicrob Agents Chemother. 2018;62:e02240-17. https://doi.org/10.1128/AAC.02240-17
- 8. Clinical and Laboratory Standards Institute. Performance standards for antifungal susceptibility testing of filamentous fungi, 2nd ed. (document M61). Wayne (PA): The Institute; 2020.
- 9. Espinel-Ingroff A, Diekema DJ, Fothergill A, Johnson E, Pelaez T, Pfaller MA, et al. Wild-type MIC distributions and epidemiological cutoff values for the triazoles and six *Aspergillus* spp. for the CLSI broth microdilution method (document M38-A2). J Clin Microbiol. 2010;48:3251–7. https://doi.org/10.1128/JCM.00536-10
- Verweij PE, Ananda-Rajah M, Andes D, Arendrup MC, Brüggemann RJ, Chowdhary A, et al. International expert opinion on the management of infection caused by azoleresistant *Aspergillus fumigatus*. Drug Resist Updat. 2015;21-22:30–40. https://doi.org/10.1016/j.drup.2015.08.001

Address for correspondence: Kennedy Bradley, Department of Public Health, 1101 Market St, Philadelphia, PA 19107, USA; email: kennedy.bradley@phila.gov