#### RESEARCH LETTERS

Jpn J Infect Dis. 2017;70:92–5. https://doi.org/10.7883/ yoken.JJID.2015.626

- Kim SY, Kim MS, Chang HE, Yim JJ, Lee JH, Song SH, et al. Pulmonary infection caused by *Mycobacterium conceptionense*. Emerg Infect Dis. 2012;18:174–6. https://doi.org/10.3201/ eid1801.110251
- Thibeaut S, Levy PY, Pelletier ML, Drancourt M. *Mycobacterium conceptionense* infection after breast implant surgery, France. Emerg Infect Dis. 2010;16:1180–1. https://doi.org/10.3201/eid1607.090771
- Gonzalez-Santiago TM, Drage LA. Nontuberculous mycobacteria: skin and soft tissue infections. Dermatol Clin. 2015;33:563–77. https://doi.org/10.1016/j.det.2015.03.017
- Faria S, Joao I, Jordao L. General overview on nontuberculous mycobacteria, biofilms, and human infection. J Pathogens. 2015;2015:809014. https://doi.org/10.1155/2015/809014
- Covert TC, Rodgers MR, Reyes AL, Stelma GN Jr. Occurrence of nontuberculous mycobacteria in environmental samples. Appl Environ Microbiol. 1999;65:2492–6. https://doi.org/10.1128/AEM.65.6.2492-2496.1999
- Prevots DR, Marras TK. Epidemiology of human pulmonary infection with nontuberculous mycobacteria: a review. Clin Chest Med. 2015;36:13–34. https://doi.org/ 10.1016/j.ccm.2014.10.002

Address for correspondence: Nupur Singh, University of Tennessee Health Science Center, 910 Madison Ave, Memphis, TN, 38103, USA; email: nsingh8@uthsc.edu, nupursingh2799@gmail.com

## Acute Gastroenteritis Associated with Norovirus GII.8[P8], Thailand, 2023

## Watchaporn Chuchaona, Sompong Vongpunsawad, Weerasak Lawtongkum, Nattawan Thepnarong, Yong Poovorawan

Author affiliations: Chulalongkorn University, Bangkok, Thailand (W. Chuchaona, S. Vongpunsawad, Y. Poovorawan); Vachira Phuket Hospital, Phuket, Thailand (W. Lawtongkum, N. Thepnarong)

DOI: https://doi.org/10.3201/eid3001.231264

Acute gastroenteritis associated with human norovirus infection was reported in Phuket, Thailand, in June 2023. We amplified GII.8[P8] from the outbreak stool specimens. Retrospective sample analysis identified infrequent GII.8[P8] in the country beginning in 2018. In all, the 10 whole-genome GII.8[P8] sequences from Thailand we examined had no evidence of genotypic recombination. N orovirus is the most common cause of acute viral gastroenteritis among adults and children and has no currently approved vaccine (1). Norovirus is genetically diverse and is classified into 10 genogroups (GI-GX) representing ≈50 genotypes, of which GI and GII predominantly infect humans (2). Currently, dual-typing of the RNA-dependent RNA polymerase (RdRp) gene in the open reading frame 1 region and the major capsid protein (VP1) gene in the open reading frame 2 region is required for proper genotype assignment and detection of viral recombinants (3).

In June 2023, health officials in Thailand were investigating diarrheal outbreaks that occurred on Phuket Island in southern Thailand, which is frequented by international travelers (https://www. bangkokpost.com/thailand/general/2592541/ phukets-diarrhoea-outbreak-wanes-cause-stillunknown). Two stool specimens were eventually sent to our laboratory at the Center of Excellence in Clinical Virology at Chulalongkorn University (Bangkok) for molecular typing. The study was approved by Chulalongkorn University Institutional Review Board (approval no. 549/62). After viral RNA extraction from the stool specimens, quantitative real-time reverse transcription PCR (4) identified GII norovirus in both specimens. Confirmation assays using conventional reverse transcription PCR (5) with additional primers (Appendix 1 Table 1) and nucleotide sequencing yielded near-complete genomes, which we subjected to the norovirus genotyping tools of the Netherlands' National Institute for Public Health and the Environment (https:// www.rivm.nl/mpf/norovirus/typingtool) and the US Centers for Disease Control and Prevention (https://calicivirustypingtool.cdc.gov).

Both specimens from Phuket were human norovirus GII.8[P8]. Because GII.8[P8] is relatively uncommon and rarely linked to large outbreaks, we retrospectively examined archived stool specimens dating back to 2018 to determine the frequency of past infection in the country. We identified 8 additional GII.8 strains (Table), all of which were GII.8[P8]. We deposited these complete genome sequences in GenBank (accession nos. OR546391–OR546400).

All 10 patients who tested positive for GII.8[P8] were relatively young (age range 3–29 years, mean age 10.8 years <u>+</u> 7.1 SD). Five patients had vomiting and diarrhea, 3 had vomiting only, and 2 had diarrhea only (Appendix 1 Table 2, https://wwwnc.cdc.gov/EID/article/30/1/23-1264-App1.pdf). Minor symptoms were nausea, abdominal pain, fever, and headaches. All but 1 patient required 1–2 nights of hospital stay.

#### **RESEARCH LETTERS**

Table. Human norovirus GII.8[P8] strains identified in Thailand. 2018–2023

Collection date	Specimen ID	Patient age, y/Sex	Location	Specimen type	
2018 Feb 2	B4899	5/M	Saraburi	Stool	
2018 Feb 18	B5182	7/M	Bangkok	Stool	
2018 Sep 18	B6213	6/F	Nonthaburi	Stool	
2019 Jul 30	B6941	12/M	Nonthaburi	Stool	
2020 Feb 04	B7634	29/F	Bangkok	Stool	
2023 Feb 22	B9202	3/M	Bangkok	Stool	
2023 Feb 27	B9256	12/F	Chaiyaphum	Stool	
2023 Apr 19	B9804	10/M	Bangkok	Stool	
2023 Jun 14	B10039	12/F	Phuket	Rectal swab	
2023 Jun 13	B10069	7/F	Phuket	Stool	

Λ		D			
		D	• GII/Hu/TH/2023/GII.8/B10039		
GII/Hu/JP/2002/GII.P7/SaitamaU4 (AB039777)	II.P7		<ul> <li>GII/Hu/TH/2023/GII.8/B10069</li> </ul>		
	II.F /		<ul> <li>GII/Hu/TH/2023/GII.8/B9804</li> </ul>		
			• GII/Hu/TH/2023/GII.8/B9202		
GII/Hu/JP/2002/GII.P6/SaitamaU16 (AB039778)	II.P6	[	• GII/Hu/TH/2023/GII.8/B9256		
<sup>™L</sup> GII/Hu/CN/2011/GII.P6/GZ2010-L96 (JX989075)		[	• GII/Hu/TH/2018/GII.8/B6213		
GII/Hu/US/1999/GII.P15/Sapporo/HK299 (KJ196290)	II.P15	[	<ul> <li>GII/Hu/TH/2018/GII.8/B5182</li> </ul>		
<sup>∞L</sup> GII/Hu/SA/1990/GII.P15/DS379 (MW261797)			GII/Hu/CN/2018/GII.8/SCD1061_4 (MN996298)		
GII/Hu/JP/2002/GII.P8/SaitamaU25 (AB039780)		`	<ul> <li>GII/Hu/TH/2018/GII.8/B4899</li> </ul>	GII.8	
GII/Hu/US/2020/GII.P8/CA-RGDS-1153 (OP686904)			GII/Hu/US/2014/GII.8/CA-RGDS-1133 (MZ292794)	00	
GII/Hu/TH/2019/GII.P8/B6941			GII/Hu/CA/2018/GII.8/BMH18-086 (MW661257)		
● GII/Hu/TH/2020/GII.P8/B7634			GII/Hu/CN/2017/GII.8/GZ2017-L601 (MK213549)		
<sup>33</sup> GII/Hu/CA/2018/GII.P8/BMH18-086 (MW661257)			GII/Hu/CN/2016/GII.8/JC231 (KY406948)		
GII/Hu/CN/2017/GII.P8/GZ2017-L601 (MK213549)		92	• GII/Hu/TH/2020/GII.8/B7634		
GII/Hu/US/2014/GII.P8/CA-RGDS-1133 (MZ292794)			GII/Hu/US/2020/GII.8/CA-RGDS-1153 (OP686904)		
GII/Hu/TH/2023/GII.P8/B9256			• GII/Hu/TH/2019/GII.8/B6941		
	GII.P8	d L	GII/Hu/NL/1998/GII.8/Amsterdam (AF195848)		
<ul> <li>GII/Hu/TH/2018/GII.P8/B4899</li> <li>GII/Hu/TH/2018/GII.P8/B4899</li> </ul>			GII/Hu/JP/2002/GII.8/U25GII (AB067543)		
• GII/Hu/TH/2018/GII.P8/B5182			GII/Hu/JP/2002/GII.8/SaitamaU25 (AB039780)		
<ul> <li>GII/Hu/TH/2018/GII.P8/B6213</li> <li>GII/Hu/TH/2022/GII.P8/B6213</li> </ul>			GII/Hu/US/1999/GII.14/M7 (AY130761)	1	
<ul> <li>GII/Hu/TH/2023/GII.P8/B9202</li> <li>GII/Hu/TH/2023/GII.P8/B9804</li> </ul>			GII/Hu/JP/2013/GII.14/HO-47 (LC122869)		GII.14
			GII/Hu/US/2002/GII.17/CS-E1 (AY502009)		
<ul> <li>Image: GII/Hu/TH/2023/GII.P8/B10039</li> <li>Image: GII/Hu/TH/2023/GII.P8/B10069</li> </ul>			GII/Hu/JP/2001/GII.17/OC01031-1 (AB545151)		GII.17
GII/Hu/US/2016/GII.P12/AnnArbor-0027 (MK073886)			GII/Hu/GT/2013/GII.17/Guatemala0082-GII17 (OL94380	02)	
	GII.P12		GII/Hu/HK/2015/GII.21/CUHK-NS-601 (KR921939)	,	GII.21
Gli/Hu/JP/2005/Gli.P12/Sakai-04–179 (AB220922)	) OILF12	14	II/Hu/GB/1990/GII.7/Leeds (AJ277608)		
GII/Hu/AU/2004/GII.P4/Hunter504D/04O (DQ078814)	1		II/Hu/AU/2008/GII.7/NSW743L (GQ849130)		GII.7
• GII/Hu/US/2011/GII.P4/MI001 (KC631814)	GII.P4	l l <sub>G</sub>	II/Hu/JP/2012/GII.7/EH-58 (LC122888)		
r GII/Hu/AU/2012/GII.P31/NSW0514 (JX459908)	i		GII/Hu/GB/1993/GII.4/Bristol (X76716)	1	
GII/Hu/TW/2014/GII.P31/D001 (KY488575)	GII.P31	051	GII/Hu/HK/2019/GII.4 Hong Kong/CUHK-NS-2200 (MN	400355)	
GII/Hu/HK/2012/GII.P31/CUHK6080 (KC631827)		95 99 T	GII/Hu/TH/2016/GII.4 Hong Kong/B2717 (MW521097)		
GII/Hu/US/1971/GII.P1/Hawaii (U07611)	GII.P1	ПЦ	GII/Hu/AU/2012/GII.4 Sydney/NSW0514 (JX459908)		
GII/Hu/NP/2014/GII.P21/ETR-NV-104 (MK396773)		93	GII/Hu/TH/2020/GII.4 Sydney/B7470 (MW521128)		GII.4
<sup>*7</sup> ↓ GII/Hu/RU/2016/GII.P21/NS16-C32 (KY210919)	GII.P21	97	GII/Hu/NL/2007/GII.4 Apeldoorn/317 (AB445395)		
- GII/Hu/JP/2004/GII.P2/MK04 (DQ456824)	GII.P2	I	GII/Hu/AU/2008/GII.4 New Orleans/NSW001P (GQ845	367)	
GII/Hu/US/2016/GII.P2/Washington0526 (MK753007)	GILPZ		GII/Hu/US/2009/GII.4 New Orleans/1805 (GU445325)		
GII/Hu/DE/2000/GII.P16/Neustrelitz260 (AY772730)		<sub>™</sub> г (	GII/Hu/JP/2003/GII.22/Yuri (AB083780)		GII.22
GII/Hu/US/2018/GII.P16/Washington0207 (MK754446)	GII.P16		GII/Hu/BD/2012/GII.22/Dhaka1940 (MG495082)		011.22
GII/Hu/BT/2013/GII.P16/ETR-NV-203 (MK396777)		~	GII/Hu/US/2011/GII.1/CA-RGDS-1112 (MZ265063)		
GII/Hu/NL/1995/GII.P3/NV_Amsterdam_1994 (KJ194504)	GII.P3		GII/Hu/BR/2015/GII.1/Ananas (MT242482)		GII.1
	GII.F3		GII/Hu/US/1971/GII.1/Hawaii (U07611)		
GII/Hu/JP/2014/GII.P17/Kawasaki323 (AB983218)		-∣∥ г	GII/Hu/GB/1994/GII.2/Melksham (X81879)		
<sup>**</sup> GII/Hu/HK/2014/GII.P17/CUHK-NS-463 (KP998539)	GII.P17		GII/Hu/US/2017/GII.2/CA-RGDS-1075 (MN493873)		GII.2
<sup>99</sup> GII/Hu/JP/2015/GII.P17/Kawasaki308 (LC037415)			GII/Hu/JP/2014/GII.2/Yamaguchi-014 (LC209469)		
┌ GII/Hu/BD/2012/GII.P25/Dhaka1928 (MG495083)			GII/Hu/DE/2000/GII.10/Erfurt546 (AF427118)		GII.10
GII/Hu/CN/2021/GII.P25/N21051 (OL451532)	GII.P25		GII/Hu/US/2022/GII.10/CA-RGDS-1159 (OP691294)		
<sup>101</sup> GII/Hu/US/2022/GII.P25/CA-RGDS-1158 (OP690505)			GII/Hu/CN/2021/GII.3/N21051 (OL451532)		
GI/Hu/US/1968/GI.P1/Norwalk (M87661)			GII/Hu/CA/1991/GII.3/TV24 (U02030)		011.0
0.05		11 1	GII/Hu/NL/2014/GII.3/Groningen (LN854569)		GII.3
0.05		2] 2]	GII/Hu/US/2016/GII.3/AnnArbor-0027 (MK073886)	,, I	
		<sup>%</sup>	GII/Hu/RU/2022/GII.3/Nizhny Novgorod589 (OP901693	5)	
		11 1	GII/Hu/GB/1990/GII.6/Seacroft (AJ277620)		
		91	GII/Hu/US/2012/GII.6/BethesdaD21 (KY424344)		GIL6

Figure. Phylogenetic analysis of the complete nucleotide sequences of noroviruses identified in Thailand, 2018–2023 (black dots), and reference sequences. A) RNA-dependent RNA polymerase (RdRp) region; B) major capsid protein (VP1) region. Trees were generated using the maximum-likelihood method based on the general time reversible model, with 1,000 bootstrap replications for branch support as implemented in MEGA software version 11 (http://www.megasoftware.net). Bootstrap values >80 are indicated at the branch nodes. GenBank accession numbers for reference sequences are provided in parentheses. Scale bar indicates nucleotide substitutions per site.

GII/Hu/JP/2021/GII.6/178F (LC726059) GII/Hu/CN/2016/GII.6/016Q02 (KY407214) - GI/Hu/US/1968/GI.1/Norwalk (M87661)

GII.6

0.20

From the complete nucleotide sequences of the RdRp and VP1 genes, the GII.8[P8] strains from Thailand phylogenetically clustered with strains identified in Canada (GenBank accession no. MW661257), China (GenBank accession nos. MK213549 and MN996298), and the United States (GenBank accession nos. MZ292794 and OP686904) during the previous 10 years (Figure). Collectively, nucleotide sequence identities of GII.8[P8] strains from Thailand and other strains were 85%-99% over the entire genome compared with the prototypic GII.8[P8] SaitamaU25 (GenBank accession no. AB039780) (Appendix 1 Figure). However, Phuket GII.8[P8] appeared to diverge most from other GII.8[P8] strains in parts of the nonstructural protein 1-2 (p48), nonstructural protein 3 (NTPase), and VP1 shell domain.

To address whether Phuket GII.8[P8] strains had developed notable amino acid changes on its genome, we compared their deduced residues to other GII.8[P8] strains. Phuket GII.8[P8] shared many unique residue changes with the most recent strain from Thailand (B9804) identified in Bangkok 2 months prior (Appendix 2 Table, https://wwwnc.cdc.gov/EID/ article/30/1/23-1264-App2.xlsx). No apparent mutations to suggest increased virulence or viral transmissibility were obvious, although ≥10 residue positions scattered throughout the GII.8[P8] genome identified in Thailand in 2023 were not shared by other known GII.8[P8] sequences. Most residue variations were conservative changes; however, T479S on VP1 is a highly conserved position among GII noroviruses.

The potential for GII.8[P8] to cause the recent norovirus outbreak in Phuket was unexpected given that the last reported outbreak in Thailand wad caused by a novel GII.3[P25] recombinant in Chanthaburi Province (6). Of note, GII.8[P8] outbreaks are infrequent (7), and the most recent occurrence was foodborne (through contaminated raspberries) (8). No specific food source was identified and laboratory-confirmed for norovirus, and anecdotal evidence suggests probable person-to-person norovirus transmission in the Phuket outbreak. Reports of GII.8[P8] infection in the literature have not identified RpRp–VP1 recombinants, and comprehensive historical analysis of norovirus sequences suggests that GII.8 RdRp and VP1 rarely recombine with other genotypes (9).

Molecular analysis in this study was limited because <40 complete GII.8[P8] genomes were available in the public database. This study was also constrained by the scarcity of specimens sent for laboratory testing, which underscored limited awareness and importance placed by health officials toward timely etiologic diagnosis. A study suggests that antibodies elicited by GI.1 and GII.4 (2 genotypes in the norovirus vaccine candidate under consideration) minimally block the binding of GII.8 VLPs to histoblood group antigens (*10*). Although unlikely, any potential increase in the prevalence of GII.8[P8] could affect real-world norovirus vaccine effectiveness. In summary, GII.8[P8] genomes identified in this study are expected to contribute to the ongoing molecular and epidemiologic surveillance of community-acquired norovirus infection, which could benefit the tracking of global norovirus transmission.

This study was supported by the Center of Excellence in Clinical Virology of Chulalongkorn University and Hospital. Support for W.C. was provided by the Second Century Fund of Chulalongkorn University.

### About the Author

Dr. Chuchaona is a postdoctoral fellow at the Center of Excellence in Clinical Virology in the Faculty of Medicine at Chulalongkorn University. Her primary research interests are molecular epidemiology and evolution of human noroviruses.

### References

- Ahmed SM, Hall AJ, Robinson AE, Verhoef L, Premkumar P, Parashar UD, et al. Global prevalence of norovirus in cases of gastroenteritis: a systematic review and meta-analysis. Lancet Infect Dis. 2014;14:725–30. https://doi.org/10.1016/ S1473-3099(14)70767-4
- Chhabra P, de Graaf M, Parra GI, Chan MC, Green K, Martella V, et al. Updated classification of norovirus genogroups and genotypes. J Gen Virol. 2019;100:1393–406. https://doi.org/10.1099/jgv.0.001318
- Kroneman A, Vega E, Vennema H, Vinjé J, White PA, Hansman G, et al. Proposal for a unified norovirus nomenclature and genotyping. Arch Virol. 2013;158:2059–68. https://doi.org/10.1007/s00705-013-1708-5
- Debbink K, Costantini V, Swanstrom J, Agnihothram S, Vinjé J, Baric R, et al. Human norovirus detection and production, quantification, and storage of virus-like particles. Curr Protoc Microbiol. 2013;31:15K.1.1–15K.1.45.
- Chhabra P, Browne H, Huynh T, Diez-Valcarce M, Barclay L, Kosek MN, et al. Single-step RT-PCR assay for dual genotyping of GI and GII norovirus strains. J Clin Virol. 2021;134:104689. https://doi.org/10.1016/j.jcv.2020.104689
- Chuchaona W, Khongwichit S, Luang-On W, Vongpunsawad S, Poovorawan Y. Norovirus GII.3[P25] in patients and produce, Chanthaburi Province, Thailand, 2022. Emerg Infect Dis. 2023;29:1067–70. https://doi.org/10.3201/ eid2905.221291
- Eftekhari M, Kachooei A, Jalilvand S, Latifi T, Habib Z, Ataei-Pirkoohi A, et al. The predominance of recombinant norovirus GII.4Sydney[P16] strains in children less than 5 years of age with acute gastroenteritis in Tehran, Iran, 2021– 2022. Virus Res. 2023;334:199172. https://doi.org/10.1016/ j.virusres.2023.199172
- 8. Lysén M, Thorhagen M, Brytting M, Hjertqvist M, Andersson Y, Hedlund KO. Genetic diversity among food-borne and

waterborne norovirus strains causing outbreaks in Sweden. J Clin Microbiol. 2009;47:2411–8. https://doi.org/10.1128/ JCM.02168-08

- Kendra JA, Tohma K, Parra GI. Global and regional circulation trends of norovirus genotypes and recombinants, 1995–2019: a comprehensive review of sequences from public databases. Rev Med Virol. 2022;32:e2354. https://doi.org/10.1002/rmv.2354
- Gao J, Xue L, Liang Y, Wang L, He F, Meng L, et al. Receptor profile and immunogenicity of the non-epidemic norovirus GII.8 variant. Virus Res. 2021;306:198603. https://doi.org/10.1016/j.virusres.2021.198603

Address for correspondence: Yong Poovorawan, Center of Excellence in Clinical Virology, Faculty of Medicine, Chulalongkorn University, 1873 Rama 4 Rd, Pathumwan, Bangkok 10330, Thailand; e-mail: yong.p@chula.ac.th

# Use of Doxycycline to Prevent Sexually Transmitted Infections According to Provider Characteristics

William S. Pearson, Brian Emerson, Matthew Hogben, Lindley Barbee

Author affiliation: Centers for Disease Control and Prevention, Atlanta, Georgia, USA

DOI: https://doi.org/10.3201/eid3001.231152

Use of doxycycline to prevent sexually transmitted infections (STIs) may lead to antimicrobial resistance. We analyzed attitudes toward this practice between US providers who commonly and less commonly treat STIs. Providers who more commonly treat STIs are more likely to prescribe prophylactic doxycycline and believe that benefits outweigh potential for increased antimicrobial resistance.

**R**(STIs) (e.g., chlamydia, gonorrhea, and syphilis) in the United States are at the highest level in several decades (1). A useful tool for preventing STIs may be prophylactic use of doxycycline taken within 72 hours after a sexual encounter (2–5). However, concerns about development of antimicrobial resistance (AMR) (e.g., in *Neisseria gonorrhea*, which is listed by the Centers for Disease Control and Prevention as an urgent AMR threat), may affect provider attitudes toward prophylactic use of doxycycline (6). To determine differences in the practices and beliefs of providers who work with STI patients (STI providers) and do not work with STI patients (non–STI providers) with regard to prophylactic use of doxycycline for STIs and their concerns about potential AMR consequences, we analyzed survey responses.

We analyzed data from the DocStyles panel survey (https://styles.porternovelli.com/docstyles) conducted by SERMO, a social network platform for physicians (https://www.sermo.com) in conjunction with Porter Novelli during September 9–November 3, 2022. Of 1,755 US healthcare providers who responded (response rate 67.0%), we focused on a sample of 1,504 healthcare providers, including family physicians (457, 30.4%), internists (545, 36.2%), obstetrician/gynecologists (251, 16.7%), and nurse practitioners/physician assistants (251, 16.7%). We excluded 251 pediatricians.

We further stratified analyses by the percentage of the providers' practice focused on clinical management of STIs. Providers were asked, "What proportion of your visits include screening for, diagnosing, or treating sexually transmitted infections?"; the 5 possible responses were "none," "some, but less than 10%," "more than 10% up to 25%," "more than 25% up to 50%," or "more than 50%." The 743 respondents whose practice consisted of <10% STI management were considered non-STI providers, and the 761 others were considered STI providers. We further ascertained provider age, sex, specialty, and number of years in practice.

We asked 4 questions about use and beliefs with regard to doxycycline prophylaxis and antimicrobial resistance (Figure), and the 5 response choices were "strongly disagree," "somewhat disagree," "neither agree nor disagree," "somewhat agree," or "strongly agree." We used  $\chi^2$  tests to compare the percentage of respondents who chose "strongly agree," and "agree" between STI providers and non–STI providers. We further tested those differences by using adjusted logistic regression models controlling for provider age, sex, number of years in practice, and specialty (Table).

Among STI providers, 41.9% said that they had ever prescribed doxycycline for STI prophylaxis, compared with 21.0% non–STI providers (p<0.01). Among STI providers, 57.4% either strongly agreed or agreed with the statement, "I have seen an increase in antibiotic resistant infections among my patients over the past 5 years," compared with 57.6% of non– STI providers (p = 0.94). Among STI providers, 63.5% either strongly agreed or agreed with the statement,